Monitoring Report for CSMRI Site Second Quarter 2010

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August 2010

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1. Introduction

This report presents the second quarter (April, May, June) 2010 results for groundwater and surface water monitoring conducted at the Colorado School of Mines Research Institute (CSMRI) site in Golden, Colorado. The monitoring was conducted by the S.M. Stoller Corporation (Stoller).

2. Sampling and Analysis

Stoller obtained quarterly samples of groundwater and surface water on June 7, 8, 9, and 10, 2010, from 11 groundwater monitor wells and three Clear Creek surface water sample locations. Groundwater quality samples were obtained on June 7 (CSMRI-7B); June 8 (CSMRI-1B, CSMRI-4, CSMRI-5, CSMRI-6C, CSMRI-8, and CSMRI-11B); June 9 (CSMRI-1B, CSMRI-9, and CSMRI-10); and June 10 (CSMRI-1 and CSMRI-7B). Monitor wells CSMRI-1B and CSMRI-7B required purging on one day and sample collection on subsequent visits over the following days to obtain sufficient sample volume.

Clear Creek surface water samples were collected on June 9, 2010, from sampling locations SW-1 and SW-2. A third Clear Creek surface water sample, SW-3, was collected on June 10, 2010 at the request of the Colorado Department of Public Health and Environment (CDPHE). This new surface water sample location is shown on Figure 1.

All aqueous samples were placed on ice in coolers and couriered to ALS Laboratory Group in Fort Collins, Colorado or to TestAmerica, Inc. in Arvada, Colorado for analyses.

Figure 1 presents the monitor well and Clear Creek surface water sample locations at the CSMRI site. The figure also shows the groundwater potentiometric surface elevations posted adjacent to each monitor well location. Potentiometric surface elevations are based on depth to groundwater relative to the surveyed top-of-casing and represent groundwater elevations as measured before purging. Groundwater levels are measured at each well to the nearest 1/100th of a foot (0.01) prior to purging and sample collection. Depth to groundwater was measured in monitor wells on the bench terrace on June 7, 2010, and monitor wells and test pit piezometers in the flood plain on June 17, 2010. Figure 1 shows a northeasterly component of flow for groundwater on the bench terrace above the Clear Creek flood plain. This is consistent with historical groundwater flow direction for this area.

Figure 2 presents hydrographs of groundwater potentiometric elevations for monitor wells CSMRI-1, CSMRI-1B, CSMRI-4, CSMRI-5, CSMRI-6B (abandoned July 2008), CSMRI-6C, CSMRI-7B, CSMRI-8, CSMRI-9, CSMRI-10, CSMRI-11 (abandoned July 2008), and CSMRI-11B. Gaps in the graph denote the intermittent presence of groundwater in the monitor wells because sometimes groundwater is below the bottom depth of a monitor well. Monitor wells included on Figure 2 are located within the CSMRI site proper and illustrate historical trends in the water table fluctuations throughout the site.

Figure 3 is a hydrograph of monitor well CSMRI-2. Monitor well CSMRI-2 is located near the southeast corner of the freshman parking lot on West Campus Drive and the former Welch Ditch. CSMRI-2 is upgradient of the CSMRI site and historically has been used to provide background

groundwater quality conditions. Early potentiometric data reflect the use of and leakage from the nearby irrigation ditch. For example, this hydrograph shows a marked seasonal rise in the potentiometric surface during the summer months in 2005 and 2006. In 2007, the Welch Ditch was diverted upstream of CSMRI and piped to Washington Avenue in Golden and then to down-ditch users. From late 2006 through late 2007, the water level remained elevated. From early 2008 through mid-2010, water levels reflect a more typical seasonal pattern of fluctuation.

2.1 Groundwater Sampling

Water quality samples were collected following the procedure outlined in Appendix A, Groundwater Sampling Procedures.

Sample collection forms provide a record of water quality parameters as measured in the field as groundwater was purged from monitor wells. These forms also indicate the volume of water removed from each well. Sample collection forms are provided as Appendix B. After three casing volumes of groundwater were purged, water samples were filtered through a 0.45 micron (μ) filter, collected in laboratory-provided containers, and preserved in the field as appropriate for the analyte and analytical method. Monitor wells CSMRI-1B, CSMRI-2, and CSMRI-7B were purged dry before three casing volumes of groundwater were removed. Monitor wells CSMRI-1B and CSMRI-7B required multiple visits to collect sufficient sample volume because they recharge so slowly.

2.2 Surface Water Sampling

Surface water samples from Clear Creek were collected on June 9, 2010 from two locations: one upstream of the site (SW-1) and one downstream of the site (SW-2) (Figure 1). Clear Creek surface water sample SW-3 was collected at a location between the upstream and downstream sample locations and due north of monitor well CSMRI-8. Surface water at this location was sampled on June 10, 2010. All surface water samples were collected following the procedure outlined in Appendix C, Surface Water Sampling Procedures. Surface water samples are filtered through a 0.45µ filter, collected in laboratory-provided containers, and preserved in the field as appropriate for the analyte and analytical method.

Discharge data of stream flow for Clear Creek, as measured by the U.S. Geological Survey (USGS), at Clear Creek gauging station #06719505 (USGS Surface Water Online Database) for the quarter from April 1, 2010 through June 30, 2010 are presented as Figure 4. The figure shows ice affected the stream flow measurement devices from April 1, 2010 through April 5, 2010 and USGS data indicate that sampling of the CSMRI monitor wells was conducted during the peak stream flow. Tabulated stream flow data for the time period of June 8, 9, and 10, 2010 indicates the mean stream flow was recorded at 1,160, 1,120 and 1,060 cubic feet per second, respectively.

2.3 Analyses

All samples collected were analyzed using a Comprehensive Environmental Response, Compensation and Liability Act (CERCLA)-certified analytical laboratory. The results received from the laboratory were evaluated based on the following parameters:

• Data completeness

- Holding times and preservation
- Instrument initial calibrations
- Instrument performance checks
- Preparation blanks
- Duplicate sample results
- Laboratory control sample results
- Compound quantization and reporting limits (full validation only)

As a quality control/quality assurance (QA/QC) check, an equipment blank sample was collected in the field by pouring distilled water through a sample bailer. The equipment blank sample was submitted for the identical analytical parameters as the groundwater and surface water samples. The results of the equipment blank analyses did not identify interferences or anomalies in the laboratory data.

Results of the QA/QC review did identify several issues regarding analytical laboratory results. The Ra-228 analytical results from monitor wells CSMRI-1B and CSMRI-7B are rejected [**R**] due to the combination of the detector failure, the batch laboratory control sample failure, and the inductively coupled plasma-atomic emission spectroscopy barium carrier concentration being less than the known amount added.

One sample cooler arrived at the analytical laboratory at a temperature of 5° Celsius. The samples in this cooler were not identified on a separate chain-of-custody; therefore, determining which samples may have been affected by the slightly elevated temperature cannot be segregated to assess usability of the data.

Data validation results are presented in Appendix D.

2.3.1 Groundwater Quality Analyses

Summaries of groundwater results for radioisotopes, metals, and inorganic anions and cations are presented in Table 2-1, Table 2-2, and Table 2-3, respectively. Groundwater parameters are reported as picoCuries per liter (pCi/L) for radioisotopes, micrograms per liter (μ g/L) for uranium and iron, and milligrams per liter (mg/L) for all other metals and ions.

ALS Laboratory Group in Fort Collins, Colorado and TestAmerica Laboratories, Inc. in Arvada, Colorado conducted laboratory analyses of the aqueous samples. Analytical samples submitted to ALS were analyzed for radium isotopes (Ra-226 and Ra-228), uranium (U), calcium (Ca), potassium (K), magnesium (Mg), sodium (Na), phosphorus (P), chloride (Cl), sulfate (SO₄), carbonate as calcium carbonate (CO₃), bicarbonate as calcium carbonate (HCO₃), alkalinity, and dissolved organic carbon (DOC). Pursuant to an agreement with CDPHE, a broader analyte list of metals is sampled only during the second quarter (June) sampling event. Specifically these metals include: silver (Ag), arsenic (As), barium (Ba), cadmium (Cd), chromium (Cr), mercury (Hg), and vanadium (V).

Analytical samples submitted to TestAmerica were tested for the presence of nitrate (NO₃), nitrite (NO₂), ferrous (Fe²) iron, ferric (Fe³) iron, sulfide and total dissolved solids (TDS),

TestAmerica conducts the short holding time analyses because of their close proximity to the CSMRI site.

Groundwater samples were measured onsite for temperature, pH, specific conductance, dissolved oxygen (DO), oxidation-reduction potential (ORP), and turbidity as nephlometer turbidity units (NTU) during the purging and sampling process using a Horiba U-22 multi-probe. Onsite parameters measurements of groundwater and purge volumes are presented on the sample collection forms in Appendix B.

From the first quarter of monitoring in February 2005 until the last quarter of 2006, the concentration of total uranium in μ g/L was analyzed using ALS (formerly Paragon) method 714R9. With this method, the concentration of uranium is calculated based on the activity of the uranium isotopes U-234, U-235, and U-238. Effective since the first quarter 2007 sampling event, the concentration of uranium has been analyzed using mass spectrometry method EPA 6020, which analyzes for total uranium and not for the activity of the individual isotopes.

Analytical data from ALS and TestAmerica were transmitted as an electronic data deliverable (EDD) and are included in Appendix E on a compact disk. Data are formatted as series of Excel spreadsheets. Appendix F presents copies of the chain-of-custody for the CSMRI samples.

2.3.2 Surface Water Analyses

Clear Creek surface water results for radioisotopes, metals, and inorganic anions and cations are presented in Table 2-4, Table 2-5, and Table 2-6, respectively. Surface water parameters are reported as pCi/L for radioisotopes, μ g/L for uranium and iron, and mg/L for all other metals and ions. Surface water samples were measured onsite for temperature, pH, specific conductance, DO, ORP, and NTU as the sampling was conducted. Onsite parameter measurements are presented on the sample collection forms in Appendix B.

2.4 Health and Safety Program

Stoller implements a program to protect the health and safety of field personnel for during the environmental monitoring at the CSMRI site. This program has been developed in accordance with requirements of 29 Code of Federal Regulations (CFR) 1910.120.

3. Results

Groundwater analytical results for samples collected from the CSMRI site during the second quarter 2010 for radioisotopes, metals, and anions and cations are summarized on Table 2-1, Table 2-2, and Table 2-3, respectively. Surface water analytical results for samples collected from the CSMRI site during the second quarter 2010 for radioisotopes, metals, and anions and cations are summarized on Table 2-4, Table 2-5, and Table 2-6, respectively. Table 2-7 presents historical data collected by previous consultants for select contaminants of potential concern in groundwater at the site. The historical uranium data presented in Table 2-7 are presented in pCi/L as "activity," more recent (2005 through 2010) analytical data are presented in µg/L as "mass concentration." The December 7, 2000, *Federal Register* discusses the final uranium maximum contaminant level (MCL) and presents a conversion factor of a geometric average mass: activity ratio of 0.9 pCi/g for values near the National Primary Drinking Water Standards MCL, based on data from the National Inorganics and Radionuclides Survey.

Tables G-1 and G-2 in Appendix G present the quarterly historical groundwater radioisotopic and metals sample results, respectively, collected by Stoller since February 2005. Tables G-3 and G-4 in Appendix G present the quarterly historical Clear Creek surface water radioisotopic and metals sample results, respectively, collected by Stoller since February 2005.

3.1 Groundwater Conditions

Groundwater monitor wells are located in areas likely to detect impacts, if any, to groundwater emanating from the site and at locations that represent background water quality. Monitor wells CSMRI-4 and CSMRI-5 are downgradient of the site in the Clear Creek flood plain. Well CSMRI-1 is located along Clear Creek upstream of the site, and well CSMRI-2 is located offsite in the southeast corner of the freshman parking lot on West Campus Drive. Both monitor wells CSMRI-1 and CSMRI-2 are upgradient of the site.

In February 2007, seven new groundwater monitor wells were installed to assess the effectiveness of the source removal excavation that was conducted in 2006. Monitor well CSMRI-8 is located along Clear Creek within the flood plain area; and monitor wells CSMRI-1B, CSMRI-6B, CSMRI-7B, CSMRI-9, CSMRI-10, and CSMRI-11 are located on the bench terrace and essentially encircle the CSMRI site.

In July 2008, two monitor wells (CSMRI-6B and CSMRI-11) were abandoned due to construction activities at the CSMRI site. These two wells were replaced in December 2008 with CSMRI-6C and CSMRI-11B, respectively.

3.2 Groundwater Quality

Groundwater samples were collected from 11 monitor wells and tested for the presence of metals and radioisotopes as identified in Section 2.3.1. The sample volumes collected from monitor well CSMRI-7B was limited due to insufficient water. Because of the limited water volume at this well, analytical testing was only conducted for radioisotopes and uranium.

Uranium was detected in monitor wells CSMRI-4 at 56 μ g/L, CSMRI-7B at 84 μ g/L, CSMRI-8 at 540 μ g/L, and CSMRI-9 at 48 μ g/L at concentrations exceeding the State of Colorado groundwater standard of 30 μ g/L. Uranium was also detected in the remaining seven groundwater monitor wells but at concentrations below the groundwater standard.

In the flood plain area, uranium was detected in monitor wells CSMRI-4 at 56 μ g/L, CSMRI-5 at 8.7 μ g/L, and CSMRI-8 at 540 μ g/L. Monitor well CSMRI-4 historically has had elevated concentrations of uranium. Values had been declining since 1991 until the last several quarterly sampling events as depicted on Figure 5. Historically, the concentration of uranium in this monitor well spiked once in 1999 and again in 2003. The spike in the uranium concentration in 2003 was attributed to precipitation effects and removal of asphalt and concrete as discussed in New Horizons RI/FS, Section 4.2.2 of the (New Horizons 2004). The recent (2009) rise in the uranium concentration in this monitor well appears to be attributed to stormwater discharge from the new Colorado School of Mines (CSM) artificial turf soccer field subdrains. Precipitation collected in the subdrains, discharged near the northern edge of the bench terrace, and flowed down a riprap-embedded concrete rundown onto the flood plain. During the process, the discharge water became oxygenated and was introduced into the poorly oxygenized environment

of the flood plain area. Uranium present in the saturated sediments of the flood plain was mobilized by the oxygenated discharge water and flowed toward monitor well CSMRI-4. In early 2010, the soccer field discharge pipe was relocated to the east. Analytical data suggest discharge is no longer affecting this monitor well.

Figure 6 presents the potentiometric surface elevation of groundwater in CSMRI-4 (left Y axis) and the uranium concentration (right Y axis) from 2005 through the second quarter 2010. The figure indicates the uranium concentration had previously been fluctuating seasonally from slightly above to slightly below the groundwater standard of 30 μ g/L for seven quarterly sampling events in 2005 and 2006. An ice chest from the fourth quarter 2006 (December) sampling event was lost by the courier service resulting in a gap in the analytical data for CSMRI-4. The concentration of uranium in this well has increased since the 2006 surface soil remediation activities and also spiked during the third quarter 2009 sampling event. The uranium concentration in this monitor well has decreased significantly since the third quarter 2009 sampling event and reflects the relocation of the soccer field discharge pipe.

The uranium concentration detected in CSMRI-8 decreased to 540 μ g/L from the previous quarterly value of 960 μ g/L. Monitor well CSMRI-8 was installed in February 2007. Figure 7 presents the potentiometric surface elevation of groundwater at this well (left Y axis) and the uranium concentration (right Y axis) from 2007 through the second quarter 2010. Quarterly monitoring will continue at this location to document seasonal variability and trends in the uranium concentration in groundwater.

The uranium concentration in CSMRI-9 of 48 μ g/L is consistent with the past four sampling events, but it is still slightly elevated relative to the 2007 and 2008 analytical data. This monitor well is located at the top of the bench terrace that rises above the flood plain and is downgradient of the CSMRI site.

3.2.1 Ionic Balance Evaluation

The ionic testing and balancing is conducted to determine the different groundwater hydrochemical facies within the CSMRI site and to assess the analytical laboratory quality control procedures since the sum of the major anions should equal the sum of the major cations when the ionic concentrations are converted to millequivalents per liter. Significant differences in the ionic balance of a groundwater sample suggest either an ionic species is not tested for or laboratory analytical procedures are not consistent.

Groundwater and surface water samples were collected and tested for major anions and cations, DOC, and at select locations (CSMRI-1, CSMRI-4, CSMRI-5, and CSMRI-8) ferric/ferrous iron. The presence of sulfide was also analyzed for in samples from the three flood plain monitor wells (CSMRI-4, CSMRI-5, and CSMRI-8). Analytical results for these parameters are presented in Table 2-3 for groundwater and Table 2-6 for surface water. AqQA[®] geochemical software is used to calculate ionic balances of water samples and to present the graphical representation of anions and cations. Ionic balance calculations for the anions and cations for the water samples generally range from 6 to 9 percent. Only monitor wells CSMRI-9 at 10.6 percent and CSMRI-2 at 16.7 percent are greater than a 10 percent balance difference.

Summary sheets from the AqQA® geochemical software for each of the water samples are presented in Appendix H. Dominant water types identified at the CSMRI site include Ca-Cl (CSMRI-1, CSMRI-1B, CSMRI-5, CSMRI-6C, CSMRI-9, CSMRI-10, and CSMRI-11B); Ca-HCO₃ (CSMRI-2, CSMRI-8, and SW-2); and Ca-SO₄ (CSMRI-4, SW-1, and SW-3).

Historically, surface water samples SW-1 and SW-2 have always been a Ca-SO4 type waters. However, with the second quarter 2010 sampling event, SW-2 appears to be a Ca-HCO3 type water while SW-1 and new sample location SW-3 are Ca-SO4 type waters, even though the ion concentrations in all three samples are very close.

A Piper quadrilateral diagram is included in Appendix H and presents the overall ionic properties for each water sample. The cation triangle in the lower left of the figure indicates the cation composition of the water samples is similar as exhibited by the tight grouping of the plotted results. The anion triangle in the lower right of the figure indicates the anion composition of the water samples is more diverse as exhibited by the dispersed nature of the plotted results.

3.2.2 Comparison of Upgradient and Downgradient Groundwater Quality

Monitor wells CSMRI-4, CSMRI-5, and CSMRI-8 are downgradient from the upper terrace portion of the site and are located on the Clear Creek flood plain. Monitor wells CSMRI-7B and CSMRI-9 are located downgradient of the CSMRI site at the top of the bench terrace above the flood plain. Monitor wells CSMRI-10 and CSMRI-11B are located at the eastern edge of the site, and monitor wells CSMRI-1B and CSMRI-6C are located upgradient of the site.

Uranium was detected in monitor wells CSMRI-4 at concentrations of 56 μ g /L, CSMRI-8 at 540 μ g/L, and CSMRI-9 at 48 μ g/L, exceeding the groundwater standard of 30 μ g/L. Monitor well CSMRI-8 is located at the western edge of the flood plain area and is upgradient of monitor wells CSMRI-4 and CSMRI-5. Monitor well CSMRI-9 is located at the downgradient position on the bench terrace at the CSMRI site.

3.2.3 Comparison with Previous Groundwater Quality Analyses

Table 2-7 presents historical groundwater analytical results from past sampling events dating back to 1991 for radioisotopes of concern. The data indicate fluctuating concentrations of tested analytes, particularly for monitor well CSMRI-4.

As additional data are collected for each sampling quarter and trends become more defined, graphs of concentration versus time are produced and presented. These analytical data are incorporated to show long-term trends and correlation between the detected concentration of uranium in groundwater, the fluctuating water table, and seasonal variability if present.

3.2.4 Comparison with Colorado Groundwater Standards

As stated previously, the groundwater standard of 30 μ g/L for uranium in groundwater was exceeded in monitor wells CSMRI-4 (56 μ g/L), CSMRI-7B (84 μ g/l), CSMRI-8 (540 μ g/L), and CSMRI-9 (48 μ g/L). In January 2008, the CDPHE Water Quality Control Commission adopted the surface water quality standard of 30 μ g/L as the groundwater quality standard in an effort to keep both uranium standards consistent.

The historic spikes in the concentration of uranium in CSMRI-4, as shown on Figure 5, have been attributed to increased precipitation from December 2006 through February 2007 and surface soil remediation activities in the flood plain. Recent analytical results indicate a decreasing trend back to 2007 and 2008 levels prior to impacts from the soccer field storm drain.

No exceedances of the MCL for Ra-226 and Ra-228 were detected in any of the groundwater samples for this sampling quarter.

Monitoring well CSMRI-8 will be temporarily abandoned in late-2010 to accommodate soil characterization and source removal activities in the flood plain. After soil characterization is complete, this monitor well will be replaced and integrated into the quarterly sampling schedule. Sampling of this new well and other site wells will continue and the reason(s) for observed elevated levels of uranium will be evaluated.

3.3 Surface Water Quality

Surface water samples are collected from three locations at the site. Location SW-1 is located upstream from the site, SW-2 is downstream from the site, and SW-3 is located adjacent to the Clear Creek bank in the vicinity of monitor well CSMRI-8. All surface-water concentrations of tested parameters detected at the CSMRI site from stations SW-1, SW-2, and SW-3 are similar. Established water quality standards were not exceeded at any location.

4. Future Activities

During the first week of June 2010, a characterization study of subsurface soils, bedrock, and groundwater within the flood plain was conducted. Eight test pits were excavated down to bedrock and samples of the unsaturated soil, saturated soil, and bedrock were collected and submitted for analytical testing. Grab groundwater samples were collected from six of the test pits and submitted for analytical testing. Small-scale pump tests were conducted on monitor wells CSMRI-4, CSMRI-5, and CSMRI-8 to determine aquifer properties associated with alluvial deposits within the flood plain.

A geochemical modeling firm was retained to conduct testing to determine a partitioning coefficient for uranium in soil in the flood plain. This value was used to determine a soil screening level, the level at which soil has the potential to impact groundwater to levels above the MCL.

A copy of the *Preliminary Flood Plain Characterization* was presented as Appendix A in the Flood Plain Work Plan that was presented to CDPHE on August 9, 2010.

5. References

Colorado Department of Public Health and Environment, Water Quality Control Commission, Regulation No. 41, *The Basic Standards for Ground Water*. Amended: January 14, 2008, Effective: May 31, 2008.

ERO Resources, Inc. 2010. *Biological Assessment, CSMRI Flood Plain Site in Golden, Jefferson County, Colorado.* ERO Project #4633. February 9, 2010.

New Horizons Environmental Consultants, Inc. Remedial Investigation/Feasibility Study and Proposed Plan, Colorado School of Mines Research Institute Site, Golden, CO, January 21, 2004.

Stoller 2010. Draft Work Plan, Environmental Assessment and Characterization Flood Plain Area, Colorado School of Mines Research Institute Site, Golden, Colorado, Prepared by The S.M. Stoller Corporation For Colorado School of Mines, January 8, 2010.

USGS Surface Water website: http://nwis.waterdata.usgs.gov

Summary of Radioisotopes in Groundwater													
			226 Ci/L)	Ra- (p0	Total U (µg/L)								
Sample Station	Sample Date	Result	Uncertainty	Result	Uncertainty	Result							
CSMRI-1	6/10/10	0.1	0.25	0.93	0.41	2.4							
CSMRI-1B	6/9/10	0.23	0.19	-0.03 R	0.33	18							
CSMRI-2	6/10/10	0.27	0.24	2.25	0.76	1.6							
CSMRI-4	6/8/10	0.6	0.34	1.42	0.53	56							
CSMRI-5	6/8/10	2.42	0.82	0.75	0.38	8.7							
CSMRI-6C	6/8/10	0.34	0.23	1.48	0.58	12							
CSMRI-7B	6/10/10	0.21	0.34	0.17 R	0.9	84							
CSMRI-8	6/8/10	0.45	0.28	0.68	0.4	540							
CSMRI-9	6/9/10	0.41	0.38	0.64	0.4	48							
CSMRI-10	6/9/10	0.37	0.31	0.66	0.41	9.8							
CSMRI-11B	6/8/10	0.28	0.25	0.4	0.43	10							
M	CL*		Total F	Ra = 5		30							

Table 2-1Summary of Radioisotopes in Groundwate

*Maximum Contaminant Level - National Primary Drinking Water Regulations

pCi/L = picoCuries per Liter

 $\mu g/L = micrograms per Liter$

NT – Not Tested

R = data rejected due to laboratory QA/QC (see Section 2.3 Analyses)

Table 2-2Summary of Metals in Groundwater

			•	(T)	
- ()	$\Delta \Pi$	results	1n	$m\sigma/L$)

Sample Station	Sample Date	Ag	As	Ва	Са	Cd	Cr	Hg	ĸ	Mg	Na	Pb	V
	6/10/10	ND	ND ND			ND	ND	0.000023 B	K	15		-	ND
CSMRI-1				0.11	51				4.1		42	ND	
CSMRI-1B	6/9/10	ND	ND	0.1	170	ND	ND	0.000023 B	4.8	85	61	ND	ND
CSMRI-2	6/10/10	ND	ND	0.098 B	93	ND	ND	0.000024 B	6.9	43	25	ND	0.00094 B
CSMRI-4	6/8/10	ND	ND	0.1	140	0.00029 B	ND	0.00007 B	16	55	59	ND	ND
CSMRI-5	6/8/10	ND	ND	0.11	150	ND	ND	0.000028 B	5	54	53	ND	ND
CSMRI-6C	6/8/10	ND	ND	0.1 B	120	ND	ND	0.000026 B	5.2	56	49	ND	ND
CSMRI-7B	6/10/10	NT	NT	NT	NT	NT	NT	NT	NT	NT	NT	NT	NT
CSMRI-8	6/8/10	ND	ND	0.052 B	170	ND	ND	0.000024 B	14	60	48	ND	ND
CSMRI-9	6/9/10	ND	ND	0.07 B	150	ND	ND	0.000019	6.7	69	69	ND	ND
CSMRI-10	6/9/10	ND	ND	0.098 B	130	ND	ND	0.000025	4.7	48	49	ND	ND
CSMRI-11B	6/8/10	ND	ND	0.091 B	130	ND	0.0013 B	0.000026	5.6	55	49	ND	ND
Detection	Limits	0.01	0.01	0.1	1	0.005	0.01	0.0002	1	1	1	0.003	0.01
MCL	*	NE	0.010	2	NE	0.005	0.1	0.002	NE	NE	NE	0.015	NE

*Maximum Contaminant Level - National Primary Drinking Water Regulations

NE - Not Established

ND - Tested but not detected above the Detection Limits

NT - Not Tested

B - Reported value was less than the contract lab required detection limit but equal to or greater than the lab MDL.

						Summary o	f Anions and	Cations in G	roundwater						
Sample Station	Sample Date	Bicarbonate as CaCO ₃ (mg/L)	Carbonate as CaCO ₃ (mg/L)	Total Alkalinity as CaCO₃ (mg/L)	Chloride (mg/L)	Total Dissolved Solids (mg/L)	Dissolved Organic Carbon (mg/L)	Ferric Iron (mg/L)	Ferrous Iron (mg/L)	Total Iron (ug/L)	Nitrate (mg/L)	Nitrite (mg/L)	Dissolved Phosphorous (mg/L)	Sulfate (mg/L)	Sulfide (mg/L)
CSMRI-1	6/10/10	83	ND	83	89	360	ND	ND	ND		0.64	ND	NT	75	NT
CSMRI-1B	6/9/10	230	ND	230	250	930	2	NT	NT	NT	4.7	ND	NT	270	NT
CSMRI-2	6/10/10	330	ND	330	29	510	ND	NT	NT	NT	ND	ND	NT	95	NT
CSMRI-4	6/8/10	280	ND	280	140	820	3.4	ND	ND	ND	ND	ND	NT	230	ND
CSMRI-5	6/8/10	200	ND	200	270	840	1.4	ND	ND	ND	0.66	ND	NT	120	ND
CSMRI-6C	6/8/10	220	ND	220	230	700	1.4	NT	NT	NT	4	ND	NT	83	NT
CSMRI-7B	6/10/10	NT	NT	NT	NT	NT	NT	NT	NT	NT	NT	NT	NT	NT	NT
CSMRI-8	6/8/10	350	ND	350	150	910	4.7	ND	ND	ND	ND	ND	NT	200	ND
CSMRI-9	6/9/10	300	ND	300	210	920	1.6	NT	NT	NT	9.2	ND	NT	180	NT
CSMRI-10	6/9/10	180	ND	180	250	760	ND	NT	NT	NT	4.4	ND	NT	76	NT
CSMRI-11B	6/8/10	220	ND	220	250	720	ND	NT	NT	NT	4.4	ND	NT	79	NT
Reporti	ng Limits	5, 10 or 20	5, 10 or 20	5 or 20	1, 2 or 4	10	1	0.20	0.20	100	0.50	0.50	0.05	5, 10 or 20	0.050

Table 2-3

ND - Non Detect NT - Not Tested

Table 2-4 Summary of Radioisotopes in Surface Water

Sample	Sample	Ra-226	(pCi/L)	Ra-228	(pCi/L)	Total U (µg/L)					
Station	Date	Result	Uncertainty	Result	Uncertainty	Result					
SW-1	6/9/10	0.07	0.2	0.44 J	0.36	0.46					
SW-2	6/9/10	0.07	0.34	-0.06	0.39	0.52					
SW-3	SW-3 6/10/10 0.39		0.3	0.01	0.37	0.49					
M	CL*		Total I	Ra = 5		30					

*Maximum Contaminant Level - National Primary Drinking Water Regulations

pCi/L = picoCuries per Liter $\mu g/L = micrograms per Liter$

J = Estimated value

Table 2-5 Summary of Metals in Surface Water

(All results in milligrams per liter)

					(υ	1	,					
Sample Station	Sample Date	Ag	As	Ba	Ca	Cd	Cr	Hg	K	Mg	Na	Pb	V
SW-1	6/9/10	ND	ND	0.012 B	8.4	ND	0.001 B	0.000027 B	0.47 B	1.9	2.8	ND	ND
SW-2	6/9/10	ND	ND	0.012 B	8	ND	ND	0.000024 B	0.49 B	1.9	2.7	ND	ND
SW-3	6/10/10	ND	ND	0.012 B	8.4	ND	ND	0.000024 B	0.5 B	1.9	2.7	ND	ND
Detecti	on Limits	0.01	0.01	0.1	1	0.005	0.01	0.0002	1	1	1	0.003	0.01
M	MCLs*		0.010	2	NE	0.005	0.1	0.002	NE	NE	NE	0.015	NE

*Maximum Contaminant Level - National Primary Drinking Water Regulations

NE - Not Established

ND = Not Detected at or above the detection limit

B = Reported value was less than the contract lab required detection limit but equal to or greater than the lab MDL.

					•	Jummary of A	mons and C		flace water					
Sample Station	Sample Date	Bicarbonate as CaCO ₃ (mg/L)	Carbonate as CaCO ₃ (mg/L)	Total Alkalinity as CaCO₃ (mg/L)	Chloride (mg/L)	Total Dissolved Solids (mg/L)	Dissolved Organic Carbon (mg/L)	Ferric Iron (mg/L)	Ferrous Iron (mg/L)	Total Iron (µg/L)	Nitrate (mg/L)	Nitrite (mg/L)	Total Phosphorous (mg/L)	Sulfate (mg/L)
SW-1	6/9/10	15	ND	15	4.4	61	2.6	NT	NT	NT	ND	ND	NT	12
SW-2	6/9/10	15	ND	15	4.6	61	2.7	NT	NT	NT	ND	ND	NT	11
SW-3	6/10/10	14	ND	14	4.5	61	2.5	NT	NT	NT	ND	ND	NT	12
Reportir	ng Limits	5	5	5	0.2	10	1	NT	NT	NT	0.50	0.50	0.05	2

Table 2-6 **Summary of Anions and Cations in Surface Water**

ND = Not Detected at or above the Reporting Limits

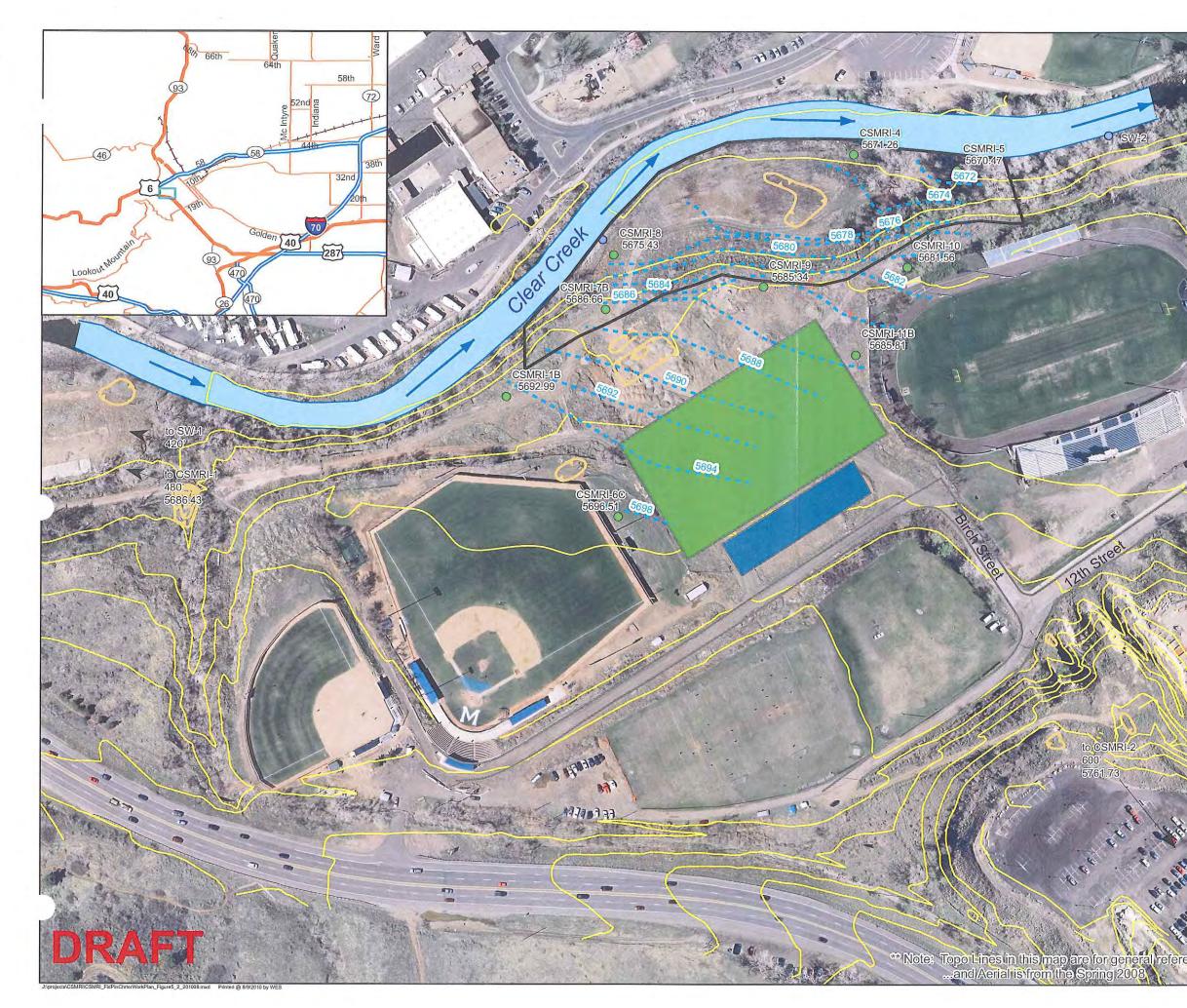
NT = Not Tested

				(All resu	ults in picoCuries	per liter)				
Well ID (d)	Analyte	1/1991 (a)	6/1991 (a)	3/1999 (b)	6/1999 (b)	10/1999 (b)	2/2003 (c)	4/2003 (c)	7/2003 (c)	10/2003 (c)
	Ra-226			0.1	0.3	0.2	< 0.55	< 0.45	ND (<0.38)	ND (<0.31)
CSMRI-1	U Total			2.09	2.59	1.44	2.4	2.9	0.87	1.4
	Th-230			0.4	0.2	0.2	<0.19	0.21	ND (<0.13)	<0.15
	Ra-226		1.9	1.9	1.4	1.4	1.4	2.8	2.1	1.7
CSMRI-2	U Total	11	5.7	0.55	1.46	0.71	1.5	1.3	1.9	1.3
	Th-230		0	0.1	0.1	0.9	<0.17	0.43	0.20	0.31
	Ra-226		0.6	1.5	1.2	1.6	<0.75	<0.81	ND (<0.49)	<0.98
CSMRI-3	U Total	17	10.4	8.41	12.4	10	12	12	9	10
	Th-230		0	0.3	0.3	1.1	<0.12	ND (<0.15)	ND (<0.17)	ND (<0.14)
	Ra-226		1	<0.4	0.6	0.4	<0.85	< 0.42	<0.32	ND (<0.64)
CSMRI-4	U Total	86	57.3	23.4	58.6	33.7	16	34.2	53	19
	Th-230		0	0.7	0.3	0.4	< 0.099	ND (<0.15)	ND (<0.17)	ND (<0.12)
	Ra-226		0.6	2.4	3.3	2.7	ND (<0.49)	1.1	2.6	1.59
CSMRI-5	U Total	14	16.8	3.6	3.6	4	2.8	2.3	2.7	3.3
	Th-230		0	0.2	0.2	1.4	0.062	ND (<0.14)	ND (<0.19)	ND (<0.13)

Table 2-7 **CSMRI** Historical Groundwater Data (Previous Consultants)

Notes: ND = Not Detected

a - Samples collected by Grant and Associates and analyzed by Barringer Labs
b - Samples collected by URS Greiner Woodward Clyde and analyzed by CORE Labs
c - Samples collected by New Horizons Environmental Consultants and analyzed by Paragon Analytics; Total U activity (pCi/L) calculated from concentration (μg/L) reported by Paragon.
d - Well Identification numbers changed from the 1991 data to the 1999 data. Data presented account for this change



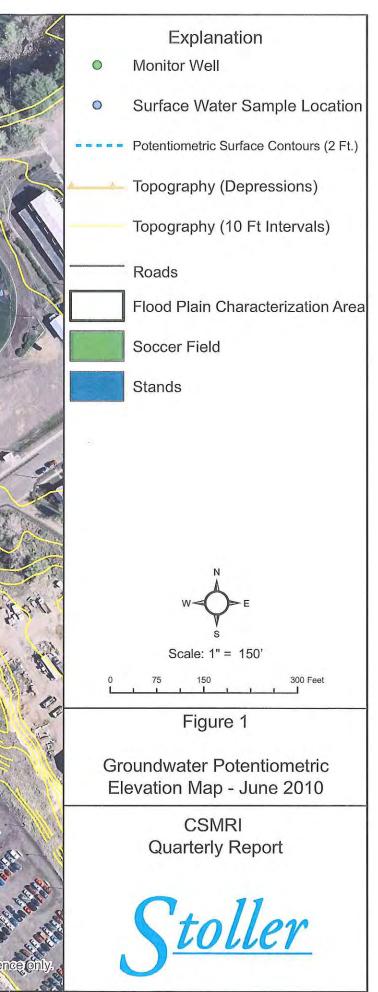
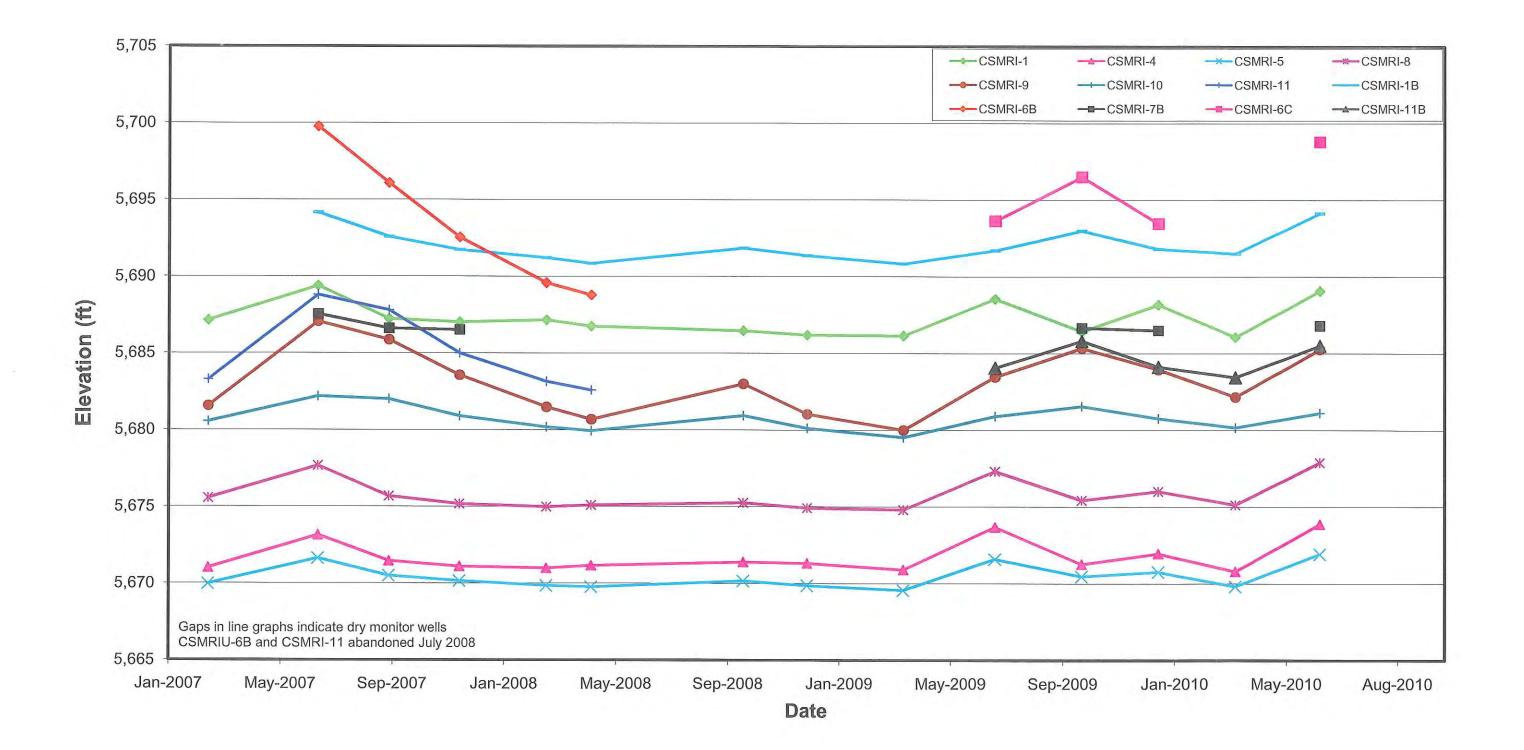


Figure 2 CSMRI All Monitor Wells Hydrograph



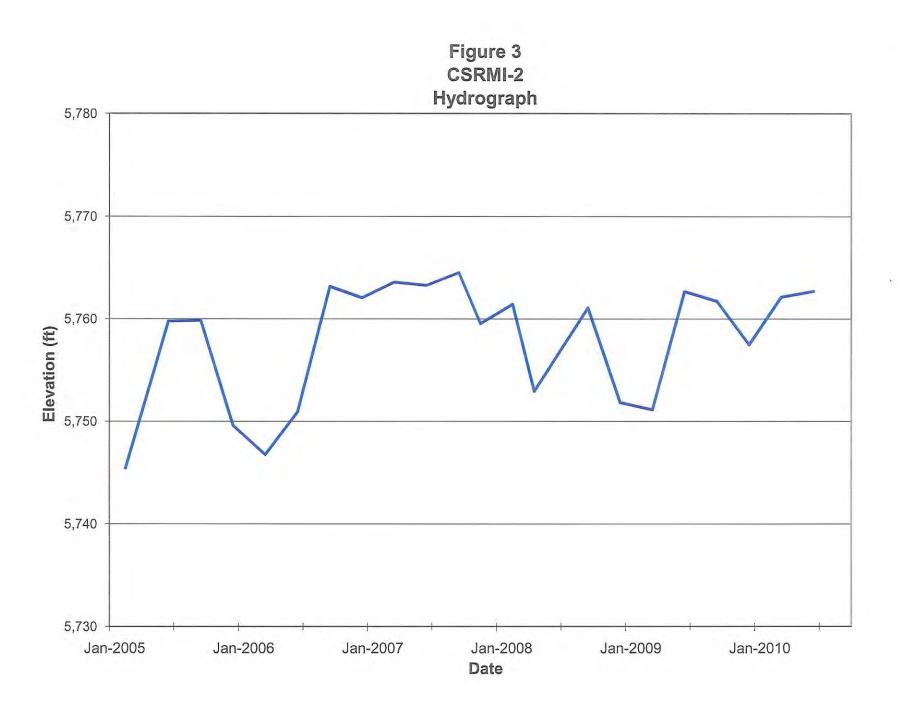


Figure 4 Clear Creek Gauging Graph April - June 2010

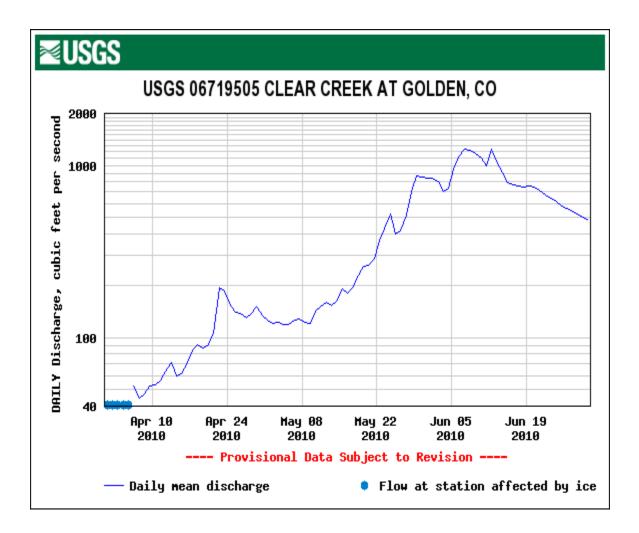


Figure 5 CSMRI-4 Historical Total Uranium Concentration (1991 - 2010)



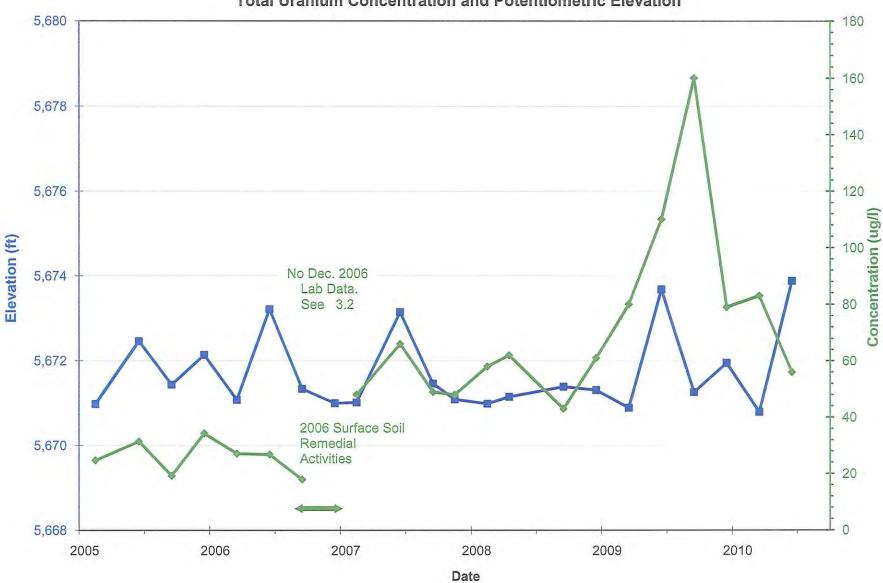
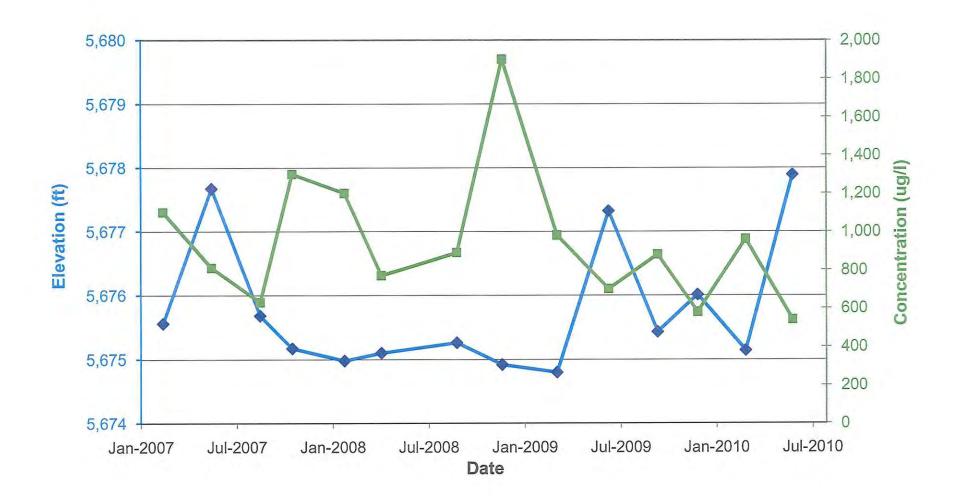


Figure 6 CSMRI-4 Total Uranium Concentration and Potentiometric Elevation

Figure 7 CSMRI-8 Total Uranium Concentration and Potentiometric Elevation



Appendix A Groundwater Sampling Procedures

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Groundwater Sampling

1.0 Purpose

This procedure describes actions to be used to sample groundwater from monitoring wells and piezometers. Monitoring wells are generally sampled on a semiannual, quarterly, or monthly basis, or by special request in support for specific projects. All wells are to be sampled using this procedure unless superseded by specific site, facility, or client procedures.

This procedure describes equipment decontamination and transport, site preparation, detection and sampling of immiscible layers, water level measurements, well purging, sample collection, field and analytical parameters, quality assurance/quality control (QA/QC) requirements, and documentation that shall be used for field data collection.

2.0 Scope

This document describes acceptable methods for the sampling of wells and piezometers.

3.0 Responsibilities and Qualifications

Personnel performing groundwater sampling procedures are required to have completed the initial 40-hour OSHA classroom training that meets the Department of Labor requirements at 29 CFR 1910.120(e)(3)(i), and must maintain a current training status by completing the appropriate annual 8-hour OSHA refresher courses. Personnel must also have read the appropriate project, site, or facility Health and Safety Plan(s). Prior to engaging in groundwater sampling activities, personnel must have a complete understanding of the procedures described within this procedure and, if necessary, will be given specific training regarding these procedures by other personnel experienced in the methods described within this procedure.

4.0 Groundwater Sampling Procedures

4.1 Introduction

Many monitoring wells are constructed of either 2-inch stainless steel, or 2- or 4-inch flush threaded PVC casing. Some piezometers are completed as monitoring wells, and they are usually constructed of ¾-inch inside diameter, flush threaded PVC casing. Some wells have been constructed to incorporate a sump below the well screen. Because these vary in length, the well construction diagrams should be consulted to determine the sump lengths for specific wells. Most piezometers are constructed with a flush threaded cap at the bottom of the well screen. However, the well construction diagrams should also be consulted for information about specific piezometers.

Procedures for groundwater sampling are designed to obtain a sample that is representative of the formation water beneath the site in question. Since an analysis of the quality of formation water is desired, standing water within the well must be purged before sampling. Also, a measure of the static water elevations is important to determine the effect of seasonal horizontal and vertical flow gradient changes during site characterization activities.

Groundwater sampling procedures can be initiated after sampling personnel take the required water level measurements and purge the well in accordance with this procedure. Methods for accomplishing each of these activities are included in this procedure in the following sequence:

- Collection of immiscible layers samples, if present
- Well purging
- Groundwater sampling using a bailer

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- Groundwater sampling using a peristaltic pump
- Groundwater sampling with a bladder pump

4.2 General Equipment Requirements

Down-hole sampling equipment shall be constructed of inert material such as polytetrafluoroethylene (Teflon[®]) or stainless steel. This equipment shall be assessed on an individual basis prior to use in the field.

The following is a primary list of well sampling and associated equipment:

- Bailers Teflon[®], stainless steel, or other appropriate inert materials
- Teflon[®] coated stainless steel cable with reels
- Peristaltic pumps and tubing
- Water level measuring devices sufficiently accurate to measure water levels to the nearest 0.01 foot
- Graduated purge water containers
- Plastic sheeting
- Distilled or deionized water
- Decontamination equipment and supplies
- Organic vapor detector (OVD)
- Gloves (nitrile)
- Calculator and watch
- Sample containers precleaned to EPA specifications
- pH paper
- Custody tape
- Coolers with sufficient blue ice to cool samples to 4°C
- Preservatives (trace metals grade)
- Disposable in-line 0.45-micron membrane filters
- Logbooks and field forms
- Black waterproof pens
- Portable laboratory equipment for measuring field parameters for pH, temperature, specific conductance, and turbidity
- Total alkalinity reagent
- Beakers and graduated cylinders

Additional equipment may be required to meet project or client health and safety standards, to perform specialized sampling, or to meet personnel and equipment decontamination requirements.

4.3 Equipment Decontamination and Transport

Equipment associated with the tasks involved in groundwater sampling shall be decontaminated upon arrival at the sampling location. All sampling equipment shall be decontaminated between

sample locations. Decontamination frequency shall be increased appropriately as field conditions dictate.

Transportation of all equipment shall be performed in a manner that eliminates any possibility of cross-contamination. Calibration solutions, fuel, decontamination solutions and wastewater, and all other sources of contamination shall be segregated from sampling equipment during transport. Purge water being transported to holding areas shall be kept in closed containers.

If the decontamination of downhole equipment is not performed at the well, used downhole equipment shall be wrapped in plastic sheeting and/or segregated from clean equipment to eliminate the possibility of cross contamination. The equipment shall then be decontaminated as soon as possible.

4.3.1 Routine Field Decontamination

Decontamination of delicate equipment and the routine decontamination of sampling equipment prior to use at each well shall consist of the following steps:

- Vigorously scrub the equipment with a brush and solution of phosphate-free laboratory grade detergent (e.g., Liquinox) and distilled water.
- Rinse the equipment thoroughly with approved distilled water.
- If the decontaminated equipment is not immediately packaged to eliminate any adhesion of airborne impurities, perform an additional final rinse, or decontamination and rinse, immediately prior to actual sampling operations.

4.3.2 Routine Decontamination of Sampling Pumps

The external surfaces of all non-dedicated pumping equipment shall be decontaminated as described in Subsection 4.3.1. Internal surfaces shall be decontaminated according to the following procedures, except under special situations where the pump(s) must be disassembled and the internal parts cleaned separately (see Subsection 4.3.3). For routine decontamination, the following procedures shall be followed.

- Pump several pump volumes of a solution of a phosphate-free laboratory grade detergent (e.g., Liquinox) and water through the equipment.
- Displace the soap solution immediately by pumping approved distilled water, equivalent to three or more volumes of the pump storage capacity, through the equipment.
- If any detergent solution remains in the pump, continue pumping distilled water through the system until the detergent is no longer visibly present. Sudsing is the common indicator used to determine incomplete rinsing.
- 4.3.3 Unusual Decontamination Requirements

When equipment becomes grossly contaminated, such as from the collection of immiscible layer samples (see Subsection 4.5), routine decontamination of sampling equipment is not considered sufficient and thus is not allowed. This situation and other unusual equipment decontamination problems shall be reported to the field site supervisor. Under certain circumstances, a pump can be disassembled and the parts cleaned separately using approved solvents (i.e., hexane, alcohol, etc.). If specific instructions are required, the field site supervisor shall consult with a management representative for proper decontamination procedures.

4.3.4 Disposition of Decontamination Water

All water generated during the decontamination of equipment used for the sampling of wells shall be containerized in either a satellite container or in the purge water container in the groundwater sampling vehicle. It will then be disposed of according to the procedure designated in Subsection 4.6.3 of this procedure.

4.4 Site Preparation

Sheet plastic may be used to protect clean equipment from contacting contaminated surfaces. Plastic bags and sheeting, along with the segregation of clean and dirty equipment, can be used to reduce the chances of cross contamination. If a mechanical bailer retrieval system is used, the amount of plastic appropriate for protection of sampling equipment may be lessened. The sampling crew members are responsible for determining the amount of plastic sheeting required.

Disposable nitrile gloves, or gloves made of other approved materials, shall be used at all times when handling sampling equipment. Gloves shall be changed between each site and as often as necessary to ensure the integrity of clean sampling equipment.

4.5 Collection of Immiscible Layer Samples

When specified in the project sampling plan, or when the well to be sampled contains immiscible layers, immiscible phases must be collected before purging activities begin. The method of choice for collecting light non-aqueous phase liquids (LNAPLS) is a bottom valve bailer or peristaltic pump. Dense non-aqueous phase liquids (DNAPL) or "sinkers" shall be collected with a bottom double check valve bailer or peristaltic pump.

In all cases, the bailer shall be carefully lowered into the well so that agitation of the immiscible layer is minimal. Any bailer used to collect immiscible layers shall be dedicated to the well that is sampled. Peristaltic pumps shall be equipped entirely with silicon, or other chemical compatible tubing, when sampling immiscible layers. The project manager shall be responsible for determining the type materials to be used for specific projects. Dedicated equipment used for collecting immiscible layers shall be decontaminated prior to and after use as described in Subsection 4.3 of this procedure, if removed from the well.

Immiscible layer sampling shall be performed as follows.

- Remove dedicated bailers from the well and decontaminate as specified in Subsection 4.3 of this procedure. Decontaminate dedicated pump tubing, if used, prior to use.
- For LNAPLs, carefully lower the bailer intake or sampling port to the midpoint of the immiscible layer and allow it to fill while it is held at this level. The bailer must be lowered into the immiscible layer slowly so that minimal agitation of the immiscible layer occurs. Peristaltic pump intakes must also be lowered to the midpoint of the immiscible layer.
- If a DNAPL layer is being sampled, use either the double check valve bailer or peristaltic pump. Lower the bailer into the well until bottom is encountered. Lower peristaltic pump intakes also to the well bottom. Care must be taken not to immerse the pump intake into accumulated sediments.
- Do not allow the bailer or line to touch the ground at any time or allow the ground to come in contact with other physical objects that might introduce contaminants into the well.
- Decontaminate all equipment immediately after sampling is completed. Suspend dedicated bailers in the well from the well cap above the high water level. Discard silicon tubing used with peristaltic pumps.

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4.6 Well Purging

Purging stagnant water from a well is required so that the collected sample is representative of the formation groundwater. The device used (bailer or pump) depends upon aquifer properties, individual well construction, and data quality objectives. Wells that contain immiscible layers will not be purged unless specified in the site-specific work plan. Any well scheduled for purging and sampling that subsequently is found to contain immiscible layers must be reported to the site supervisor or project manager. The project manager shall be notified immediately prior to continued activities.

Before obtaining water level elevations or initiating purge activities, obtain the following information in reference to the well to be sampled, and enter the applicable information on the sample collection log.

- Location code (well number)
- Previous purge volume (information only)
- Depth to top of screen (bailed wells only)
- Well sample number
- Report Identification Number (RIN)
- Sample event number

Record the location code (well number), date, sampling team members, visitors, well condition, and any other pertinent information on the sample collection log. Enter the well number, time well is opened, and other information regarding the field activities on the Field Activity Daily Log.

The field instruments shall be standardized (to check calibration) and the results recorded on the sample collection form.

Measure the depth to the top of the water column and the total depth of the well in order to determine the height of the water column in the well. Calculate the well casing volume using the well casing inner diameter and the height of the water column in the well. The formula for calculating the volume in gallons of water in the well casing is as follows:

 $(\pi r^2 h)$ 7.481 = gallons; where

 $\pi = 3.142$

r = inside radius of the well pipe in feet

h = linear feet of water in well

7.481 = gallons per cubic foot of water

1 gallon = 3785 ml

Calculations of the volume of water in typical well casings may be done as follows:

- a. 2" diameter well:
 - 0.16 gal./ft x (linear ft of water) = gallons of water
- b, 4" diameter well:
 - 0.65 gal./ft x (linear ft of water) = gallons of water
- c. 3/4" diameter well:

87 ml./ft x (linear ft of water) = milliliters of water

4.6.1 Purging Duration

Purging shall be considered complete if any of the following conditions are met.

- Purging is complete if at least three casing volumes of water are removed from the well, and the last three consecutive pH, specific conductance, and temperature measurements do not deviate by more than the following: 1) pH = ±0.1 pH units; 2) Specific Conductance = ±10% and; 3) temperature ±0.5°C. A turbidity measurement will be taken for every other purge sample for wells that are purged using a bailer. For wells that are equipped with a dedicated bladder pump, the turbidity will be measured each time the parameters are taken. The purge rate should be such that the turbidity is maintained at 5 NTU units or less (if possible). If the readings are not stabilized after three volumes, continue purging until stabilization or until five volumes have been removed. Field parameter measurements shall be collected after every half-casing volume (approximate) is removed from the well. When casing volumes are less than 1-liter, parameter measurements will be collected after each whole casing volume is removed. If readings do not stabilize after five well volumes have been recovered, obtain additional guidance from the project manager concerning the proper course of action.
- 2. A well is considered dewatered when only a few milliliters of water (or none) can be recovered each time the bailer is lowered into the well. When this occurs, a 10-minute recharge rate will be calculated (linearly). If, at the end of the 10-minute period, the well has not recovered sufficiently to continue the purge in thirty minutes, the purge is considered completed. If, at the end of the 10-minute period, there is sufficient water to collect the VOA samples, the samples may be collected at that time. If the well has not recovered sufficient water during the 10 minutes, and depending upon the well history, the samplers may elect to return to the well the same day (preferably within two hours), check the water level, and collect the VOA samples (first), and other samples as feasible. If the sample team cannot return the same day, the well will be checked in 24 hours to determine if sample collection is feasible. If an extended period of time is required to collect samples, the procedures in Subsection 4.8.1 shall be followed. The well will not require an additional purge before sampling.

Wells that dewater (have a slow recharge rate as specified in 2 above) will not be restricted by parameter stabilization requirements. Sampling of these wells will follow the protocol established in Subsection 4.8.

4.6.2 Purging Methods

Wells will be purged by either bailing or pumping. When purging a well, the rate of water withdrawal during purging should not exceed the rate of withdrawal at which the well was developed (if known). All purge times (initiation and completion) and the rate of purging will be recorded on the field log sheets.

4.6.2.1 Bailing

Generalized procedures for purging a well with a bailer are as follows.

Prepare the sampling site as discussed in Subsection 4.4. Use properly decontaminated equipment to determine the static water level of the well.
 Measure the total depth of the well. Use this information to determine the volume of water in the well casing.

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- Decontaminate all dedicated bailers prior to initiating purging as described in Subsection 4.3 of this procedure.
- Use a mechanical reel equipped with Teflon[®] coated stainless steel cable attached to a bailer for bailing and sampling operations. Lower the bailer slowly into the well until water is encountered. Minimize agitation of the well water. Avoid lowering the bailer to the bottom of the well so sediments accumulated in the bottom do not become suspended. For wells that dewater, do not allow the bailer to strike the well bottom with force. Raise and lower the bailer carefully to limit surge energy and ensure that cable does not come in contact with any potentially contaminated surfaces. Do not allow the cable to drag along the well casing or against other objects that will cause fraying. Monitor the amount of water purged.

Wells with significant levels of contamination may have dedicated bailers installed. Dedicated bailer systems shall consist of a Teflon[®] bailer with check valve or double check valve for DNAPLS and a 5-foot leader of Teflon[®] coated stainless steel cable. Bailer sampling attachments and the stainless steel reel cable will not be dedicated to individual wells.

Dedicated bailers will be decontaminated at the conclusion of sampling activities and suspended from the well cap above the high water table. If the well interval above the high water table is not adequate to allow for storage in the casing, the dedicated bailers will be stored in labeled and sealed plastic bags at the equipment trailer.

4.6.2.2 Pumping

Pump designs that meet the following criteria are allowed for purging.

- The pump is constructed of a material that does not introduce a source of contamination to the well.
- The pump drive system does not introduce a source of contamination into the well.
- All downhole parts to the pump can be easily decontaminated.
- A return check system that does not allow pumped water to return to the well is integral in the pump design.
- The pump is easily used and does not require excessive amounts of time to install, use, remove, and decontaminate.

The pumps currently in use to purge groundwater include peristaltic pumps and dedicated submersible bladder pumps. A procedure for the use of each style of pump is specific to its applications. User manuals, which accompany each pump, shall be referenced for operating procedures.

Basic operating procedures common to all pumps are as follows.

- Prepare the sampling site as described in Subsection 4.4 regardless of the type of pump being used.
- Use properly decontaminated equipment to determine the static water level and the total depth of the well. This information is utilized to determine the volume of water in the well casing.

- For wells with dedicated pumps, calculate the minimum purge volume using the pump storage volume and the volume of the discharge tubing. A total depth of a 2-inch well cannot be taken without the removal of the pump.
- Position a dedicated pump near the bottom of the well or according to the information on the well construction form. Monitor the discharge rates and the amount of water purged during purging. The pumping rate for purging can be higher than the pumping rate for sampling, however, the water level in the well should be monitored during purging to avoid excessive water level drawdown.
- Ensure that any tubing that enters the well casing is composed of inert material. Disposable silicon tubing will be used in the drive mechanism of peristaltic pumps and discarded after each well is purged. The air supply for all air-driven pumps (dedicated bladder pumps) will be free of oil (i.e., no hydrocarbon containing substances will be added to the compressor).

4.6.3 Disposition of Purge Water

All water removed from a well during sampling operations shall be collected either in a satellite container or the purge water collection container in the groundwater sampling vehicle. The water from these containers will then be transferred to another approved collection container on the sampling or project site. When the collection container is filled, or is near capacity, it will be transported for disposition or treatment in accordance with approved project plans.

4.7 Measurement of Field Parameters

The following field parameters will be measured during groundwater purging operations unless otherwise specified by the project manager or the approved project work plans.

Parameter	Relative Precision	Minimum Calibration	
pН	0.01 pH units	Daily	
Conductivity	$10 \ \mu \text{S/cm}$	Daily	
Temperature	0.1 °C	Weekly	
Total Alkalinity (unfiltered)	1 mg/l	None	
Turbidity (photometric)	2 FTU (or NTU)	Specified purge samples (bailed wells) Daily (dedicated bladder pump wells)	

The measuring equipment shall be stored and handled in a manner that will maintain the integrity of the equipment. Appropriate field manuals will accompany each instrument in the field. Each instrument will also be given an identification number. All logbook and field form references to individual instruments will refer to this number for ease of identification.

Field parameters will be measured at the following intervals.

• Conductivity, pH, temperature, and turbidity shall be measured from the first water removed from the well when initiating well purging procedures. For bailed wells, the initial bail of water will be carefully removed from the well and the water transferred to a sample beaker by decanting the bailer through a bottom control valve. For wells

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purged with a peristaltic pump, similarly collect the first water removed in a sample beaker and then measure parameters. For wells with dedicated pumps, measure the parameters of the first recovered water that is collected in the continuous sampler.

- During purging operations, conductivity, pH, and temperature shall be measured for every half-casing volume (one half of the initial casing volume as calculated on the sample collection log form) of water removed from the well (because of the accuracy of the graduated containers for the purge water, the purge volume will be estimated as close as feasible). For wells that have half volumes less than the volume of a sample bailer (approximately 1 liter), only measure parameters after each full casing volume of water is removed from the well. Turbidity will be measured on every other sample recovered for parameters for bailed wells, or wells purged with a peristaltic pump. All parameters, including turbidity, will be measured at predetermined intervals while purging wells with dedicated pumps.
- During purging, if a well is dewatered prior to the measurement of the final required set of parameters, then conductivity, pH, temperature, and turbidity shall be measured immediately before the start of sample collection. These parameters may be delayed until sampling is completed if, at the discretion of the sampling crew, the well recharge has provided insufficient water volume to collect all the samples and also measure parameters. If there is insufficient water for samples and field parameters, the parameters will not be measured.
- Total alkalinity measurements shall be collected only once upon completion of purging. For wells that do not dewater and sample collection proceeds to completion immediately after purging, alkalinity will be measured after the completion of all other final purge field parameters. Wells that dewater and require repeated visits for the collection of samples will have alkalinity measured subsequent to the collection of the sample for inorganic water chemistry. Alkalinity will not be measured if sufficient water is not available.
- For micro purged wells, a purge is considered completed when the parameters have stabilized.
- Whenever a method used to remove well water is changed, a set of field parameters shall be recorded from water removed with the new method.

4.8 Groundwater Sampling

Techniques used to withdraw groundwater samples from a well shall be based on consideration of the parameters of interest. The order of collection, collection techniques, choice of sample containers, preservatives, and equipment are all critical to ensuring that samples are not altered or contaminated. The preferred methods for collection of groundwater samples are either bailing and/or the use of bladder pumps.

Sites shall be prepared prior to sampling as described in Subsection 4.4. All necessary and appropriate information will be recorded on the sample collection log and on the Field Activity Daily Log.

4.8.1 Sample Collection

The following discussion involves collection of groundwater samples using bailers and peristaltic or bladder pumps. Regardless of the collection method, care shall be taken not to alter the chemical nature of the sample during the collection activity by agitating the sample or allowing prolonged contact with the atmosphere. To minimize the potential for

altering the sample and to maximize the available water, the following sample collection sequence is preferred.

- Radiation Screening
- VOC
- Nitrate/Nitrite, as N
- Dissolved Metals TAL, with Cs, Li, Sr, Sn, Mo, Si
- ^{239/240} Plutonium, ²⁴¹ Americium
- ^{233/234}U, ²³⁵U, ²³⁸U
- Gross alpha and beta
- ^{89/}Strontium
- ¹³⁷Cesium
- ^{226,228}Radium
- Tritium
- Total Metals TAL, with Cs, Li, Sr, Sn, Mo, Si
- TDS, CL, F, SO⁴, CO3, HCO³
- TSS
- BNA
- Pesticides/PCB
- Cyanide
- Orthophosphate

VOC samples shall be collected first and as soon as possible after the well has been purged. If a well is purged using a peristaltic pump, then all other samples shall be collected prior to removing the pump from the well. The VOC sample will then be collected using a bailer.

For wells that dewater, if a sufficient volume of water for VOC sample collection has still not accumulated within 48 hours after the completion of purging, VOCs will not be collected for that well. Other samples may be collected using a maximum of five attempts to recover sufficient sample water for analysis. This procedure is discussed in the following paragraph.

The containers used for sample collection from poor producing wells may differ from those used for high yield wells in some instances due to constraints on obtaining enough sample to fill sample containers. In some instances smaller containers may be utilized, or analyte samples normally collected in separate containers may be combined into a single container. Well histories can be used to identify which wells may require a modified sample suite and an extended sampling period. These wells will initially be sampled for a period of 48 hours after the completion of purging, with the exception of VOC sample collection, which is discussed in the previous paragraphs. The completion of purging will be considered 0 hour. At the end of 48 hours, any partial sample will be measured. The accumulated sample will be compared to the minimum volume requirement identified in Table 1 and the allowed sample holding time. If the minimum volume requirement for the target analyte has not been achieved, then sampling may continue as determined from the well recharge

history. All analyte samples that have only minimum sample volumes collected, and all uncollected samples will be documented on the sample collection log.

Table 1

Sample Container	s and Preservatives	or Groundwater	Samples
------------------	---------------------	----------------	---------

Parameter	Minimum Container ¹	Preservative	Holding Time
Radiation Screen	120 ml poly	None	NA
VOC - CLP	3 – 40 ml amber glass	Cool to 4° C	4 Days
BNA	1 L amber glass	Cool to 4° C	7 Days
Pesticides/PCB	1 L amber glass	Cool to 4° C	7 Days
TSS	125 ml poly	Cool to 4° C	7 Days
TDS, CI, F, SO4, CO3, HCO3	1 L poly	Cool to 4° C	7 Days
Dissolved Metals - CLP, with Cs, Li, Sr, Sn, Mo, Si	1 L poly	*Filtered, HNO ₃ to pH <2, Cool to 4° C	6 Months
TOC	125 ml poly	H ₂ SO ₄ < pH2, Cool to 4° C	28 Days
COD	125 ml poly	H ₂ SO ₄ < pH2, Cool to 4° C	28 Days
Total Metals - CLP with Cs, Li, Sr, Sn, Mo, Si	1 L poly	Unfiltered, HNO ₃ to pH <2, Cool to 4° C	6 Months
Orthophosphate	250 ml poly	Filtered, Cool to 4° C	2 Days
Nitrate / Nitrite as N	250 ml poly	H ₂ SO ₄ to pH <2, Cool to 4° C	28 Days
Cyanide	1 L poly	NaOH to pH >12, Cool to 4° C	14 Days
Gross Alpha / Beta	550 ml poly	HNO3 to pH <2	6 Months
233/234U, 235U, 238U	100 ml poly	Filtered, HNO ₃ to pH <2	6 Months
239/240Pu	1 L poly	HNO₃ to pH <2	6 Months
²⁴¹ Am	1 L poly	HNO₃ to pH <2	6 Months
^{89/90} Sr	700 ml poly	Filtered, HNO ₃ to pH <2	6 Months
^{226/228} Ra	750 ml poly	Filtered, HNO ₃ to pH <2	6 Months
¹³⁷ Cs	2.5 L poly	Filtered, HNO ₃ to pH <2	6 Months

¹ The volume listed is the minimum amount required for analysis. Actual sample volumes may be slightly higher and some parameters may be combined in a single container.

* Some samples may not require filtering if taken from a well with a dedicated pump and turbidity of 5 NTU or less.

The order of sample collection may be changed at the discretion of the sampling team. Changes in the order shall be based on the predicted volume of water that will be recovered and the priority stated in the controlling document. The sampling team shall document their sample selections on the sample collection log.

Sample containers shall be stored away from sunlight and cooled to 4°C prior to filling. Immediately after collection, samples requiring cooling shall be cooled to 4°C. A chilled cooler shall be used as the storage container. Whenever a sample bottle that requires chilling is not being physically handled, it will be placed in the cooler to prevent heating or freezing, exposure to sunlight, and possible breakage.

VOC samples shall be collected using a bailer equipped with a bottom-decanting control valve or directly from the pump discharge line on wells equipped with bladder pumps. The procedures for collecting VOC samples are discussed in Subsections 4.8.1.1 and 4.8.1.2 of this procedure.

VOC vials shall never be filled and stored below capacity because of insufficient quantities of water in the well. Except for the VOC vials, adequate air space should be left in the sample bottles to allow for expansion.

Samples shall be placed in the appropriate containers and packed with ice in coolers as soon as practical. VOC samples will be stored in the cooler in an inverted position immediately after collection. When sampling is complete, the well cap shall be replaced and locked.

Sampling tools, instruments, and equipment shall be protected from sources of contamination before use and decontaminated after use as specified in Subsection 4.3. *Liquids from decontamination operations will be handled in accordance with the procedures in Subsection 4.6.3 of this procedure.* Sample containers shall also be protected from sources of contamination. Sampling personnel shall wear chemical-resistant gloves (e.g., nitrile) when handling samples, and the gloves will be disposed of between well sites.

4.8.1.1 Groundwater Sampling Using a Bailer

This subsection describes the use of a bailer for collecting groundwater samples that may be used to obtain physical, chemical, or radiological data.

A bailer attached to a Teflon[®] coated stainless steel cable is carefully lowered into the well. After filling within the well, the bailer is withdrawn by rewinding the bailer line, and the bailer contents are drained into the appropriate containers. Certain recommendations and/or constraints should be observed when using bailers for sampling groundwater monitoring wells, as follows.

- Use only bottom-filling Teflon[®] bailers or bailers made of other inert materials.
- Ensure that bailers are attached to a Teflon[®] coated stainless steel line that is pre-wound on a reel.
- Do not use bailers constructed with adhesive joints.
- Lower the bailer slowly to the interval from which the sample is to be collected.

VOC samples shall be collected using a bailer equipped with a bottom-decanting control valve. The first water through the valve assembly will be discarded into the purge water container. Vials will be filled by dispensing water through the control valve along the inside edge of the slightly tilted sample vial. Care shall be taken to climinate aeration of the sample water. The vials will be filled beyond capacity so the resulting meniscus will produce an airtight seal when capped. The capped vial will be checked for trapped air by lightly tapping the vial in an inverted position. If air becomes trapped in the vial, the sample water shall be discarded, and the vial refilled. If two consecutive attempts to fill a VOC vial result in trapped air bubbles, the vial shall be discarded.

The remainder of the sampling water shall be collected in a stainless steel container from which the remaining sample bottles will be filled. Samples requiring filtration shall be filtered and then containerized.

4.8.1.2 Groundwater Sampling Using a Peristaltic Pump

Use of peristaltic pumps shall generally be limited to collecting sample aliquots for radionuclides, metals, and other species that are not subject to volatilization and degassing. Peristaltic pumps shall never be used to collect VOCs or other

volatile species in routine wells, although such samples may be collected for special screening applications. All downhole tubing shall be Teflon[®] except in areas of special concern (e.g., where immiscible layers exist) where special tubing, such as stainless steel or Viton[®], may be required. If so, the project manager will make this determination. Only the portion of tubing that is inserted into the mechanical drive shall be made of silicon. This drive portion of the tubing shall be discarded after each use.

4.8.1.3 Groundwater Sampling Using a Downhole Bladder Pump

Some wells are equipped with dedicated downhole bladder pumps for purging and sampling. These are wells that will normally produce an adequate amount of water during a single visit to complete the required sampling suite. The equipment required to purge and sample a well consists of a pump control unit, a portable air compressor, a continuous sampler for measuring the field parameters, and the necessary sample containers, graduated cylinders, and container(s) to collect the purge and excess water. The following precautions should be observed during the sampling operation.

- Locate the compressor used to power the pump downwind from the well to eliminate the contamination of equipment and samples with exhaust.
- If the flow-through cell will not maintain a full sample chamber (tends to drain back), then clean the check valve on the pump if it is fouled, or replace the pump.
- Calculate the minimum purge volume using the procedure in Section 4.6. Note that a purge is considered completed only when the groundwater parameters have stabilized.
- Upon completion of purging, initiate sampling with the collection of the VOC sample(s). The pump should operate with minimum interruptions while the full sample suite is collected. Allowing the pump to stop for an extended period of time will cause the water trapped in the discharge lines to equilibrate to ambient temperatures, which is not acceptable. During sampling, the pump can be slowed to any rate that allows efficient sampling while also maintaining stable field parameters.
- Measure groundwater parameters periodically during sample collection and record them on the sample collection log to document conditions during sampling.
- Because micropurging is the method used for sampling, adjust the flow rate to limit the drawdown in the well. Also adjust the rate such that the turbidity is below 5 NTU for sampling. If this criterion is met, the samples need not be filtered.
- Operate the pump, pump control unit, and the flow-through cell according to the manufacturer's recommendations.
- 4.8.1.4 Groundwater Sampling Using a Push Type Sampler

This portion of this procedure describes the use of a Geoprobe[®] Screen Point 15 Groundwater Sampler, or similar type equipment, for collecting groundwater samples at predetermined depths. These samples may be used to obtain physical, chemical, or radiological analyses.

A Geoprobe[®] Screen Point 15 Groundwater Sampler, or equivalent tool, is driven to a predetermined depth by a push type-sampling rig. The Screen Point 15 Groundwater Sampler is equipped with a 41-inch retractable screen and expendable drive point. It can then be partially or fully withdrawn (up to 41 inches) to expose a portion or the entire deployed well screen. After groundwater enters the exposed screen, a sample is collected using either the procedures in Subsection 4.8.1.1, Groundwater Sampling Using a Bailer, or in Section 4.8.1.2, Groundwater Sampling Using a Peristaltic Pump. Note that these samples are collected only for screening purposes because the sampling tool hole has not been completed as a well.

The method for obtaining QC samples using the push type-sampling tool is provided in Subsection 4.8.4.1 for groundwater sampling. Duplicate groundwater samples shall be collected only if there is enough water to collect two full suites of analytes without dewatering the annulus. If insufficient water is available for the collection of a planned QC sample, it shall be explained and documented in the field log book, and the project manager informed. If insufficient water is available for two full suites of analytes, it may be come necessary to prioritize the analyte list. The prioritization sequence should be described in the project-specific work plan.

4.8.2 Sample Filtering and Preservation

Samples for dissolved metals, Gross Alpha/Beta, ^{233/234}Uranium, ²³⁵Uranium, ²³⁸Uranium, ^{89/90}Strontium, ¹³⁷Cesium, ²²⁶Radium, ²²⁸Radium, and orthophosphate shall be filtered in the field at the well location during the sampling event through a disposable 0.45-micrometer membrane filter. If a peristaltic or bladder pump is used, a disposable filter may be attached directly to the sample delivery line so that the sample is filtered directly into the sample container as it exits the delivery line. Discharge pressure shall be gauged so it does not exceed 50 psi. Alternatively, sample water may be collected in a stainless steel container and filtered with a peristaltic pump. Before sample collection, 100 to 200 milliliters of sample water shall be passed through the filter in order to rinse the filter and filtration apparatus of possible contaminating substances.

Preservatives shall be added to the sample bottles prior to the introduction of the filtered sample water. The preservative shall be added in aliquots appropriate to the size of the bottle.

After sample collection has been completed, the pH of preserved samples shall be checked as follows.

- Pour a small amount of sample from the sample bottle directly onto approved pH paper. Use care so that the threaded neck of the bottle does not contact the pH paper. Do not, under any circumstances, insert the pH paper into the sample bottle.
- Check the pH paper against the supplied color chart. If the appropriate pH has not been achieved, add additional preservative to the sample in 5 ml aliquots and repeat the pH test after each addition.

4.8.3 QA/QC Samples

The frequency and types of field QA/QC samples collected during groundwater sampling are described in project-specific work plans or quality assurance plan documents. These documents detail the applicable criteria for collecting QA/QC samples.

4.8.3.1 Duplicates

Duplicate samples shall be collected only from wells that produce enough water to collect two full suites of analytes without dewatering. Wells that produce sufficient water shall be incorporated into the sampling program such that the required duplicate frequency can be maintained.

Wells scheduled for duplicate sample collection shall be sampled as described in Subsection 4.8 of this procedure, and in relevant sections of project-specific work plans and/or quality assurance documents. Field duplicates are collected following the same sampling procedures used to obtain the real samples. With the exception of VOCs, the typical procedure for a location is to collect the real and duplicate of each sample at the same time, in two equal portions, with each portion going to the laboratory in separate containers. This is accomplished by alternately filling two sample bottles one half at a time to minimize heterogeneity. Note that real and duplicate VOC samples shall be collected independently to reduce the possibility of volatilization of the sample.

When a well with a dedicated pump is being used for sample collection, all samples shall be collected in the normal order, with duplicate VOC samples being collected first. The remaining samples will be sampled as described above.

If a well is being used for matrix spike (MS) and matrix spike duplicate (MSD) samples, the duplicate shall be collected after collection of the MS and MSD.

All duplicate samples shall be given a sample number different from the original sample and the information recorded on the sample collection log and/or the field QC sample collection log.

4.8.3.2 Matrix Spike and Matrix Spike Duplicate

MS and MSD samples shall be collected only from wells that produce enough water to collect the required suites of analytes without dewatering. MS and MSD samples are not collected on a routine basis, but will be collected if so designated in a site-specific sampling plans, or if requested by the project manager.

MS and MSD samples shall be collected as follows.

- Purge the well as described in Subsection 4.6 of this procedure..
- After completion of purging, collect VOC samples. Collect the real sample followed by the MS and MSD. Collect these samples in immediate succession.
- Collect the remaining samples not requiring filtering. For each sample parameter, collect the original sample, MS, and MSD concurrently. Fill the original sample bottle one-third full followed by the MS and MSD sample bottles, which are also filled one-third full. Rotate each bottle in the sequence, filling in one-third full until all three bottles are full. For analytes not requiring an MSD, collect only the original sample and the MS.
- After the real sample, MS, and MSD (where appropriate) are collected for one parameter, repeat the process for the next parameter.
- Similarly, collect samples requiring filtering. When a bailer is used, fill a stainless steel bucket with sample water. As samples are collected and the reservoir of water in the bucket is depleted, add more water with discretion. When a pump is used, attach the filter directly to the discharge line. Fill

sample bottles as described above, partially filling the original sample, MS, and MSD in rotating sequence until each parameter bottle is full.

- Radiochemistry samples may have more than one bottle for each parameter group. In this case, include all required bottles in the rotating sequence.
- Field parameter measurements are not be required for MS and MSD samples.
- Retain the original sample number for MS and MSD samples. However, add a suffix of MS or MSD to the sample number to correspond with each QA/QC sample. Record all information on the field QC groundwater sample collection log.
- 4.8.3.3 Replicates and Splits

Replicate and split samples shall be collected in the same manner as described for the MS and MSD. Seek instruction from the project manager for replicates and splits exceeding three samples. Record all information will be recorded on the groundwater sample collection logs.

4.8.3.4 Field Equipment Rinses

Wells scheduled for equipment rinsate samples shall be sampled as described in Subsection 4.8 of this procedure, and field equipment rinses shall be collected as described in this Subsection and in relevant portions of project-specific QC documents and work plans. Field equipment rinses shall be collected in a manner designed to reflect sampling techniques. All equipment used during sampling will be fully decontaminated as described in Subsection 4.3, then rinsed with distilled or deionized water. The rinse water will then be collected in bottles identical to those used for the original sample, and assigned a separate sample number. Analytes requiring filtration will be filtered using a new filter and tubing as required for the real sample. All information will be recorded on groundwater sample collection logs.

4.8.3.4.1 Bailed Wells

After completion of sampling, all equipment shall be decontaminated. Prior to leaving the well location, the equipment rinse will then be collected as follows.

- Fill the bailer with distilled or deionized water by pouring the water into the top opening.
- Decant the rinse water to the VOC vials through the bottom valve just as was done during sample collection.
- For the remaining unfiltered samples, fill the bailer with distilled or deionized water each time additional rinsate is needed. Transfer the rinsate to sample bottles or to a stainless steel bucket and then to sample containers in the same manner used during collection.
- Collect filtered samples in an identical manner as the real samples. Fill the bailer with distilled or deionized water. Then transfer the rinse water to a stainless steel bucket. Filter the rinse water in the bucket through a new disposable filter.

- Preserve rinse samples in the same manner as the real samples.
- 4.8.3.4.2 Pumped Wells

Rinsate samples are not routinely collected from wells that are equipped with dedicated bladder pumps because the samples from these wells are collected directly from the pump discharge line. However, wells sampled using peristaltic pumps for sampling may be selected for rinsate sampling, with equipment used in sample collection (down hole tubing, filter tubing and the stainless steel bucket used for sample water collection, etc.) being decontaminated prior to rinsate sampling. The tubing at the pump head will be replaced, and a new filter used for filtered analytes. To collect the samples, distilled or deionized water will be poured into the decontaminated stainless steel bucket and pumped, using the decontaminated tubing, into the sample containers. The equipment used to collect the real VOC samples will also be decontaminated, rinsed, and used to collect the VOC rinse samples. All samples will be preserved at the same pH levels as the real samples.

4.8.3.5 Distilled Water Blanks

Distilled water sample blanks are not submitted on a routine basis, but will be made up if so designated in a site-specific sampling plan. Samples of the distilled or deionized water used for the final decontamination of equipment will be transferred directly to sample bottles to determine any baseline contamination the water may have introduced into the samples. Five-gallon bottles of the distilled or deionized water will be opened in a controlled area, such as the bottle storage room, and then poured directly into the appropriate sample bottle. A Teflon[®], glass, or stainless steel funnel may be used to help control flows into small mouth bottles. Blank samples will be preserved to the appropriate pH required for each analyte. All information will be recorded on groundwater sample collection logs.

4.9 Sample Handling and Control

Pre-cleaned sample containers will be obtained from a contract analytical sample container source. Preserving solution will be added to the bottles by a laboratory, the sample manager or qualified sampling personnel. The bottles will be labeled to indicate the preservative added.

The sampling containers, preservation requirements, and holding times for the various types of analyses are shown in Table 1. Groundwater samples will be properly labeled so that they can be easily identified. The sample numbering system will be assigned by project-specific sampling plan documents. A sample identification (ID) number will be assigned to each sample suite. The sample ID number will contain the following information as part of a nine to twelve character, alpha-numeric code:

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Character(s)	Description	Code
1 and 2	Project ID	GW
3 through 7	Sample Number	00001 to 99999
8 and 9	Subcontractor ID	Alpha (e.g. TE = Tierra Environmental Consultants)
10, 11, and 12	QA/QC	MS for matrix spike, MSD for matrix spike duplicate

In addition to a sample number, each well sampled will be assigned a current Record Identification Number (RIN), an event number (specific to the RIN), and bottle numbers that are specific to the RIN and event number.

5.0 Records

All field activities shall be recorded on a Field Activity Daily Log or Groundwater Sample Collection Log. Additional logs may be required to record QC samples and for recording well status. Refer to specific project, site, or facility work plans for further information. Summary information of the day's activities or other pertinent information should always be recorded on the field forms. Under some circumstances, the project manager may assign a bound field logbook to the field personnel that will remain in their custody during all sampling activities. The cover of each logbook shall contain the following information at a minimum:

- Name of the organization to which the book is assigned
- Book number
- Project name
- Start and end dates

Logbook pages shall be sequentially numbered and marked with the book number before any data are recorded. All data and information pertinent to field sampling shall be recorded in the logbook or on the field forms that identify all required data entries. Enough detail must be included in the documentation to reconstruct the sampling event. Field form entries shall include the following minimum information:

- Date and time
- Names of field personnel
- Names of all visitors
- Location of field activities
- Description of sampling sites including weather conditions
- All field observations and comments
- Field parameters
- Sample identification information
- · References to all prepared field activity forms and chain-of-custody records

Field logbooks, when required on specific projects, shall normally be kept only by the field sampling team leaders and the site supervisor and shall typically be used only to summarize field activities and to document project information not required by the procedure field forms.

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Permanent ink shall be used for all entries in the logbooks and on the field forms. Mistakes shall be crossed out with a single line, initialed, and dated. Unused pages or partial pages shall be voided by drawing a line through the blank sections and initialing and dating the mark. Any deviation from this procedure shall require documentation in the site supervisor's logbook.

The field activity daily log narrative should create a chronological record of the sampling team's activities, including the time and location of each activity. Descriptions of problems encountered, personnel contacted, deviations from the procedure, and visitors on site shall also be included. The weather conditions, date, signature of the person responsible for entries, and the number of field activity daily log sheets used to record media team activities for a given day shall also be included.

The Groundwater Levels Measurement/Calculations Form and the Chain of Custody Record (see *Containing, Preserving, Handling, and Shipping Soil and Water Samples*) shall also be completed for each site. All blank fields on the forms must be completed or voided.

6.0 References

- Environmental Protection Agency, 1982, Test Methods for Evaluating Solid Waste, SW-846, Volume II. Field Methods, 2nd edition.
- Environmental Protection Agency, 1986a, Engineering Support Branch Standard Operating Procedures and Quality Assurance Manual, EPA Region IV Environmental Service Division.
- Environmental Protection Agency, September 1986b, RCRA Ground Water Monitoring Technical Enforcement Guidance Document, OSWER-9950.1.
- Environmental Protection Agency, 1987a, A Compendium of Superfund Field Operations Methods, EPA/540/P-87/001. 1987.
- Environmental Protection Agency, 1987b, Data Quality Objectives for Remedial Activities, Development Process, EPA/540/G-87/003.
- Environmental Protection Agency, December 1988, User's Guide to the Contract Laboratory Program.

Appendix B Sample Collection Forms

Project Name Colorado	school of	Min.	5	Sample Location $CSMR - I = I$
Project Number 43	49-250			Date. 6/10/10
Sample Type	Duplicate	SW Other	ER	Sampler N Malozyt-

Purge Volume Calcula	Sample Collection					
Measured TD =	25.02 (ft (+.28)	Ra-226	Container	Preservative	Date	Time
Total Deptl) =		-228 Diss. U	cuse	HNO	6/10/10	1130
Depth to Water -	5-35 (11	and produce the second s	500ml Plastic	HNON	6/10/10	1130
Initial Water Column -	19.95	Anions	500ml Plastiz		6/10/10	1130
Initial Water Volume	3.20 (gal	DOC	125ml Amber	H- 50,	6/10/10	1130
3 X Water Volume	9-60 (gal)	Lab Mrs-	Ft. Collm	· Test /	America -	

Tuna	Volume	Temperature	pН	Conductivity	DO	ORP	Turbidity	Appearance
	(gal)	(C)FI	(SU)	(uS/cm)	(mg/L)	(mV)	(MTU)	
1108	1.60	12.07	7.06	976	9.88	162	1000+	6.on
1111	3,20	10.87	6.90	865	9.97	167	1	
1114	4.80	10.50	6.76	866	9.97	173		
1116	6.40	11.36	6.72	852	12.70	177		
1119	8.00	11.03	6.67	878	12-92	179		
1122	9.60	10,69	6.66	871	13.00	180	+	4
						1		
	1							Arm

Comm	ents Analysis	Container	Preservative	Dete	Time
	Diss. Mehls	500nd Plactor	HNOZ	6/10/10	1130
TA	Noz, NOz	16 Plactre	rene	6/10/10	1130
TA	TDS	16 Plastrz	sont	6/10/10	1130
TA	Firrows Fe	16 Plastic	nenc	6/10/10	1130
TA	Ferriz Fe	500ml Plastic	H-NO3	6/10/10	1130
TH-	Sutficte	-250mt Placht	ZnAz	6- Ha-,	ven

Project Name: Colorado	school of	Min	13	Sample Location CSMRI - IB	
Project Number.	49-250		1	Date: 6/5-110.6/9/10	
Sample Type:	Duplicate	SW Other	ΕB	Sampler N Malezyt	

Purge Volume Calculat	Sample Collection					
Measured TD =	23.41 1) Analysis	Container	Preservative	Date	Time
	1+ 281	22-226	1901		<i>i i</i>	
Total Depth =	23.69 (11	Diss. U	cuse	HNO	6/9/10	0930
Depth to Water =	18.01 (ft	0	soome	3	, ,	
		Cotions	Plastiz	HNO-	6/9/10	0930
Initial Water Column =	(ft)	soome	1	, ,	
	5-68	Anions	Plastiz	-	619/10	0930
Initial Water Volume =	0.90 191		125ml		1 1	
		DOC	Amber	Hy SOM	6/9/10	0930
3 X Water Volume	2.70 11	Lah. ALS-	Ft. Collm	, Test,	America -	Areal

Time	Volume: (gal)	T⊶mperature (℃)"F)	eH (응다)	Conductivity (uS/cm)	DO (mg/L)	ORP (mV)	Turbidity (NTU)	Appearance
1:07	0.90	11.67	7.01	1740	14,00	162	131	reddish Lorown
109	1.80	10,91	6.95	1540	14.76	162	1000+	Lown
114	2.70	11,36	6.90	1420	15.26	58	1000+	4
				1.0			11.00	
	1			1				
	1						at the second se	
	-				and the second sec			
	and the second							
								Non
ne purge	ed	25 gel						and the second second

Comme	ents: Analysis	Contract	Preservetive	Dete	Time
	Diss. Mahls	500ml Plactor	HNOZ	6/9/10	6930
TA	NO2, NO2	16 Plastre	rene	6/9/10	0930
TA	TDS	16 Plostre	LONC	49/10	0930
A	- Ferros Ec	the Plastic	near	6-110-	nen
A	-Faccil Fe	500-6 Planton	1+202	6/ 110 NO	n
=A	Suthale	250ml Plachz	-ZAAZ	6 to M	im

Project Name: Colorado	School of	Min	15	Sample Location CSMRJ - Z	
Project Number 43	49-250			Date. 6/9/10	
Sample Type	(W) Dophcate	SW Other	Eß	Sampler N Malazyk-	

Purge Volume Calcula	tions	Sample Collection					
Measured TD =	95.12 (11)	Analysis	Container	Preservative	Date	Lime	
Total Depth -	(+:28)	Re-226 -228	1901		11		
	95.40 (11)	Diss. UL	cuse	HNOZ	6/10/10	1230	
Depth to Water =	56.85 (11)	Cations	Plastic	HNON	6/10/10	1230	
Initial Water Column =	38.55	and the second second	SOOML Plastiz	-	6/10/10	1230	
Initial Water Volume =	6-16 (gal)	DOC	125ml Amber	Hz SON	dio lis	1230	
3 X Water Volume	18.48 (g.il)	Lab Ars-	Ft. Colla	Test	America -		

Tune	Volume (gal)	Temperature	pH (50)	Conductivity (u5/cm)	DC) (mg/L)	ORP (mV)	Lurbidity (NTU)	Appearance
1022	3.08	15.24	6 84	900	7.68	140	1000+	boom
1031	6.16	14.45	6.80	900	9.01	152	1000+	1
1040	9.24	14.63	6.73	900	9.67	153	1000+	
1051	12.32	14.60	6.76	986	14.04	148	1000+	t
							12.5	
							1.1.1.1.1.1.1	
								NEM
Volume purge	12.0	1					·	

Comme	ents Analysis	Conterner	Preservative	Dete	Time
	Diss. Mahals	500nd Plastiz	HNO3	6/10/10	1230
TA	Noz, NOz	16 Plactre	reve	6/10/10	1230
TA	TDS	16 Plostre	work	6/10/10	1230
PA-	-Ferrow Fe	-H_ Plastic	nort	if trom	
TA	-Ferrit Fe	- 500mb Plastra	H-102	6/ 10 Nr.	
Tat	Sutfield	250006 Plackz	ZnAFE	61 tro ve	n

Project Name Colorado	school of	Min	< 5	Sample Location.	
Project Number 43	49-250			Date 6/8/10	
Sample Type	GW Duplicate	SW Other	EB	Sampler N Malezyt-	

Purge Volume Calcula	tions	Sample Collection					
Measured TD =	17.34 (1	1) Analysis	Container	Preservative	Date	Tîme	
Total Depth =		Ra-226 -228 Diss. U	cube	HNO	6/8/10	1245	
Depth to Water =	4.37	Cations	Plastic	HNO,	6/8/10	1245	
Initial Water Column =	13.25		Plactiz	-	6/8/10	1245	
Initial Water Volume =	2.12 (gal	Dar	125ml Anler	Hz SON	blolio	17.45	
3 X Water Volume	6.36 190	Lali ALS-	Ft. Colla	Test	America -	Arenda	

Time	Molume (gal)	Temperature (C)°F)	pH (SU)	Conductivity (((South))	(rug(L)	(mV)	Turbidity (NTU)	Appearance
1225	106	11.12	7.05	1030	14.32	128	571	born
1228	2.12	10.79	6.86	980	13.94	134	668	1
1231	3.18	11.05	6.76	950	13.14	140	848	
1233	4.24	10.70	6.74	970	13.41	143	1000+	
1235	5.30	10.78	6.73	590	13.28	146	1000+	
12.39	6.36	10.89	6.72	990	12.84	147	1000+	*
-								Arria
olume purge	di 65	sellons						

Comm	ents: Analysis	Contract	Preservative	Defe	Time
	Diss. Makly	500nd Plactiz	HNOZ	6/8/10	12.45
TA_	Noz, NOz	16 Plastre	rene	6/8/10	1245
TA	TDS	16 Plostre	NONE	6/8/10	1245
TA	Ferrors Fe	16 Plastic	NOAK	6/8/10	1245
TA	Ferriz Fe	500ml Plastic	14-NO2	6/8/10	1245
FA	Sulfide	250mL Plashz	ZnAz	6/8/10	1245

Project Name Colorado	school of	Min	< <u>s</u>	Sample Location
Project Number 43	49-250			Date 6/8/10
Sample Type-	GW Duplicate	: W Other	ΕB	Sampler N Malczyt-

Purge Volume Calculations			Sample Collection					
Measured TD =	10.48	(ft)	Analysis	Container	Preservative	Date	Time	
	(+ <u>2</u> 8)	15	6-226	1901				
Total Depth =	11.26	(11) D	TSS U	cuse	HND	6/2/10	1150	
Depth to Water =		(11)		soome	5			
	100	C	tions	Plastiz	HNOZ	6/8/10	1150	
Initial Water Column –		ft)		sooml.				
	6.58	1	Inions	Plastiz		6/8/10	1150	
Initial Water Volume =	1.05 14	at)		izsme				
1		D	DOC	Amber	Hy SOM	6/8/10	1.50	
3 X Water Volume	3,15 (4)	aliLa	1) AZS-	Ft. Collon	· Test A	America -		

June	Volume (gal)	Temperature	pH (SU)	Conductivity (uS/cm)	DO (mg/L)	ORP (my)	Turbidity (NTU)	Appearance
1140	1.05	13,63	7.04	/170	12.41	-11	440	brown
1142	2.10	13.35	6.86	1140	12.42	-1	540	1
1145	3.15	13,34	6.80	1130	12.19	15	387	V
								Non

ents. Analysis	Contract	Preservetive	Dete	Time
Diss. Mehls	500nh Plactiz		6/8/10	1150
Nez, NO3 -	16 Plastiz	reve	6/8/10	1150
TDS	16 Plastrz	LOAL	6/8/10	1150
Ferrors Fr	IL Plastic	near	6/8/10	1150
Ferriz Fe Sulfrda	250ml Plastic 250ml Plastic	HNO, ZnAz	6/8/10	1150
	Diss. Mehls Nozi Noz TDS Ferrors Fe Ferriz Fe	Diss. Mehls SDOrte Plastic Nort NO3 14 Plastic TDS 14 Plastic Ferrors Fe 14 Plastic Ferric Fe SOOML Plastic	Diss. Makels 500ml Plastic HN03 Noz, NO3 16 Plastic rene TDS 16 Plastic rene Ferrors Fe 16 Plastic none Ferrir Fe 500ml Plastic HN03	Diss. Mehls SDORL Plactic HNO3 6/8/10 Noz, NO3 IL Plastic rene 6/8/10 TDS IL Plastic sone 6/8/10 Ferrors Fe IL Plastic none 6/8/10 Ferrors Fe SODAL Plastic HNO3 6/8/10

Project Name	School of	- Min	15	Sample Location	
Project Number 43	49-250			Date 6/8/10	
Sample Type	GW Duplicate	SW Other	EB	Sumpler N Maleryt	

Purge Volume Calcula	tions	Sample Collection					
Measured TD =	29.94 (1) Analysis	Container	Preservative	Date	Tîmê	
	(+.28)	225-28	1901				
Total Depth =	30.22 Itt	Diss. U	cuse	HNO	6/0/10	0950	
Depth to Water	24.23 (11	and the second se	soome	3-	ang free		
	29.05	Cations	Plaster	HNO,	6/8/10	0950	
Inifial Water Column =	5.99 (11)		500ml				
		Anions	Plactiz	-	6/8/10	0950	
Indial Water Volume	0.96 (gal)		125ml				
1		DOC	Amber	Hr SON Fest	6/8/10	0950	
3 X Water Volume	Z.88 (gal)	Lab: Azs-	Ft. Collm	· Test	America	Arend	

Lune	Volume (gal)	Temperature	pH (SU)	Conductivity (0SFG01)	DC) (mg/L)	ORP (m∀)	Turbidity (NTU)	Appearance
0937	0.96	12.60	6.18	1020	11.93	150	1000+	brown
0539	1.92	11,97	6.38	990	13.34	155	1000+	1
0941	2.88	11.90	6.44	990	13.80	160	1000+	+
~		10000						
	1							
	1							
olume purge								208-4

Comm	ents. Analysis	Contrainer	Preservetive	Defe	Time
	Diss Mohals	500nd Plastic	HNOZ	6/8/10	0950
TA	No2, NO3_	16 Plactre	reve	6/8/10	0950
TA	TDS	16 Plostre	LOAL	6/8/10	0750
A	Firms Ec	16 Plantic	nonc	of tion	
FA	-FRACT FL	500ml Plack	1=202	6/ 10-	nen
ra	-Selferte	750ml Plachz	ZnAc	61-110-	nen

	546	

Project Name.	School of	Min	Sample Location CSMRI - 7B	
Project Number 43	49-250			6/7/10,6/10/10
Sample Type	CV> Duplicate	SW Other	EB	Sampler Maleryt

Purge Volume Calcula	ations	Sample C	Sample Collection					
Measured TD =	16.82	(ft) Analysis	Container	Preservative	Date	Time		
Total Depth =	(+ 28) 17.10	-228 Diss U	cuse	HNO	6/10/10	1430		
Depth to Water		(ft) Cotions	Soomt Plastic	HNOZ	6/-110	-		
Initial Water Column -	0.83	(11) Anions	Soomi Plastiz	-	61-110			
Initial Water Volume =	0.13 (0	al)	125ml	11. 10				
3 X Water Volume	0.39 (0)	al) Lab. ALS-	Amber Et colla	Hz SOU	61-1101 America -	A 1		

Time	Volume (gal)	Temperature	pH (SU)	Conductivity (uS/cm)	DQ (mgd.)	()PP (mV)	Turtadity (NTU)	Appearance
1404	0.13	1614	7.30	1290	13,04	120	Austanting	dear
~	0.26						7.2	
	0.29							
	1				14			
	1	1						
								Non
alume purge	-18 g							No

Comme	ents Analysis	Conterner	Preservetive	Dete	Time
	Diss Makels	500nd Plactiz	HNOZ	6/-/10	
TA	Noz, NOz	16 Plastre	rene	11-110	
TA	TDS	16 Plastre	SONE	4-110	-
T-A	- Frerows Fe	-H. Pleatic	none	it tro	10cm
TA	Front Fe	- 500 plustor	- 14 Ala	6/ 110	ven
r <u>A</u>	-SLIFFAR	-250mal Plachz	-ZhAZ	6/ 110-	NEM

Project Name Colorado	> School of	Min	15	Sample Location	
Project Number 43	49-250			Date 6/5/10	
Sample Type	Duplicate	SW Other	EB	Sampler N. Malarit	

Purge Volume Calculat	ions		Sample Collection					
Measured TD	17.03	(ft)	Analysis	Container	Preservative	Date	Tune	
	(+ 28)	- 13	Ra-226 -228	1901			-	
Total Depth =	17.31	(ft)	Diss. U	cuse	HND	6/8/10	1340	
Depth to Water =	5.71	$(\hat{\mathbf{f}})$		soome	5			
	3.71		Cations	Plastiz	HNOZ	6/8/10	1340	
Initial Water Column =		(11)		SOOML				
	11.60		Anions	Plastiz	· · · · ·	6/8/10	1340	
nifial Water Volume -	1.86	(al)		125ml	-			
			DOC	Amber	Hz SOW	6/2/10	1340	
3 X Water Volume	5.58 10	ral)	Lab: ALS-	Ft Calla	· Test	America -	Acerto	

Time	Volun⇔	Temperature	рН	Conductivity	F)(;)	ORP	Furbidit,	Appearance
	(gal)	$(\mathbb{C})^{n}\mathbb{F})$	(SU)	(uS/cm)	(mg/L)	()u/=)	([1111])	
1320	0.93	10.92	7.13	1240	13.80	134	122	cloudy
1323	1.86	11.32	7.02	1170	13,76	136	749	
1326	2.79	11.93	6.99	1150	13.31	137	1000+	
1329	3.72	12,42	6.98	1150	13.12	139	1000+	-
1322	4.65	12.62	6.99	1150	12.43	140	10007	
1335	5.58	12.93	6.99	1160	12.96	141	1000+	4
					1			
				1.				VEM
Volume purged	5.75							ALC 1

Comme	ents: Analysis	Contriner	Preservative	Dete	Time
	Diss Makels	500rd Plactiz	HNOZ	6/8/10	1340
TA	No2, NO2	16 Plastre	rene	6/8/10	1340
TA	TDS	16 Plastre	sone	4,8/10	134D
TA	Ferrors Fe	14 Plastic	near	6/8/10	1340
TA	Ferriz Fe	500ml Plastic	HNOZ	6/5/10	1340
TA	Sulfrale	zsom L Plashz	ZnAz	6/8/10	1340

Project Name Colorado	School of	Min	< s	Sample Location
Project Number	49-250			Date 6/9/10
Sample Type	GW	SW	Εß	Sampler
	Duplicate	Other_		N Maleryt

Purge Volume Calculat	tions	Sample C	ollection			
Measured TD =	33 . // (it (+.28)	Analysis	Container	Preservative	Date	Time
Total Depth =		-228 Diss. U	cuse	HNO	6/9/in	1100
Depth to Water	22.45 (H)	Cations	500ml Plastic	HNO.	6/9/10	1150
nihal Water Column =	10.94 ⁽¹¹⁾	Anions	Soome Plastiz	-	6/9/10	1150
Initial Water Volume =	1.75 (gal)		125ml		1-1	
3 X Water Volume	5.25 (gal)	Lab: ALS-	Ft. Colla	Hz SOU	6/9/10 -	1150 A.

Tune	Voltume (gal)	Temperature (O'F)	pH (SU)	Conductivity (uS/cm)	()() (mg/L)	QRP (mV)	Lurbidity (MTU)	Appearance
1132	0.88	13.15	6.97	1600	9.82	149	10007	brown
1.35	1.75	12.20	6.70	1520	9.07	153		1
1138	2.63	11.87	6.70	1470	9.09	156		
1141	3.50	12.34	6.65	1420	9.46	159		
144	4.38	12.16	6.64	1400	10.30	167		
1146	5.25	12.55	6.66	1380	10.36	165	-	4
			_		4404686 denete		1	
Volume purge								Nen

Comm	ents Analysis	Contrainer	Preservetive	Dete	Time
	Diss Mahls	500mb Plactiz	HNOZ	6/9/10	1150
TA	Nez, Noz	16 Plastre	rene	6/9/10	1150
TA	TDS	16 Plastrz	sont	19/10	1150
T.A.	- Ferrors Fe	-H Plestic	- hoar	1/ 40 00	1150 m
TA	-FRACE FL	500ml Plantiz	IFNO.	6/ 110 m	
TA	-Suttende	250ml Plaster	ZnAr	61 tron	n

Project Name: Colorado	School of	Min	Sample Location		
Project Number 43	49-250			Date 6/9/10	
Sample Type	GVV Duplicate	SVV Other	EB	Sampler N Malezy 5-	

Purge Volume Calcula	ntions	Sample C	Sample Collection					
Measured TD =	27.86 (ft (+ 28)	20-226	Container	Preservative	Date	Time		
Total Depth	28.14 (ft	-228 Diss. U	cuse	HNOS	6/9/10	1730		
Depth to Water -	23.18 ^{(ft}		500ml Plastiz	HNOZ	6/9/10	1230		
Initial Water Column =	4.96 ^{(ft}		SOOML Plastiz	-	6/9/10	1230		
Initial Water Volume	0.80 (gal	DOC	125ml Amber	H 50.	6/9/10	1230		
3 X Water Volume	2.40 (gal)	Lab: ALS-	Ft. Colla	Test	America -			

Time	Volume (gal)	Temperature OF)	pH (SU)	Conductivity (u\$/cm)	DO (mgil.)	ORP (mV)	Forbidity (PFU)	Appearance
1214	0.80	14.44	7.22	1190	11.31	161	55.1	born
1217	1.60	12.67	7.04	1120	11.65	161	882	1
1726	2.40	12.36	6. 91	1110	11.38	16Z	724	47
								Nik

Comme	ents Analysis	Container	Preservative	Dete	Time
	Diss. Mahls	500rd Plactic	H-NO2	6/9/10	1230
TA	Noz, NOz	16 Plastre	rene	6/ 9/10	12.30
TA	TDS	16 Plastre	sont	69/10	1230
A	- Ferras Fe	- H_ Plastic	MONC	of to a	en
TA	FACTIR FL	500ml Plastic	HAD2	6/ 110 N	
r <u>A</u>	Sattrate	250ml Plache	ZnRc	61 tro N	en

Project Name Colomolo	School of	Min	Sample Location		
Project Number 43				Date 6/8/10	
Sample Type	GW Duplicate	SW Other	EB	Sampler N Malczyk-	

Purge Volume Calcula	tions		Sample Collection					
Measured TD =	28.55	(ft)	Analysis Re-226	Container	Preservative	Date	Tume	
Total Depth =	(+ 28) 28.83	(ft)	-228 Diss. U	cuse	HNO	6/8/10	1040	
Depth to Water =	26.41	(ft)	Cations	500ml Plastiz	HNON	6/8/10	1040	
Initial Water Column	2.42	(ft)	Anions	500ml Plastiz	-	6/ 110	1040	
Initial Water Volume =	0.38	gal)		125ml	H. SON	itali		
3 X Water Volume	1.14	gali	DOC Lab: Azs-	Amber Ft. Collm	Ht SUN	America -	Armla	

Time-	Volume	Temperature	рН	Conductivity	DO	ORP	Turbidity	Appearance
	(gal)	(°C)"F)	(SU)	(uS/cm)	(mg/L)	(mV)	(NTU)	1
1027	0.38	12.79	6,94	1030	16.25	163	1000T	branish
1029	0.76	12.60	6.89	1050	16.05	163	1000t	1
1032	1.14	12.51	6.80	1000	16.67	165	1000+	4
~								lie-
	1.00		······································					
				and the second second second	*****			
					1.			
								NEM

Comme	ents: Analysis	Container	Preservetive	Defe	Time
	Diss. Mohls	500nd Plactor	HNOZ	6/8/10	1040
TA	Noz, NO3	16 Plaster	rent	6/8/10	1040
TA	TDS	16 Plostre	SONC	6/8/10	1040
TA	-Farros Es-	11 Pleatic	- DOAK	4 110 m	in
TA	-Ferrit Fe	500ml Plack	11-100	-6/ HO M	in
TA	-Setforte-	250001 Plaster	ZnAZ	6 tron	2

Project Numbe	4349	school of	Mines		0				
					Date				
	4544	1-1							
Santhe Type			Tet		6/9/1	0			
		GW		ŧΒ	Sampler	1	1		
		Duplicate	Other		N	Malery	15		
Purge Volui	me Calc	ulations		Sample C	ollection				
Measured TD =		1	i)A		Container	10	1	1	
			(+ 28)		1901	Preservative	Date	Tune	
Total Depth =			/	Re-226 -228	1901		. In I	1400	
Depth to Water				Diss. U	CLSC	HNO3	6/7/10	700	
copinitio viater	7		(ft)	1			11	1.5261	
Initial Water Col	1		1	Cations	Plastoz Soome	HNOZ	6/9/10	1400	
miniar water Col	ionu) =	1.	(ft) ~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~						
nitial Water Vol		1-1-1		Anions	Plactiz 125ml		6/9/10	1400	
undar water vol	une		(141)				1.1	1	
2 VIN-I-I		1		DOC	Amber	Ho SON	6/9/10	1400	
3 X Water Volun	ile.	1	(gal)	Lan Mas-	Ft. Collin	s, Test,	America	- Actor	
Time	Volume (gal)	Temperature	p14 (SU)	Conductivity (uS.cm)	DQ (mg/L)	ORP (mV)	Turbidity (NTU)	Appearai	
1252	V/A	12.41	6.97	100	13.09	173		1	
the second the					The second se	114	90.7	brown	
						1.1.4	70.1	brown	
							90.7	brown	
							70.7	brown	
							70. 1	brown	
							70.7	brown	
							70.1	brown	
							70.7	barn	
							70-1	Ven	

Comme	ents: Analysis	Contriner	Preservetive	Dete	Time
	Diss Mohls	500nh Plastiz	HNO,	6/9/10	1400
A	Noz, NOz	16 Plasta	rene	6/9/10	1400
A	TDS	16 Plastre	LONK	6/9/10	1400
4	From Er	-H- Plestic	-ponc	of lion	
4	-Farrie Fe	500ml Plasta	12-ACO-	6/ 110 m	
A	-selfide	-250ml Plache	-ZRAZ	6/ ten	rem

Project Name	School of Mines	Sample Location.
Project Number 43	49-250	Date. 6/9/10
Sample Type	GW SW EB Duplicate Other	Sampler N Maletyt

Purge Volume Calc	ulations	Sample C	ollection			
Measured TD =	(A)	Analysis	Container	Preservative	Date	Time
Total Depth =) + 28) (ft)	Re-226 -228 Diss. U	1gel cuse	HNO	6/9/10	1245
Depth to Water =	(ft)	Cations	500ml Plastic	HNON	6/4/10	1245
Initial Water Column =	(it) ven		500ml Plactiz		6/9/10	1245
Initial Water Volume =	(gal)	DOC	125ml Amber	Hy SOM	6/4/10	1245
3 X Water Volume	(gal)	Lab: Mrs-	Ft. Colla	· Test	America -	Arendo

Time	Volume (gal)	Temperature	pH (SU)	Conductivity (uS/cm)	DO (mg/L)	ORP (m\/)	Turbidily (NTU)	Appearance
1240	1 mm	11.67	7.68	107	13.99	156	171	brown
								ven.

Time
1245
1245
1245
m
1
n

1.01	ne salo s	icheol of	Ma		Sample Loc			
Project Nur	nber.	cheel of	2 Cine	\$	Date	2		
		-250				10		
Sample Typ	1371 DR		AND	FB	6/10/1 Sampler	0		
		Duplicate	Other	LD			1	
		copilitate	ound		N	Malez	x5	
Purge Vo	olume Calc	ulations		Sample C	Collection			
Measured T	D -		(ft) Analysis	Container	Preservativo	Date	Time
			(+28) /	11	1		2013 Th	and the second s
Total Depth	8		1	-228	144	410	1 list.	13115
Depth to Wa	iter =		1 1	1)	soont	11-003	0110/10	1.070
		1	/	1 time	Plactor	UNA	the los	12110
Initial Water	Column	1	(f	t)	Soome	3-	0110 110	1340
		1/1	rn	Anions	Plactiz	-	6/10/11	1340
Initial Water	Volume =	1 /	(ga	1)	125ml			1
					the second se			10.1
				DOC	Amber	H. SO.	6/10/10	1340
3 X Water Vo	olume		(ga	DOC Lab: ALS-	Amber Ft. Collm	Hz SON	6/10/10 America	- Acro A
		/	(ga	1) Cotions 1) Cotions 1) Cotions 1) Cotions 1) DOC 1) DOC	PAmber Pt. Collm	Hz SOy s, Test.	6/10/10 America	- Arm
Purge Vol		Field Water	(ga Quality M	DOC Lab: Azs-	Amber Ft. Collm	Hz SOy s, Test.	6/10/10 America	- Arm
		Temperature	(gal Quality N pH	DOC Lab. Ats-	Amber Fl. Collin ts	Hz SON S, Test.	6/10/10 America	
Purge Vol	lumes and Volume (gal)		Quality N	leasuremen	ts		Turbidity	r
Purge Vol	umes and Volume	Temperature	Quality N pH (SU)	Conductivity (uS/cm)	DO (mg/L)	ORP (m섯)	Turbidity (NTU)	Appearanc
Purge Vol Tīma	lumes and Volume (gal)	Temperature (°C, °F)	Quality N pH (SU)	Conductivity	ts DO	ORP	Turbidity	Appearanc
Purge Vol Time	lumes and Volume (gal)	Temperature (°C, °F)	Quality N pH (SU)	Conductivity (uS/cm)	DO (mg/L)	ORP (m섯)	Turbidity (NTU)	Appearanc
Purge Vol Time	lumes and Volume (gal)	Temperature (°C, °F)	Quality N pH (SU)	Conductivity (uS/cm)	DO (mg/L)	ORP (m섯)	Turbidity (NTU)	Appearanc
Purge Vol Tīma	lumes and Volume (gal)	Temperature (°C, °F)	Quality N pH (SU)	Conductivity (uS/cm)	DO (mg/L)	ORP (m섯)	Turbidity (NTU)	Appearanc
Purge Vol Tīma	lumes and Volume (gal)	Temperature (°C, °F)	Quality N pH (SU)	Conductivity (uS/cm)	DO (mg/L)	ORP (m섯)	Turbidity (NTU)	Appearanc
Purge Vol Tīma	lumes and Volume (gal)	Temperature (°C, °F)	Quality N pH (SU)	Conductivity (uS/cm)	DO (mg/L)	ORP (m섯)	Turbidity (NTU)	Appearanc
Purge Vol Tīma	lumes and Volume (gal)	Temperature (°C, °F)	Quality N pH (SU)	Conductivity (uS/cm)	DO (mg/L)	ORP (m섯)	Turbidity (NTU)	Appearanc
Purge Vol Tīma	lumes and Volume (gal)	Temperature (°C, °F)	Quality N pH (SU)	Conductivity (uS/cm)	DO (mg/L)	ORP (m섯)	Turbidity (NTU)	Appearanc
Time	Volume (gal) N/A	Temperature (°C, °F)	Quality N pH (SU)	Conductivity (uS/cm)	DO (mg/L)	ORP (m섯)	Turbidity (NTU)	Appearance Low

	Inclusis	Conterner	Presecutive	Dete	Time
	TSS. Mohly	500nd Plactiz	HNOZ	6/10/10	1340
TA 1	102, 103 _	If Plactor	rene	6/10/10	1340
TA T	DS	16 Plastre	sone	6/10/10	1340
	Trous Fr	It Plastic	none	6/ 110	12M
	The Fe	- 500ml Plantin_	1+202		ven
-ASc	Hidt	-250mt Plashe	ZnAc	6/ 170.	NEM

Project Name: Colorado	School of	Mine	Sample Location. Equipment Blank
Project Number	49-250		Date 6/10 /10
Sample Type	GW	SW	EB Sampler
	Duplicate	Other	N Maleryt

Purge Volume Calci	ulations	Sample C	ollection		1	
Measured TD =	jr.) Analysis	Container	Preservative	Date	Tune
Total Depth =	(+.28)	Re-226 -228 Diss. U	1gel cuse	HNO	6/10/10	1040
Depth to Water =	(fl)	the second second second second	500ml Plastiz	HNO-	6/10/10	1040
Initial Water Column -	(ft)		SOOML Plactiz	-	6/10/10	1040
Initial Water Volume =	(gal)		125ml Amber	H. SON	6/10/10	1040
3 X Water Volume	(gal)	Lab. Ars-	Ft. Collm	s. Test	America -	Arende

Time	Volume (gal)	Temperature (°C, °F)	pH (SU)	Conductivity (uS/cm)	DQ (mg/L)	QRP (mV)	Turbidity (NTU)	Appearance
1								
		and the second s						
			and the second	Non Anna				
						X		
-								win

Comme	ents: Analysis	Contriner	Preservetive	Dete	Time
	Diss Makels	500nh Plactiz	HNO2	6/10/10	1040
TA	Noz, NOz	16 Plactor	rene	6/10/10	1040
TA	TDS	16 Plastic	LOAL	6/10/10	1040
FA	-Ferrors Fe	It Plastic	none	of 100	ven
TA	Farrithe	SOOML Plashi	1+203	6/ 110 -	ven
TA	Sulfalt	250ml Plashi	-ZnAz	6/ 110 /	ven

Appendix C Surface Water Sampling Procedures

Surface Water Sampling

1.0 Scope and Objective

1.1 Scope

This procedure provides instructions and establishes requirements for the collection and documentation of surface water samples by Stoller personnel. This procedure applies to the collection of surface water samples from streams, rivers, ponds, lakes, seeps, impoundments, and other surface sources.

1.2 Objective

The objective of this procedure is to establish a uniform method for the collection of surface water samples that provides representative samples in a safe and responsible manner.

2.0 Definitions

Composite Sample – A sample that is comprised of roughly equal amounts of water collected from a set of sample locations known as a sample group.

Grab Sample – A single sample collected at one sampling point over a short period of time. Grab sample results are representative of the sample location at the time of sample collection. Also called a catch sample.

Peristaltic Pump – A self-priming, low volume pump consisting of a rotor and ball bearing rollers. Tubing placed around the rotors is squeezed by the rotors as they revolve. The squeezing produces a wavelike contractual movement which causes water to be drawn through the tubing. The peristaltic pump is limited to sampling at depths of less than 25 feet.

3.0 Responsibilities and Qualifications

3.1 Project Manager

The Project Manager is responsible for ensuring that surface water samples are properly and safely collected. This will be accomplished through staff training and by maintaining quality control (QC). At a minimum, project management shall:

- 3.1.1 Verify that personnel have reviewed, and are familiar with, site-specific work plans which address surface water sampling, this procedure, and any associated procedures.
- 3.1.2 Ensure that hazards are identified and analyzed with respect to collecting surface water samples, and develop and implement controls to minimize hazards.
- 3.1.3 Provide personnel with training in the operation of surface water sampling equipment and the requirements of this procedure.
- 3.1.4 Periodically review field generated documentation associated with surface water sampling to ensure compliance with project requirements and implement corrective action if necessary.
- 3.1.5 Receive feedback from field sampling personnel in order to continually improve surface water sampling process.

3.2 Site Supervisor

The Site Supervisor is responsible for directing and overseeing all field activities, including sampling, to ensure that site-specific plan requirements are met in a safe and efficient manner within the established safety envelope.

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3.3 Field Sampling Personnel

Field sampling personnel are responsible for the proper sample collection and documentation of the sampling event in accordance with this procedure. At a minimum, field sampling personnel have the responsibility to:

- 3.3.1 Familiarize themselves with site-specific work plans, surface water sampling procedures, potential hazards, and health and safety plan.
- 3.3.2 Implement the controls to minimize hazards.
- 3.3.3 Be familiar with sampling equipment and its proper use.
- 3.3.4 Properly complete field documentation.
- 3.3.5 Provide feedback to project manager in order to improve sampling process.

4.0 Equipment/Materials and Calibration

4.1 Equipment/Materials

A number of devices are available for the collection of surface water samples. These devices are constructed of a number of materials including, but not limited to: stainless steel, glass, Teflon*, Tygon*. The sampling and analytical requirements, as well as site characteristics, must be taken into account when determining the proper surface water sampling equipment to use. The site-specific work plans should identify the specific equipment to be used, and methods for safely using equipment.

4.2 Calibration

Equipment shall be calibrated in accordance with manufacturer's recommendations and calibration documentation shall be maintained in project files.

5.0 Method

5.1 Field Preparation

Field preparation requires the organization of sample containers, sample labels, and documentation in an orderly, systematic manner to promote consistency and traceability of all data.

- 5.1.1 General sampling areas will be predetermined to ensure coverage of the various impact scenarios and should be described in project-specific work plans. The location of each sampling point shall be surveyed or mapped and staked as described in Section 5.1.6 prior to sampling.
- 5.1.2 In flowing water, surface water sampling shall be conducted from downstream locations first, then proceed to upstream locations to avoid potential cross contamination from disturbing the substrate.
- 5.1.3 Prior to sampling and between sampling locations, sampling equipment shall be decontaminated.
- 5.1.4 Appropriate personal protective equipment shall be used, as specified in the project-specific health and safety plan.
- 5.1.5 All pertinent information (date, site name, identification number, and location) shall be recorded on a Field Activity Daily Log (FADL) and a Sample Collection Log, as appropriate. Field conditions, unusual circumstances, and weather conditions shall be noted.

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- 5.1.6 Due to the nature of sampling an aqueous environment, additional steps are required to verify and mark sample locations. Depending on the project needs, it may be useful to use a Global Positioning System (GPS) to verify and mark the sample locations. Refer to *Field Mapping with a Global Positioning System* for details. The following steps shall be followed by the sampler in addition to the field preparation requirements described in Section 5.1.1.
 - 5.1.6.1 Place a marker (stake) on the shore approximately perpendicular to the sampling location and mark the sample number on the stake.
 - 5.1.6.2 If the sample location is accessible by foot, use a measuring tape to measure the distance between the marked point and the sample location station. Record the compass bearing from the sample location to the shore marker.
 - 5.1.6.3 If the sample location is accessible only by boat, use a rangefinder to estimate the distance to the shore marker to obtain the most accurate measurement. Record the compass bearing from the sample location to the shore marker. It is recommended that the boat's position on the water be stabilized to prevent drifting.
 - 5.1.6.4 Determine and record the distance and direction of each shore marker from a reference point shown on the topographic map and mark all points on a map or use a GPS, if available.
- 5.1.7 Quality Control samples, including field and source blanks, shall be collected in accordance with the project-specific work plan.

5.2 Surface Water Sample Collection Using a Transfer Container

The device most commonly used to collect grab surface water samples is a transfer container (beaker, flask, etc.) made of inert material such as glass, stainless steel or Teflon*. When sampling with a transfer container, the procedure is as follows:

- 5.2.1 Survey and clearly map sampling points as described in Section 5.1.6 prior to sampling. The sample should be collected as close to the mapped location as possible. If the collection point must be moved, the new location must be approved and documented.
- 5.2.2 Dip the transfer container into the surface water. Always use a clean, properly decontaminated transfer container at each sample location.
- 5.2.3 Filter the sample if required.
- 5.2.4 Fill the sample bottle, allowing the sample stream to flow gently down the inside of the bottle with minimal turbulence.
- 5.2.5 Cap the bottle and handle the sample according to the procedures outlined in Project *Sample Shipping*.
- 5.2.6 Label the sample and document the sampling event.

5.3 Surface Water Sample Collection Using a Peristaltic Pump

A device used to collect composite surface water samples is a peristaltic pump. Samples to be analyzed for volatile organic analysis cannot be composited. When sampling with a peristaltic pump, the procedure is as follows:

5.3.1 Survey and clearly map sampling points as described in Section 5.1.6 prior to sampling. The sample should be collected as close to the mapped location as possible. If a collection point must be moved, the new location must be approved and documented.

- 5.3.2 Attach the appropriate tubing to the peristaltic pump. Always use new tubing at each sample location. Do not try to decontaminate and reuse tubing.
- 5.3.3 If filtering is required, attach the filtering device to the discharge end of the tubing.
- 5.3.4 Lower the intake end of the tubing into the water and begin pumping. If the pump is computerized, program the pump to collect the sample at the desired intervals and flow rate. If the pump is not programmable, record the discharge rate (compute discharge rate by dividing an amount of water collected by the time it took to collect it). Collect the sample at the desired interval.
- 5.3.5 Fill the sample bottle, allowing the sample stream to flow gently down the inside of the bottle with minimal turbulence. The programmable pump will perform this automatically.
- 5.3.6 Cap the bottle and handle the sample according to the procedures outlined in Project *Sample and Shipping.*
- 5.3.7 Label the sample and document the sampling event.

6.0 Required Inspection/Acceptance Criteria

None.

7.0 Records

The following records generated as a result of implementation of this procedure shall be maintained in a safe manner and submitted to project central files for storage and disposition.

Field Activity Daily Log

Sample Collection Log

Chain of Custody

8.0 References

8.1 Others

- U.S. Environmental Protection Agency. 1987. EPA Compendium of Superfund Field Operations Methods, EPA 540/P-87/001a, OSWER 9355.0-14. Washington, DC.
- U.S. Environmental Protection Agency. 1988. EPA Guidance for Conducting Remedial Investigation and Feasibility Studies under CERCLA, Interim Final OSWER Directive 9355.3-01. Washington, DC.
- American Public Health Association, American Water Works Association, Water Pollution Control Federation. 1985. Standard Methods for the Examination of Water and Wastewater, 16th Edition, American Public Health Association, Washington, DC.

DATA VALIDATION REPORT

To:Robert HillFrom:John GarrettDate:July 29, 2010Project/Site:Colorado School of MinesProject No.:4060SDG No.:1006125

This report presents the inorganic anions data validation for the data obtained for sixteen CSMRI water sample collected on June 08, 2010, June 09, 2010, and June 10, 2010 and submitted to ALS Laboratory Group on June 11, 2010 for the above referenced work assignment. The purpose of this review is to provide a technical evaluation of the inorganic anions results that were obtained by preparation method MCAWW, May 1994, and EMSL Rev 2.1 Alkalinity, Bicarbonate, and Carbonate by Method 310.1 ALS SOP 1106R8, Sulfate, Fluoride, Chloride, Nitrate as N, and Nitrite as N by Method 300.0 Rev 2.1 ALS SOP 1113R11, Total Dissolved Solids (TDS) by Method 160.1 ALS SOP 1101R10 from ALS Laboratory Group. (Fort Collins, CO). The water samples were analyzed for Bicarbonate, Carbonate, and Total Alkalinity, on June 21, 2010, TDS on June 17, 2010, and Sulfate, Fluoride, Chloride, Nitrate as N, and Nitrite as N on June 15, 2010, and June 17, 2010. All analyses were conducted by ALS Laboratory Group. The field sample numbers and corresponding laboratory numbers are presented below:

Client Sample Number	Laboratory Sample Number	Matrix	Collection Date
CSMRI-1	1006125-1	Water	June 10, 2010
CSMRI-1B	1006125-2	Water	June 09, 2010
CSMRI-2	1006125-3	Water	June 10, 2010
CSMRI-4	1006125-4	Water	June 08, 2010
CSMRI-5	1006125-5	Water	June 08, 2010
CSMRI-6C	1006125-6	Water	June 08, 2010
CSMRI-7B	1006125-7	Water	June 10, 2010
CSMRI-8	1006125-8	Water	June 08, 2010
CSMRI-9	1006125-9	Water	June 09, 2010
CSMRI-10	1006125-10	Water	June 09, 2010
CSMRI-11B	1006125-11	Water	June 08, 2010
SW-1	1006125-12	Water	June 09, 2010
SW-2	1006125-13	Water	June 09, 2010
SW-3	1006125-14	Water	June 10, 2010
EQUIPMENT BLANK	1006125-15	Water	June 10, 2010
West Seep	1006125-16	Water	June 10, 2010

Data validation was conducted in accordance with the USEPA Contract Laboratory Program National Functional Guidelines for Inorganic Data Review.

The Inorganic data were evaluated based on the following parameters:

- * Data Completeness
- * Holding Times and Preservation
- * Initial and Continuing Calibration Verification
- * Contract Required Detection Limit (CRDL)
- * Preparation/ Initial (ICB)/ and Continuing (CCB) Calibration Blanks
- * Interference Check Sample (ICSA) Results
- * Matrix Spike Results
- * Duplicate Sample Results
- * Laboratory Control Samples (LCS) Results
- * Serial Dilution Sample Results
- * Compound Quantitation and Reporting Limits (full validation only)
- * All criteria were met for this parameter

Data Completeness

The data package was complete. No results were qualified as a result of the missing data.

Holding Times and Preservation

Analytical holding times were evaluated and all criteria were met with the following exception:

Nitrate as N, and Nitrite as N which were analyzed over 2X the holding time in sample West Seep. The sample reported detects for both Analyte and are qualified as Estimated [J].

The water samples were all found to be field filtered and had a pH less than 2. Cooler #1 was received at a temperature of 5°C. However there is no documentation as to which samples were in the cooler. This should be taken into account when determining the usability of the data for all analyses except for certain radiochemistry analyses.

Initial and Continuing Calibration Verification

Initial and Continuing Calibration Verification standards were analyzed at the required frequency and all were within the required 90-110%. No action was necessary.

Contract Required Detection Limit (CRDL)

All CRDL %Rs CRI %Rs were within 80-120% limits. No action was necessary.

Preparation and Initial/ Continuing Calibration Blanks

Preparation and Initial/Continuing Calibration Blank analyses were performed at the required frequency. Preparation and Initial/ Continuing Calibration Blanks are evaluated to assess the level of contamination in the preparation and analytical processes.

Preparation and Initial/ Continuing Calibration Blanks were prepared and analyzed at the required frequencies.

All of the blanks that were analyzed had concentrations that were below their respective Reporting Limits (RLs).

However, if blank results were above the Instrument Detection Limits (IDLs) and below the RLs, it caused the associated sample results to be qualified for contamination as estimated and non-detected [UJ 107]. If blank results were below the negate IDL and above the negate RL, it caused the associated sample results to be qualified for negative contamination as estimated [J 107]. No sample results were qualified due to blank contamination.

Matrix Spike/Matrix Spike Duplicate Results

MS/MSD analyses were performed at the required frequency. All MS/MSD percent recoveries were within 75-125% limits with the following exception:

The chloride concentration was above the analytical range in the native sample CSMRI-1 and MS/MSD recoveries could not be evaluated. The associated LCS, ICV, and CCV results were within control limits and no action was necessary.

Duplicate Sample Analysis

Duplicate analyses were performed at the required frequency. All original sample/duplicate sample and MS/MSD differences were less than 20% RPD or less than the RDL for results less than (5)(RDL). No actions were necessary.

Laboratory Control Samples

LCS analyses were performed at the required frequency. The laboratory analyzed laboratory control samples for all analytes. All recoveries were within 80-120% limits. No action was necessary.

Serial Dilution Results

All %Ds were less than 10% for all analytes.

Analyte Quantitation and Reporting Limits

Analyte quantitation was evaluated for all samples. No calculation or transcription errors were found. The results and reporting limits were correctly reported.

Overall Comments

Nitrate as N, and Nitrite as N which were analyzed over 2X the holding time in sample West Seep. The sample reported detects for both analytes and are qualified as Estimated [J].

The chloride concentration was above the analytical range in the native sample CSMRI-1 and MS/MSD recoveries could not be evaluated. The associated LCS, ICV, and CCV results were within control limits and no action was necessary.

The water samples were all found to be field filtered and had a pH less than 2. Cooler #1 was received at a temperature of 5°C. However there is no documentation as to which samples were in the cooler. This should be taken into account when determining the usability of the data for all analyses except for certain radiochemistry analyses.

Reduced aliquots were analyzed for samples CSMRI-1, CSMRI-1B, CSMRI-2, CSMRI-4, CSMRI-5, CSMRI-6C, CSMRI-8, CSMRI-9, CSMRI-10, CSMRI-11B and West Seep for chloride, sulfate, alkalinity, bicarbonate, and carbonate. The laboratory elevated the reporting limits accordingly.

DATA QUALIFIER DEFINITIONS

For the purpose of Data Validation, the following code letters and associated definitions are provided for use by the data validator to summarize the data quality.

- R Reported value is "rejected." Resampling or reanalysis may be necessary to verify the presence or absence of the compound.
- J The associated numerical value is an estimated quantity because the Quality Control criteria were not met.
- U J The reported quantitation limit is estimated because Quality Control criteria were not met. Element or compound was not detected.
- The material was analyzed for, but was not detected above the level of the associated value. The associated value is either the sample quantitation limit or the sample detection limit.
- NR Result was not used from a particular sample analysis. This typically occurs when more than one result for an element is reported due to dilutions and reanalyses.

DATA VALIDATION REPORT

To:Robert HillFrom:John GarrettDate:July 28, 2010, 2010Project/Site:Colorado School of MinesProject No.:4060SDG No.:1006125

This report presents the inorganic metals data validation for the data obtained for fourteen dissolved metals, 16 dissolved Uranium, 15 dissolved cations, 14 dissolved mercury for the CSMRI water samples collected on June 08, 2010, June 09, 2010, and June 10, 2010 and submitted to ALS Laboratory Group on June 11, 2010 for the above referenced work assignment. The purpose of this review is to provide a technical evaluation of the inorganic metals results that were obtained by SW-846, 3rd edition, Method 6010B and ALS Laboratory Group SOP 834R7 for trace metals by Inductively Coupled Plasma (ICP) atomic emission spectrometry analysis, Method 6020A ALS Laboratory Group Procedure SOP 827R7 for dissolved metals by Inductively Coupled Plasma mass spectrometry (ICP-MS) (Uranium only) analysis, Method 7470A ALS Laboratory Group Procedure SOP 812R14 Cold Vapor Atomic Absorption (CVAA) (mercury) for SDG 1006125 from ALS Laboratory Group (Fort Collins, CO). The water samples were analyzed for dissolved ICP trace metals, cations, and on July 06, 2010, dissolved uranium by ICP-MS on July 06, 2010, and dissolved mercury by CVAA on July 2, 2010. All analyses were conducted by ALS Laboratory Group. The field sample numbers and corresponding laboratory numbers are presented below:

Client Sample Number	Laboratory Sample Number	Matrix	Collection Date
CSMRI-1	1006125-1	Water	June 10, 2010
CSMRI-1B	1006125-2	Water	June 09, 2010
CSMRI-2	1006125-3	Water	June 10, 2010
CSMRI-4	1006125-4	Water	June 08, 2010
CSMRI-5	1006125-5	Water	June 08, 2010
CSMRI-6C	1006125-6	Water	June 08, 2010
CSMRI-7B	1006125-7	Water	June 10, 2010
CSMRI-8	1006125-8	Water	June 08, 2010
CSMRI-9	1006125-9	Water	June 09, 2010
CSMRI-10	1006125-10	Water	June 09, 2010
CSMRI-11B	1006125-11	Water	June 08, 2010
SW-1	1006125-12	Water	June 09, 2010
SW-2	1006125-13	Water	June 09, 2010
SW-3	1006125-14	Water	June 10, 2010

EQUIPMENT BLANK	1006125-15	Water	June 10, 2010
West Seep	1006125-16	Water	June 10, 2010

Data validation was conducted in accordance with the USEPA Contract Laboratory Program National Functional Guidelines for Inorganic Data Review.

The metals data were evaluated based on the following parameters:

- * Data Completeness
- * Holding Times and Preservation
- * Initial and Continuing Calibration Verification
- * Contract Required Detection Limit (CRDL) Preparation/ Initial (ICB)/ and Continuing (CCB) Calibration Blanks
- * Interference Check Sample (ICSA) Results
- * Matrix Spike Results
- * Duplicate Sample Results
- * Laboratory Control Samples (LCS) Results
- Serial Dilution Sample Results
- * Compound Quantitation and Reporting Limits (full validation only)
- * All criteria were met for this parameter

Data Completeness

The data package was complete except for the missing CRDL (2B) and IDL (10) QC Summary Forms. No results were qualified as a result of the missing data.

Holding Times and Preservation

Analytical holding times were evaluated and all criteria were met.

The water samples were all found to be field filtered and had a pH less than 2. Cooler #1 was received at a temperature of 5°C. However there is no documentation as to which samples were in the cooler. This should be taken into account when determining the usability of the data for all analyses except for certain radiochemistry analyses.

Initial and Continuing Calibration Verification

Initial and Continuing Calibration Verification standards were analyzed at the required frequency and all were within the required 90-110% limits for ICP trace. No action was necessary.

Contract Required Detection Limit (CRDL)

No CRDL or CRI standard recovery summary forms (EPA Form 2b) were included in the data package. The reviewer obtained the %Rs from the instrument raw data. All CRDL %Rs for ICP and CRI %Rs for mercury were within 80-120% limits. No action was necessary.

Preparation and Initial/ Continuing Calibration Blanks

Preparation and Initial/Continuing Calibration Blank analyses were performed at the required frequency. Preparation and Initial/ Continuing Calibration Blanks are evaluated to assess the level of contamination in the preparation and analytical processes.

Preparation and Initial/ Continuing Calibration Blanks were prepared and analyzed at the required frequencies.

All of the blanks that were analyzed had concentrations that were below their respective Reporting Limits (RLs).

However, if blank results were above the Instrument Detection Limits (IDLs) and below

the RLs, it caused the associated sample results to be qualified for contamination as estimated and non-detected [UJ 107]. If blank results were below the negate IDL and above the negate RL, it caused the associated sample results to be qualified for negative contamination as estimated [J 107]. No sample results were qualified due to blank contamination.

Interference Check Sample (ICSA) Results

Interference Check Samples were prepared and analyzed at the required frequencies.

No aqueous concentrations of aluminum, calcium, iron, or magnesium exceeded the ICSA values in any of the samples. No action was necessary.

Matrix Spike/Matrix Spike Duplicate Results

MS/MSD analyses were performed at the required frequency. All ICP, ICP-MS and Mercury MS/MSD percent recoveries were within 75-125% limits. No action was necessary.

A Uranium MS/MSD was not evaluated for sample West Seep due to the concentration of Uranium in the sample was greater than 4X the concentration of the spike added during digestion. The associated LCS was in control and the data are not affected.

Duplicate Sample Analysis

Duplicate analyses were performed at the required frequency. All ICP, ICP-MS and CVAA original sample/duplicate sample and MS/MSD differences were less than 20% RPD or less than the RDL for results less than (5)(RDL). No actions were necessary.

Laboratory Control Samples

LCS analyses were performed at the required frequency. The laboratory analyzed laboratory control samples for all metals. All recoveries were within 80-120% limits. No action was necessary.

Serial Dilution Results

All %Ds were less than 10% for Uranium by ICP-MS and trace metals by ICP with the following exception:

Potassium in sample West Seep %D (26) the result is qualified as Estimated [J].

Analyte Quantitation and Reporting Limits

Analyte quantitation was evaluated for all samples. No calculation or transcription errors were found. The results and reporting limits were correctly reported.

Overall Comments

The data are acceptable as reported with the following exception:

The water samples were all found to be field filtered and had a pH less than 2. Cooler #1 was received at a temperature of 5°C. However there is no documentation as to which samples were in the cooler. This should be taken into account when determining the usability of the data for all analyses except for certain radiochemistry analyses.

Potassium in sample West Seep %D (26) is qualified as Estimated [J] due to serial Dilution failure.

ICP-MS samples were analyzed at a dilution in order to bring uranium into analytical range.

No CRDL or CRI standard recovery summary forms (EPA Form 2b) were included in the data package. The reviewer obtained the results from the raw data. No action was necessary.

DATA QUALIFIER DEFINITIONS

For the purpose of Data Validation, the following code letters and associated definitions are provided for use by the data validator to summarize the data quality.

- R Reported value is "rejected." Resampling or reanalysis may be necessary to verify the presence or absence of the compound.
- J The associated numerical value is an estimated quantity because the Quality Control criteria were not met.
- U J The reported quantitation limit is estimated because Quality Control criteria were not met. Element or compound was not detected.
- U The material was analyzed for, but was not detected above the level of the associated value. The associated value is either the sample quantitation limit or the sample detection limit.
- NR Result was not used from a particular sample analysis. This typically occurs when more than one result for an element is reported due to dilutions and reanalyses.

DATA VALIDATION REPORT

To:	Robert Hill
From:	John Garrett
Date:	July 30, 2010
Project/Site:	Colorado School of Mines
Project No .:	4060
SDG No.:	1006125

This report presents the Dissolved Organic Carbon data validation for the data obtained for fourteen CSMRI water sample collected on June 08, 2010, June 09, 2010, and June 10, 2010 and submitted to ALS Laboratory Group on June 11, 2010 for the above referenced work assignment. The purpose of this review is to provide a technical evaluation of eleven Dissolved Organic Carbon results that were obtained by MCAWW, May 1994, Method 415.1, SOP 670R13 Dissolved Organic Carbon by Method 415.1 from ALS Laboratory Group (Fort Collins, CO). The water samples were analyzed June 15, 2010. All analyses were conducted by ALS Laboratory Group. The field sample numbers and corresponding laboratory numbers are presented below:

Client Sample Number	Laboratory Sample Number	Matrix	Collection Date
CSMRI-1	1006125-1	Water	June 10, 2010
CSMRI-1B	1006125-2	Water	June 09, 2010
CSMRI-2	1006125-3	Water	June 10, 2010
CSMRI-4	1006125-4	Water	June 08, 2010
CSMRI-5	1006125-5	Water	June 08, 2010
CSMRI-6C	1006125-6	Water	June 08, 2010
CSMRI-8	1006125-8	Water	June 08, 2010
CSMRI-9	1006125-9	Water	June 09, 2010
CSMRI-10	1006125-10	Water	June 09, 2010
CSMRI-11B	1006125-11	Water	June 08, 2010
SW-1	1006125-12	Water	June 09, 2010
SW-2	1006125-13	Water	June 09, 2010
SW-3	1006125-14	Water	June 10, 2010
EQUIPMENT BLANK	1006125-15	Water	June 10, 2010

Data validation was conducted in accordance with the USEPA Contract Laboratory Program National Functional Guidelines for Organic Data Review (CLP).

The Dissolved Organic Carbon data were evaluated based on the following parameters:

- * Data Completeness
- * Holding Times and Preservation
- * Initial and Continuing Calibration Verification
- * Contract Required Detection Limit (CRDL)
- * Preparation/ Initial (ICB)/ and Continuing (CCB) Calibration Blanks
- * Interference Check Sample (ICSA) Results
- * Matrix Spike Results
- * Duplicate Sample Results
- * Laboratory Control Samples (LCS) Results
- * Serial Dilution Sample Results
- * Compound Quantitation and Reporting Limits (full validation only)

* All criteria were met for this parameter

Data Completeness

The data package was complete. No results were qualified as a result of the missing data.

Holding Times and Preservation

Analytical holding times were evaluated and all criteria were met.

The water samples were all found to be field filtered and had a pH less than 2. Cooler #1 was received at a temperature of 5°C. However there is no documentation as to which samples were in the cooler. This should be taken into account when determining the usability of the data for all analyses except for certain radiochemistry analyses.

Initial and Continuing Calibration Verification

Initial and Continuing Calibration Verification standards were analyzed at the required frequency and all were within the required 90-110%. No action was necessary.

Contract Required Detection Limit (CRDL)

All CRDL %Rs CRI %Rs were within 80-120% limits. No action was necessary.

Preparation and Initial/ Continuing Calibration Blanks

Preparation and Initial/Continuing Calibration Blank analyses were performed at the required frequency. Preparation and Initial/ Continuing Calibration Blanks are evaluated to assess the level of contamination in the preparation and analytical processes.

Preparation and Initial/ Continuing Calibration Blanks were prepared and analyzed at the required frequencies.

All of the blanks that were analyzed had concentrations that were below their respective Reporting Limits (RLs).

However, if blank results were above the Instrument Detection Limits (IDLs) and below the RLs, it caused the associated sample results to be qualified for contamination as estimated and non-detected [UJ 107]. If blank results were below the negate IDL and above the negate RL, it caused the associated sample results to be qualified for negative contamination as estimated [J 107]. No sample results were qualified due to blank contamination.

Matrix Spike/Matrix Spike Duplicate Results

MS/MSD analyses were performed at the required frequency. All MS/MSD percent recoveries were within 75-125% limits. No action was necessary.

Duplicate Sample Analysis

Duplicate analyses were performed at the required frequency. All original sample/duplicate sample and MS/MSD differences were less than 20% RPD or less than the RDL for results less than (5)(RDL). No actions were necessary.

Laboratory Control Samples

LCS analyses were performed at the required frequency. The laboratory analyzed laboratory control samples for all analytes. All recoveries were within 80-120% limits. No action was necessary.

Serial Dilution Results

No dilutions were required.

Analyte Quantitation and Reporting Limits

Analyte quantitation was evaluated for all samples. No calculation or transcription errors were found. The results and reporting limits were correctly reported.

Overall Comments

The overall data are acceptable as reported with the following exception:

The water samples were all found to be field filtered and had a pH less than 2. Cooler #1 was received at a temperature of 5°C. However there is no documentation as to which samples were in the cooler. This should be taken into account when determining the usability of the data for all analyses except for radiochemistry analyses.

DATA QUALIFIER DEFINITIONS

For the purpose of Data Validation, the following code letters and associated definitions are provided for use by the data validator to summarize the data quality.

- R Reported value is "rejected." Resampling or reanalysis may be necessary to verify the presence or absence of the compound.
- J The associated numerical value is an estimated quantity because the Quality Control criteria were not met.
- U J The reported quantitation limit is estimated because Quality Control criteria were not met. Element or compound was not detected.
 - U The material was analyzed for, but was not detected above the level of the associated value. The associated value is either the sample quantitation limit or the sample detection limit.
- NR Result was not used from a particular sample analysis. This typically occurs when more than one result for an element is reported due to dilutions and reanalyses.

DATA VALIDATION REPORT

То:	Robert Hill
From:	John Garrett
Date:	July 27, 2010
Project/Site:	Colorado School of Mines
Project No.:	4060
SDG No.:	1006125 Radium-226

This report presents the radiological data validation for the data obtained during the field activities for the above referenced work assignment. The purpose of this review is to provide a technical evaluation of the radiological results that were obtained by ALS Laboratory Group Procedure SOP 783R8 for Radium-226 by Radon Emanation Counting for SDG 1006125 from ALS Laboratory Group (Fort Collins, CO). This report consists of fifteen water samples for the Colorado School of Mines/4060 project collected on June 08, 2010, June 09, 2010, and June 10, 2010 and submitted to ALS Laboratory Group on June 11, 2010. The samples were analyzed for Radium-226 by Radon Emanation Counting on June 25, 2010. All analyses were conducted by ALS Laboratory Group. The field sample numbers and corresponding laboratory numbers are presented below:

Client Sample Number	Laboratory Sample Number	Matrix	Collection Date
CSMRI-1	1006125-1	Water	June 10, 2010
CSMRI-1B	1006125-2	Water	June 09, 2010
CSMRI-2	1006125-3	Water	June 10, 2010
CSMRI-4	1006125-4	Water	June 08, 2010
CSMRI-5	1006125-5	Water	June 08, 2010
CSMRI-6C	1006125-6	Water	June 08, 2010
CSMRI-7B	1006125-7	Water	June 10, 2010
CSMRI-8	1006125-8	Water	June 08, 2010
CSMRI-9	1006125-9	Water	June 09, 2010
CSMRI-10	1006125-10	Water	June 09, 2010
CSMRI-11B	1006125-11	Water	June 08, 2010
SW-1	1006125-12	Water	June 09, 2010
SW-2	1006125-13	Water	June 09, 2010
SW-3	1006125-14	Water	June 10, 2010
EQUIPMENT BLANK	1006125-15	Water	June 10, 2010

Data validation was conducted in accordance with the Analytical Services Statement of Work for the following modules: Gas Proportional Counting Module RC04-v2, October 1, 2002, and U.S. DOE Quality Systems for Analytical Services Revision 2.5 (QSAS).

The radiological data were evaluated based on the following parameters:

- * Data Completeness
- * Holding Times and Preservation
- * Instrument Initial Calibrations
- * Instrument Performance Checks
- * Preparation Blanks
- * Duplicate Sample Results
- * Laboratory Control Samples (LCS) Results
- * Laboratory Control Samples Duplicate (LCSD) Results
- * Compound Quantitation and Reporting Limits (full validation only)

Data Completeness

The data package was complete as per ALS Laboratory Group Procedure SOP 783R8 for Radium-226 by Radon Emanation counting.

Holding Times and Preservation

Analytical holding times were evaluated and all criteria were met. However, holding time requirements are not applicable to radiochemistry analyses unless the isotopes of interest have short half-lives.

Calibrations

The instruments were calibrated at the required frequency.

Initial Calibration

All instruments were calibrated properly using NIST traceable SRM.

Instrument Performance Checks

All isotopes were within criteria.

Preparation Blanks

Preparation/Method Blanks were performed at the required frequency. All isotopes that were analyzed had activities that were below their respective MDCs in their QC batch preparation blanks.

Duplicate Sample Analysis

Duplicate analyses were performed at the required frequency. Due to limited sample volume the laboratory prepared a LCSD in lieu of a client sample Duplicate. All isotopic activities for Radium-226 LCS Duplicate and LCS original analysis were within the limits of the statistical test for equivalency. No action was required.

Matrix Spike/Matrix Spike Duplicates

Matrix spike/matrix spike duplicates were not performed for the samples in this SDG, nor were any required.

Laboratory Control Samples

LCS analyses were performed at the required frequency. All recoveries were within 75-125% limits. No calculation errors or transcription errors were found.

Analyte Quantitation and Reporting Limits

Analyte quantitation was evaluated for all samples. No calculation or transcription errors were found. The results and reporting limits were correctly reported.

Overall Comments

Overall, the data are of good quality and are usable as reported by the laboratory without qualification.

Radium-226 was detected above the RDL in sample CSMRI-5 at 2.42 pCi/L and is considered detected.

The laboratory reported that the ICP-AES measurement of the added barium carrier prior to chemical separation. Several samples showed barium concentrations less than zero. The laboratory manually adjusted the values to 0.0 in order to avoid a low bias. All QC criteria were within control limits and no action was necessary. The data are not affected.

DATA QUALIFIER DEFINITIONS

For the purpose of Data Validation, the following code letters and associated definitions are provided for use by the data validator to summarize the data quality.

- R Reported value is "rejected." Resampling or reanalysis may be necessary to verify the presence or absence of the compound.
- J The associated numerical value is an estimated quantity because the Quality Control criteria were not met.
- U J The reported quantitation limit is estimated because Quality Control criteria were not met. Element or compound was not detected.
- U The material was analyzed for, but was not detected above the level of the associated value. The associated value is either the sample quantitation limit or the sample detection limit.
- NR Result was not used from a particular sample analysis. This typically occurs when more than one result for an element is reported due to dilutions and reanalysis.

DATA VALIDATION REPORT

To:Robert HillFrom:John GarrettDate:July 27, 2010Project/Site:Colorado School of MinesProject No.:4060SDG No.:1006125 Radium-228

This report presents the radiological data validation for the data obtained during the field activities for the above referenced work assignment. The purpose of this review is to provide a technical evaluation of the radiological results that were obtained by ALS Laboratory Group PA SOP 724R10 for Radium-228 by gas proportional counting for SDG 1006125 from ALS Laboratory Group (Fort Collins, CO). This report consists of fifteen water samples for the Colorado School of Mines/4060 project collected on June 08, 2010, June 09, 2010, and June 10, 2010 and submitted to ALS Laboratory Group on June 11, 2010. The samples were analyzed for Radium-228 by Radon gas proportional counting on June 28, 2010, July 1, 2010, July 12, 2010, and July 14, 2010. All analyses were conducted by ALS Laboratory Group. The field sample numbers and corresponding laboratory numbers are presented below:

Client Sample Number	Laboratory Sample Number	Matrix	Collection Date
CSMRI-1	1006125-1	Water	June 10, 2010
CSMRI-1B	1006125-2	Water	June 09, 2010
CSMRI-2	1006125-3	Water	June 10, 2010
CSMRI-4	1006125-4	Water	June 08, 2010
CSMRI-5	1006125-5	Water	June 08, 2010
CSMRI-6C	1006125-6	Water	June 08, 2010
CSMRI-7B	1006125-7	Water	June 10, 2010
CSMRI-8	1006125-8	Water	June 08, 2010
CSMRI-9	1006125-9	Water	June 09, 2010
CSMRI-10	1006125-10	Water	June 09, 2010
CSMRI-11B	1006125-11	Water	June 08, 2010
SW-1	1006125-12	Water	June 09, 2010
SW-2	1006125-13	Water	June 09, 2010
SW-3	1006125-14	Water	June 10, 2010
EQUIPMENT BLANK	1006125-15	Water	June 10, 2010

Data validation was conducted in accordance with the Analytical Services Statement of Work for the following modules: Gas Proportional Counting Module RC04-v2, October 1, 2002, and U.S. DOE Quality Systems for Analytical Services Revision 2.5 (QSAS).

The radiological data were evaluated based on the following parameters:

- * Data Completeness
- * Holding Times and Preservation
- * Instrument Initial Calibrations
- * Instrument Performance Checks
- * Preparation Blanks
- * Duplicate Sample Results
- * Laboratory Control Samples (LCS) Results
- * Laboratory Control Samples Duplicate (LCSD) Results
- * Compound Quantitation and Reporting Limits (full validation only)

Data Completeness

The data package was complete as per ALS Laboratory Group Procedure SOP 724R10 for Radium-228 by gas flow proportional counting for SDG 1006125.

Holding Times and Preservation

Analytical holding times were evaluated and all criteria were met. However, holding time requirements are not applicable to radiochemistry analyses unless the isotopes of interest have short half-lives.

Calibrations

The instruments were calibrated at the required frequency.

Initial Calibration

All instruments were calibrated properly using NIST traceable SRM.

Instrument Performance Checks

The LCSD sample 1006125-2DUP in Batch RA100622-1 was analyzed on detector C4 which failed the daily efficiency check for alpha. The affected samples are CSMRI-1B, and CSMRI-7B. These samples are considered [R] Rejected due to the combination of the detector failure, the Batch LCS failure, and the ICP-AES barium carrier concentration

being less than the known amount added. <u>Preparation Blanks</u>

Preparation/Method Blanks were performed at the required frequency. All isotopes that were analyzed had activities that were below their respective MDC's in their QC batch preparation blanks.

Duplicate Sample Analysis

Duplicate analyses were performed at the required frequency. Due to insufficient sample volume the laboratory prepared and analyzed a Laboratory Control Sample Duplicate (LCSD) in lieu of a client sample duplicate.

The LCSD sample 1006125-2DUP in Batch RA100622-1 was analyzed on detector C4 which failed the daily efficiency check for alpha. The affected samples are CSMRI-1B, and CSMRI-7B. All other samples were re-prepped and re-analyzed with acceptable results.

All isotopic activities for Radium-228 duplicate (LCS) and original (LCSD) analysis were within the limits of the statistical test for equivalency. No action was required.

Matrix Spike/Matrix Spike Duplicates

Matrix spike/matrix spike duplicates were not performed for the samples in this SDG, nor were any required.

Laboratory Control Samples

LCS analyses were performed at the required frequency.

The LCS in Batch RA100622-1 was outside control limits low. The affected samples are CSMRI-1B, and CSMRI-7B and are considered [R] Rejected due to the combination of batch problems. All other samples associated with the batch were re-prepped and re-analyzed with acceptable results.

The LCS in Batch RA100622-2 was outside control limits low. The affected sample SW-1 and is qualified [J/UJ]. All other samples associated with the batch were re-prepped and re-analyzed with acceptable results.

All other recoveries were within 75-125% limits. No calculation errors or transcription

S.M. Stoller Corp.

errors were found.

Analyte Quantitation and Reporting Limits

Analyte quantitation was evaluated for all samples. No calculation or transcription errors were found. The results and reporting limits were correctly reported.

Overall Comments

The LCSD sample 1006125-2DUP in Batch RA100622-1 was analyzed on detector C4 which failed the daily efficiency check for alpha. The affected samples are CSMRI-1B, and CSMRI-7B. These samples are considered [R] Rejected due to the combination of the detector failure, the Batch LCS failure, and the ICP-AES barium carrier concentration being less than the known amount added.

The LCSD sample 1006125-2DUP in Batch RA100622-1 was analyzed on detector C4 which failed the daily efficiency check for alpha. The affected samples are CSMRI-1B, and CSMRI-7B. All other samples were re-prepped and re-analyzed with acceptable results.

The LCS in Batch RA100622-1 was outside control limits low. The affected samples are CSMRI-1B, and CSMRI-7B and are considered [R] Rejected due to the combination of batch problems. All other samples associated with the batch were re-prepped and re-analyzed with acceptable results.

The LCS in Batch RA100622-2 was outside control limits low. The affected sample SW-1 and is qualified [J/UJ]. All other samples associated with the batch were re-prepped and re-analyzed with acceptable results.

The laboratory reported that the ICP-AES measurement of the added barium carrier prior to chemical separation had a concentration of less than the concentration added. The laboratory manually adjusted the values to the known concentration to calculate the chemical yield in order to avoid a low bias in all samples including the QC. All samples reported barium concentrations less than that known to be added. Samples CSMRI-9, CSMRI-10, SW-1, SW-2, and EQUIPMENT BLANK have significant low bias and are qualified as [J/UJ] Estimated.

Radium-228 was detected above the RDL in sample CSMRI-2 at 2.25 pCi/L, CSMRI-4 at 1.42 pCi/L and CSMRI-6C at 1.48 pCi/L and are considered detected. The following samples are qualified:

- CSMRI-1B, and CSMRI-7B [R] Rejected due to multiple problems,
- SW-1 [J/UJ] Estimated due to LCS outside control low,
- CSMRI-10, SW-1, SW-2, and EQUIPMENT BLANK [J/UJ] Estimated due to

carrier problems.

DATA QUALIFIER DEFINITIONS

For the purpose of Data Validation, the following code letters and associated definitions are provided for use by the data validator to summarize the data quality.

- R Reported value is "rejected." Resampling or reanalysis may be necessary to verify the presence or absence of the compound.
- J The associated numerical value is an estimated quantity because the Quality Control criteria were not met.
- U J The reported quantitation limit is estimated because Quality Control criteria were not met. Element or compound was not detected.
 - U The material was analyzed for, but was not detected above the level of the associated value. The associated value is either the sample quantitation limit or the sample detection limit.
 - NR Result was not used from a particular sample analysis. This typically occurs when more than one result for an element is reported due to dilutions and reanalyses.

Appendix E Results of Analyses CD

Appendix F Chains of Custody

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-S Laboratory Group 25 Commerce Drive, Fort Collins, CO 80524

TF: 800-443-1511 PH: 970-490-1511 FX: 970-490-1522

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Return to Client UN7500Rn Method Above 0'906E ò 11890 By Lab 2W9320 E904.0 5 Analv 1-E063-1 0.5003 815 E903.0 00 0.000310 E900.0 0 L'1063 Circle: Am / Cm / Pu / Th / U 0.4155 SM/SM/2V E314.0 1'0913 39406MS 80406MS TDS; E160,1 TSS; E160,2 Total; E160,3 0.00053 9806WS NIX Shortsegid anilakiA ¥961/M5 SW6020A E200.8 SW6020A E200.8 SW6010B 7470A/7471A E200.7 E245.1 SW6010B 7470A/7471A E200.7 E245.1 (TO) A0741W2 80103W2 SW8260C SW8270D SW8081A SW8151A Circle SW8330 SW8330B SW8351 P/ LC/MS/MS AFEF8W2 SW8082 A1808W2 CI0228MS Sampler(s) 209Z8MS 200928M8 SW8015B GRO DRO Other Provide additional information as needed in Comments below Ę FAX: Project Name/No. REPORT TO: PHONE COMPANY E-MAIL: ADDRESS:

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N (2) Return to Client 11 Radon 222 UR7500Rn Above Time Time muitin 0'906E By Lab or Strontium 90 (Total RadioSr) 118SQ 0.4063 0256WS 822 muibeA 14 Relinquished By 824 muibes Anal 1.C003 Printed Name Printed Name Lab ID Received By muibeA poittim3-englA letoT 0'2063 5126MS Disposal Circle Signature Signature Company Company 2M9310 E900.0 Gross Alpha / Beta Date Date segotosl emmeð 1106E E 7 Circle: Am / Cm / Pu / Th / U Actinides - of X 14 Perchlorate SW6850 by LC/MS/MS E314.0 Due Page 1 Time Hq 1.0213 32406WS 80406WS or TDS: E160.1 TSS: E160.2 Total: E160.3 sbilo2 200 Standard X Inorganic Anions 0'0083 9906MS Relinquished By: Alkaline Digestion? Y / N Hexavalent Chromium ∀9612MS Printed Name Printed Name Received By Dissolved Metals (ICP-MS) 8.0023 A0203W8 Signature Signature Date C Company Company Turnaround (CP-MS) alstal (ICP-MS) 8.0023 A0203WS Date Date Dissolved Metals (ICP) or Hg 5W6010B 7470A/7471A E200.7 E245.1 Mathord Chain _r-Custody SW6010B 7470A/7471A E200.7 E246.1 Total Metals (ICP) or Hg Form 202r7 (5/19/09) tical TCLP Metals SW1311 A0747WS 80108WZ TCLP Organics SW1311 SW8260C SW8270D SW8081A SW8151A E = extract F = filterCircle SW8330 SW8330B SM8351 PÅ FC/WS/WS **Explosives** Herbicides AT218W2 2808WS **bCBs** 3 W = water L = liquid A1808WS **OC Pesticides** SVOCS Q0728W8 Sampler(s) BTEX + MtBE 200528WS 20928M8 SOOA Originator: Retain pink page or a photocopy! Matrix: O = pil S = soil NS = non-soil solid SW8015B GRO DRO Ofher Hdl 17 No. of Containers 11.24 TF: 800-443-1511 PH: 970-490-1511 FX: 970-490-1522 Preservative (Type HCI, etc.) 1 4 Provide additional information as needed in Comments below 225 Commerce Drive, Fort Collins, CO 80524 xinteM Lab ID 01.10 Time ' For metals or anions, please detail analyte list below. Date 3 PST. MST Project Name/No. REPORT TO: FAX COMPANY PHONE E-MAIL: ADDRESS: 1 . . Sample ID CST EST Comments: Zone (Circle):

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5 (2) Return to Client > Radon 222 UR7500Rn Time Time multinT 0'906E 5 (120ibeA letoT) 00 muitnott2 1189G Lab X 0.4063 0256WS 822 muibeA X NA N X × Relinquished By L'2063 825 muibeA Printed Name Printed Name Lab ID Received By mulbeA gnittim3-sriglA lstoT 0'E063 91E6MS Circle Disposal Signature Signature Company Company 0.00053 0166WS Gross Alpha / Beta Date Date seqotosl smmsD 1.1003 E Circle: Am / Cm / Pu / Th / U Actinides (1970) 1145 of X 0 2M6850 by LC/MS/MS E314.0 Perchlorate Due Page 2M9040B 2M9046C E150.1 Hd ime Time 0 Kar Standard, or TDS: E160.1 TSS: E160.2 Total: E160.3 sbilo2 Inorganic Anions 0'00EE 9906MS Alkaline Digestion? Hexavalent Chromium NIL ¥961/MS Printed Name Relinquished × × Dissolved Metals (ICP-MS) Received By SW6020A E200.8 Printed Nar Signature _ Company Signature Company Date V Turnaround 8.0020 E200.8 (2M-90) alstaM lstoT Method Above Date Date SW6010B 7470A/7471A E200.7 E245.1 Dissolved Metals (ICP) or Hg Total Metals (ICP) or Hg 5W6010B 7470A71A71A E200.7 E245.1 Form 202r7 (5/19/09) alvtical A0747WS 80108W2 TCLP Metals SW1311 TCLP Organics SW1311 SW8260C SW8270D SW8081A SW8151A F = fite Circle SW8330 SW8330B SM8351 PA FC/WS/WS **Explosives** $\mathbf{E} = extract$ ATE18W2 Herbicides **bCBs** SW8082 L = liquic **OC Pesticides** A1808W2 2W8270D SVOCS W = water Sampler(s) BTEX + MtBE 2M8560C 20928MS SOOA Originator: Retain pink page or a photocopy! NS = non-soil solid SW8015B GRO DRO Ofher HdT No. of Containers TF: 800-443-1511 PH: 970-490-1511 FX: 970-490-1522 2.2 Preservative (Type HCL etc.) 225 Commerce Drive, Fort Collins, CO 80524 Provide additional information as needed in Comments below 4 xinteM -S = soil Cab ID 0 = 0 Time the second For metals or anions, please detail analyte list below. Matrix: Date 1 400 400 PS MST Project Name/No. REPORT TO PHONE FAX COMPANY ADDRESS ELMAN Sample ID CST EST Comments: 1 Zone (Circle):



(2) (2) **Return to Client** UNOOSTAR Radon 222 Above Time muitinT 0'906E 10 Math Strontium 90 (Total RadioSr) 1189D By Lab 0.100E904.0 C3 822 muibeA Relinquished By Anal 1 E903'1 8adium 226 Printed Name Lab ID Received By muibeA prittim3-englA letoT 0'8063 9186MS Circle Disposal Signature Signature Company Gross Alpha / Beta 0'0063 0186MS Date sagotosl emmeð 1-106E (1) Circle: Am / Cm / Pu / Th / U Actinides 61111 of X 1 Perchlorate 2/16/20 Py LC/MS/MS E314.0 Due Page 1:0913 39406MS 80406MS Hd me ō TDS: E160.1 TSS: E160.2 Total: E160.3 sbilo2 100 14 Standard Inorganic Anions × 0'0083 9906MS 153 Alkaline Digestion? Y / N Relinquished By: Printed Name Mexavalent Chromium ABELYWE Received By (SM-90) alsteM bevlossid 8.0023 A0203WS Signature Signature Company Turnaround Date (8.0053 A0203WS (CP-90) aleteM letoT Date Dissolved Metals (ICP) or Hg SW6010B 7470A/7471A E200.7 E245.1 Chain *i*-Custody Total Metals (ICP) or Hg SW6010B 7470A/7471A E200.7 E245.1 tica TCLP Metals SW1311 A0747W2 80108W2 Ana TCLP Organics SW1311 SW8260C SW8270D SW8081A SW8151A F = filter Circle **s**avizolqx**∃** SW8330 SW8330B SW8351 PY LC/MS/MS = extract AIG18W2 Herbicides 2808WS **bCBs** ш. T = nquic A1808WS **OC Pesticides** Q0228MS SVOCS W = water Sampler(s) BTEX + MtBE 200528MS 200928MS SOOK NS = non-soil solid SW8015B GRO DRO Olher HdT No. of Containers TF: 800-443-1511 PH: 970-490-1511 FX: 970-490-1522 1 Mart Preservative (Type HCI, plc.) -S Laboratory Group 225 Commerce Drive, Fort Collins, CO 80524 Provide additional information as needed in Comments below 10 xinteM S = SOILab ID 4 Matrix: O = O()Time * 10 f For metals or anions, please detail analyte list below. Date . 12 64 PST MST. 100 Project Name/No. REPORT TO: FAX: BNOHA E-MAIL: ADDRESS: COMPANY -Sample ID. CST N EST

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Appendix G Historical Summary Tables

Total U	1.61	0.64	1.3	2.8	0.31	1.2	0.88	0.72	1.5	1.9	1.5	2	1.6	2.4	2.9	2.7	6.3	6.9	6.5	0 4	4,6	8,1	34	16	9.4	0.89	0.94	1.06	1.36	0.85	0.72	2	0.98	0.68	0.89	0.83	0.77	0.66	0.75	0.59	31.4	19.3	27.1	26.8	17.9 48	66	49	48 58	62	43 61	80	110	79 83	NT	2.8 3.57	4.4	4.1 4.1	2.8	5.8	10	6.6	6.6	10	10	12	11	10	TN
U-238 (nCi/l)	0.53	0.217	0.43	0.92	0.08	TN	TN	TN TN	t L	LN F	LN	NT	TN F	Z Z	NT	tz t	LN	NT	ty t	z Lz	NT	LZ LZ	t N	μ	NT	0.10	0.31	0.35	0.45	0.28	μŢ	۲Z :	I L	ΤN	NT	NT TN	ty :	IZ LU	τN	N1 08	10.6	6.4	9	8.9	9 TN	NT	TN TN	NT N	TN TN	TN TN	TN	NT	TN -	TN CO	0.93	1.47	1.21	0.92	22 NT	TN TN	z Ł	TN TN	μ	TN TN	E LA	TN	z L	-
U-235 (nCi/l)	0.043	0.011	0.073	0.11	0.18	LCU.U	NT	T T T	z L	TN TN	N TN	NT	L L	zŁ	NT	TN FN	NTN	NT	TZ 5	z tz	NT	IN TN	NTN	ΤN	NT	0.025	0.05	0.002	0.066	0.014	ΝŢ	TN F	IN TN	ΤN	T TN TN	NT	tZ :	IN TN	ΤN	0.53	0.49	0.33	0.43	0.4	0.35 NT	NTN	TN F	NTN	LN F	NT NT	TN	NT	TN 1	TN Seco	0.056 0.086	0.051	0.058	0.13	NT N	NT	N TN	TN TN	NT	NT	E LN E	TN TN	NT	1
U-234 (nCi/l)	0.77	0.43	0.94	1.76	0.18	C+'D	NT	T T T	zŁ	TZ F	z L	NT	E F	z L	NT	TN F	z L	μ	T F	z z	NT	IN IN	NTN	NT	NT SC	0.68	0.65	0.83	0.83	0.46	NT	LN H	Z Z	NT	T T	NTN	LN F	z tz	NT	1N 2.2	11.4	6.4	6	9.2	6.5 NT	NT	LZ :	Z T	LN F	TA TA	TN F2	NT	TZ :	LN S	1.22	1.85	1.81	1.03	N TN	NT T TN	z t	T T	NT	NT NT	E E	TA TA	z tz	=
Th-232 (nCi/l)	0.01	0.012	0.014	-0.004	0.062	0.012	-0.005	0.001	0.011	0.01	N IN	NT.	FS :	z z	NT	0.02	0.001	0.016	0.033	o TV	NT	Z	NTN	μ	TN 60	0.026	0.049	0.024	0.012	800.0-	0.011	-0.007	0.006	0.044	0.008 NT	NTN	LZ I	zŁ	τN	0.013	0.005	0.086	0.007	0.061	0.007	0.004	0.021	0.016	0.002	TN TN	LN F	NT N	tz !	IN S	0.034	0.051	0.019	0.043	0	0.013	0.01	0.051	NT	TN TN	L L	ty ty	IN IN	:
Th-230 (nCi/l)	0.07	-0.021	0.32	0.032	-0.06	-0.031	0.064	0.019	0.032	-0.004	NT	Ъ	TN F	z	NT	0.09	0.012	0.06	0.058	-0.040 NT	NT	Z	NTN	NT	NT	0.003	0.108	0.285	0.204	-0.039	-0.01	-0.01	0.035	0.085	-0.021 NIT	N TN	TN F	z t	NT	1N 1000-0-	0.014	0.164	0.174	0.17	-0.015	0.035	0.112	0.149 0.038	-0.019	TN TN	Ę	Z LZ	tz !	TZ C	0.007	1.25	0.144	0.08	-0.061	-0.023	-0,008	0.02	NT	T T	ΞŻŻ	NT NT	z z	i.
Th-228 (nCill)	0.007	0.018	-0.045	0.025	0.15	0.052	0.17	-0.031	0.035	-0.03 MT	z L	ţ	tz t	TZ Z	NT	-0.03	0.019	0.004	0.01	-0.004	μ	ZZ	TN TN	τN	NT 0.07	0.14	0.162	0.108	0.03	0.088	0.022	-0.075	0.037	0.043	0.041 NIT	TN TN	t t	z tz	τN	0.019	0.013	-0.013	0.004	0.11	0.056	0.011	0.024	0.029	0.017	tu tu	TN F	IN TN	ty !	LN CO	-0.018	0.06	0.038	0.145	-0.012	0.008	0.004	-0.011	TN	TN TN	E E E	LN LN		:
Ra-228 (nCi/l)	0.81	0.44	0.59	0.58	0.05	0.78	0.91	0.53	0.85	0,66	1.39	0.96	0.16 J	0.96	0.38	1.19	0.65	1,16	0.61	0.3	0.88	1.15	0.89	0.98	0.42	3	2.71	1.62	2.53	2.35	2.15	3.2	3.2 2.05	2.26	1.89	1.88	2.68	2.64 J 2.12	1.6	0.16	0.34	0.78	0.38	0.39	0.79	0.87	1.12	0.78	0.71	0.97	0.56	0.73	0.68	N N	0.53	0.76	0.45	0.87	0.39	0.86	0.49	0.17 0.64	0.47	0.68 1.24	1.15 J	0.85	0.4	Ł
Ra-226 (nCi/l)	-0.11	0.16	-0.19	-0.15	0.42	0.32	0.51	0.3	0.2	-0.02	0.32	60.0	0.19	2.04	0.11	0.13	-0.13	0.11	0.32	0.05	0.02	0.2	0.08	-0.03	0.09	1.47	1.78	1.35	1.25	1.01	0.76	1.4	0.45	1.37	1.08	1.1	2.37	0.63	1.02	-0.03	0.26	0.17	0	0.41	-0.05	0.07	0.99	0.33	0.11	0.32	0.54	0.11	0.21	0.38	2.51	2.5	0.57	2.13	1.78	2.22	1.52	1.05	2.87	0.78	1.96	-0.15	3.9	0.83
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Table G-1 storical Summary of Radioisotopes in Groundwater (Sto	
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3/8/2010 NT NT NT NT	-		TN
-	*		NE

Table G-2	Summary of Metals in Groundwater (Stoller)	All results in milligrams per liter)
	Historical	

Zn 0.032 0.032 0.034 0.034 0.049 0.049 0.048 0.049 0.049 0.049 0.049 0.049 0.049 0.049 0.049 0.049 0.049 0.049 0.057 NT NT NT NT	NT ND 0.0012 (B) 0.0033 (B) 0.0033 (B) 0.0033 (B) NT NT NT NT NT NT NT NT NT NT NT NT NT	NT NT NT NT NT NT NT 0.12 0.068 0.14 0.14 0.14 0.14 0.14 0.14 0.14 0.14	NT NT NT NT NT 0.067 0.047 0.047 0.083 0.117 0.055 0.055 0.055 0.054 0.025 0.025	0.093 NT NT NT NT NT NT NT ND 0.0051 (B) 0.0048 0.0099 (B) NT NT NT NT NT NT NT NT
> 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2	NT 0.0009 (B) ND 0.00055 (B) NT NT NT NT NT NT ND ND ND ND ND ND ND ND ND ND ND ND ND	NT NT NT NT NT NT ND ND 0.00056 (B) 0.00056 (B) 0.0011 (B) ND 0.0012 (B) 0.0012 (B) 0.0013 (B) 0.0013 (B) 0.0013 (B)	NT NT NT NT NT ND ND 0.0012 (B) 0.00057 (B) ND 0.00057 (B) ND 0.0012 (B) ND 0.0017 (B) ND 0.0017 (B) ND 0.0017 (B) ND ND ND ND ND ND ND ND ND ND ND ND ND	0.0011 (B) NT NT NT NT NT NT NT ND 0.00081 (B) 0.00081 (B) ND 0.00083 (B) NT NT NT NT NT NT NT NT NT NT NT NT NT
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Na Na 26 26 26 26 26 26 21 14 14 14 14 26 26 21 27 27 27 27 27 27 27 27 27 27 27 27 27	42 91 35 36 36 36 36 41 40 10 19 19 19 19 19 19 19 19 19 19 19 19 19	20 19 0,0019 19 19 19 19 19 19 19 19 19	63 48 63 63 69 (J) 69 (J) 67 55 77 23 83 34 47 47	41 47 47 47 47 55 55 0 0.0019 55 46 41 41 41 41 85 0 0 7 (J) 55 49 49 0019 0019 NT NT NT NT NT NT NT NT NT NT NT NT NT
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Mg 9.4 6.3 10 10 10 10 10 11 12 12 12 12 12 12 12 12 12	19 47 47 47 47 47 47 47 47 47 33 33 33 33 33 33 33 33 33 33 33 33 33	33 34 35 35 35 35 35 35 33 33 34 43 33 35 49 49 49 49 56 56 56 56 56 56 56 56 56 56 56 56 56	54 45 45 38 65 (J) 67 49 31 49 31 31 33 33 33 33 33 33 33 33 33 33 33 33 33 33 34 40 40	40 61 61 61 61 61 61 70 75 6 75 6 10 10 NT NT NT NT NT NT NT NT NT NT NT NT NT
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Hg ND ND ND ND 0.000034 (B) 0.000034 (B) ND ND ND ND ND NT NT NT NT NT	NT 0.000017 (B) 0.000029 (B) 0.000029 (B) ND ND NT NT NT NT NT NT ND ND ND ND ND ND ND ND ND ND	0.000024 NT NT NT NT ND ND ND 0.000045 (B) 0.000045 (B) 0.000045 (B) 0.000045 (B) 0.000045 (B) 0.000045 (B) 0.000022 (B) 0.000016 (B) 0.000015 (B) 0.000015 (B)	0.00013 NT NT NT NT NT ND ND ND 0.000048 (B) 0.000048 (B) 0.000048 (B) 0.000048 (B) 0.000048 (B) 0.000048 (B) 0.000048 (B) ND ND ND ND ND ND ND ND ND ND ND ND ND	0.000018 (8) NT NT NT NT NT NT NT NT NT ND ND ND NT NT NT NT NT NT NT NT NT NT NT NT NT
Cr ND ND ND ND ND ND ND 0.00051 (B) 0.00053 (B) ND ND ND ND ND ND ND ND ND ND ND ND ND	0.00014 (B) ND ND ND ND NT NT NT NT NT NT NT NT ND ND ND ND ND ND ND ND ND ND	0.0041 0.00041 0.00041 0.00041 0.00041 0.00041 0.0014 (B) 0.0014 (B) 0.0014 (B)	NT NT NT NT NT ND ND ND ND ND ND 0.0002 (B) ND 0.00022 (B) ND 0.00082 (B) ND ND ND ND ND ND ND ND ND ND ND ND ND	ND NT NT NT NT NT NT ND ND ND ND ND ND ND ND NT NT NT NT NT NT NT NT NT NT NT NT NT
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	0.098 (B) 0.071 (B) 0.11 0.11 0.11 0.14 NT NT NT NT NT NT 0.14 0.14 0.11 0.11 0.11 0.038 (B) 0.098 (B) 0.093 (B) 0.092 (B) 0.002 (B) 0.0			
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	A A A A A A A A A A A A A A A A A A A	0	0	0
Sample Date 2/25/2005 6/14/2005 9/7/2005 9/7/2006 9/13/2006 9/13/2006 9/14/2006 9/14/2006 9/14/2006 9/11/2007 11/27/2007 9/24/2009 6/24/2009 6/24/2009 9/24/2009	3/1/2007 3/1/2007 9/1/1/2007 9/1/1/2007 9/1/1/2008 1/1/2008 9/26/2008 9/26/2008 9/26/2009 9/26/2009 9/26/2009 9/26/2009 9/26/2005 6/14/2005 6/14/2005 6/14/2005 9/7/2005 9/7/2005 9/7/2005 9/7/2005 9/1/2005 9/2/2009 9/2/2/2005 9/1/2005 9/2/2/2005 9/1/2/2005 9/2/2/2005 9/2/2/2005 9/2/2/2005 9/1/2/2005 9/2/2/2005 9/	12/5/2009 3/18/2009 6/23/2009 9/25/2009 12/18/2009 3/11/2010 2/25/2005 6/14/2005 6/14/2005 6/14/2005 3/15/2006 9/13/2007 3/15/2006 9/13/2007 9/11/2008 9/11/	12/5/2008 3/17/2009 9/24/2009 1/2/16/2009 3/10/2010 2/25/2005 6/14/2005 9/15/2005 9/15/2005 9/13/2006 6/15/2006 9/13/2006 9/13/2006 9/13/2006 9/13/2006 9/13/2006 9/13/2007 9/11/2007 9/11/2007	4/17/2008 9/25/2008 3/17/2009 3/17/2009 5/23/2009 9/24/2009 3/16/2007 3/10/2017 2/28/2008 4/18/2009 3/16/09 1/2/18/2008 4/18/2008 1/2/3/08 6/24/2009 6/24/2009 6/24/2009 9/24/2009 9/24/2009 3/8/2010
Sample SSMRH	CSMRI-18 CSMRI-2	CSMRI-4	CSMRI-5	CSMRI-66 CSMRI-66

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Table G-2	istorical Summary of Metals in Groundwater (Stoller)	All results in milligrams per liter)
Tab	storical Summary of Mer	(All results in m

Station	Cample Date	~~	Ve	De		70	-	-					1	-		
Idlight	2/27/2007	NTN	NT	NT	NT	NT	5 12	6H TN	NT	BM	NT	Na	Pb TN	Se	NT NT	Zn
	6/26/2007	QN	DN	0.056 (B)	70	QN	QN	0.000006 (B)	5.5	37	0.024	53	QN	DN	0.00061 (B)	0.0041 (B)
	9/10/2007	Ĭ	NT	IN	IN	NT	NT	NT	NT	NT	NT	NT	NT	NT	NT	NT
	11/26/07 (DRY)	IN	IN	IN	LN.	tu	NT	NT	NT	NT	NT	NT	NT	NT	NT	NT
	2/2//2008 (DRY)	IN	IN	IN	EN .	IN	IN	NT	NT	NT	IN	TN	NT	NT	NT	NT
CCMDI 7D	4/15/08 (DRY)	IN	IN	IN	LN	E !	IN	IN	NT	NT	NT	NT	μ	NT	NT	NT
	1	NIT	IN	IN	IN	Z	IN	IN	IN	LN !	IN	TN	LN	NT	NT	NT
		TIN	LIN	IN FIN	N	z	N	IN	N	N	IN	IN	IN	IN	NT	Į
		LN	LIN	TIN	INI	N	N	IN	N	N	IN IN	N	N	IN	IN	IN
		IN	IN HA	IN	IN IN	N	N	N	Z	Z	Z	IN	IN	IN	NT	NT
	123(2009 (UKY)	NI	IN LY	IN	N	Z	IN	IN	IN	LN I	LN	NT	LN	NT	NT	τN
	12/12/2012	NIT	NI	IN	IN	IN	IN	IN	IN .	NT	LN	LN	TN	NT	NT	NT
	2/10/00/07	UN	1 0 0063 /DV	10 000 VDV	CNI	N	NUN	NI.	NS.	NS	IN O	SN	LN .	NT	NT	IN
	2/01/2/01/2	UN CI	(a) ccuu.u	0.008 (5)	230	NN	ND	- 1-	23	72	0.094	74	QN	0.034	QN	0.0024 (B
	100711710	ON CI.	UN DODO D	(B) COUO	190	QN	QN	0.0000099 (B)	19	55	0.043	52	QN	QN	QN	0.069
	1002/01/6	ND	0.0069 (B)	-	160	QN	0.00074 (B)		15	49	0.034	54	0.0018 (B)	ND	ND	0.025
	11/27/2007	QN	QN	0.091 (B)	230	QN	QN	1.1	15	67	0,026	02	QN	0.0046 (B)	0.001 (B)	0.011 (B)
	2/27/2008	QN	0.036 (B)	-	270	Q	QN	QN	15	82	0.019	100	QN	ND	ND	0.038
CSMRLR	4/1//2008	NN	ON .	-	210	QN	0.0011 (B)		13	63	0.016	73	QN	ND	DN	0.032
D-INIM	8//22/2008	N	Z		230	IN	IN		17	68	NT	70	LN	NT	NT	NT
	8002/G/21	N	IN	Z	400	N	IZ :	TN	18	35	NT	84	μ	TN	NT	NT
	3/ 18/ 2009	IN N	N		250	Z	IN	NT	13	74	NT	26	LN	NT	NT	NT
	6/23/2009	8/000.0	0.0032	0.038	170	0.00095	0.00041	0.00003	14	48	60	0.0019	0.00035	NT	NT	NT
	8/174/2008	N	Z	IN	250 (J)	IN	L	LΝ	13	63 (J)	NT	(l) 87	NT	NT	NT	NT
	2110/010	IN	N	N	210	NI	N	IN	12	59	IN	56	τN	NT	NT	NT
Ĩ	0102010	IN .	N	IN	720	N	N	N	12	11	NT	79	LN	NT	NT	NT
	2/00/1/2/2			0.040 (B)	Ad.	NN	(B) 1100.0	0.000024 (B)	12	3	0,045	33	QN	0.011	0.001 (B)	ND
	/10/2/02/04	AN N.	UN .	0.049 (B)	160	QN	QN	0.000002 (B)	8.5	11	0.0028	150	QN	0.0049 (B)	0.00096 (B)	0.0096 (B)
	/10/2/01/8	ND	0.004 (B)	0.059 (B)	100	QN		0.000016 (B)	ß	51	0.0037 (B)	49	- 1	QN	0.00071 (B)	0.0097 (B
	11/26/2007	QN	QN	0.078 (B)	110	0.00051 (B,		0.000031 (B)	5.9	56	0.0023 (B)	52		0.0054	0.0012 (B)	0.015 (B)
	8/1/2008	NN	ON.	0.079 (B)	110	QN		QN	5.4	56	QN	49		0.0033 (B)	DN	0.011
CSMRI-9	8/02//GL/4	NU	ND	0.077 (B)	100	QN	QN	0.000013 (B)	2	52	0.0017 (B)	46		QN	0.00077 (B)	0,0079 (B
	3/24/2008	IN	IN	N	011	N		IN	5.8	54	NT	50		NT	NT	NT
	2/16/2000	NT	IN LIN	NI	100	IN LA		IN	5.3	48	TN .	46		LN	NT	NT
	6/22/2009	0 00078	0.0032	0.054	750	0.00070		0 000098	4.1	400	IN	01000		IN	N	Z
	9/24/2009	NT	NT	TN	(C) (1)	NT		NT	5.6	58 (.1)	NT	64 (1)		TN	NT	IN
	12/16/2009	LN	NT	NT	140	NT		NT	6.2	67	TN	69		NT	NT	INT
	3/11/2010	NT	NT	NT	140	NT		NT	5.1	29	NT	49		NT	NT	NT
		0.00051 (B)	ND	0.064 (B)	62	ND		0.000024 (B)	7.3	33	0.01	36			0.0011 (B)	QN
	6/26/2007	ND	DN	0.079 (B)	100	QN	QN	0.0000063 (B)	4.7	44	QN	37			0.00055 (B)	QN
	9/10/2007	ND	0.0039 (B)	0.071 (B)	89	ND	0.0012 (B)	0.00002 (B)	4.2	38	0.0014 (B)	36	1	ND	0.00099 (B)	0.0042 (B
	11/26/2007	QN	QN	0.085 (B)	110	QN	QN	0.000026 (B)	4.7	43	QN	41	+		CN	UN
	2/26/2008	DN	QN	0.09 (B)	110	QN	ND	QN	4.6	46	QN	41			QN	0.0052
	4/15/2008	QN	DN	0.088 (B)	100	ND	0.0044 (B)	ND	4.5	44	QN	40			0.00059 (B)	0.0018 (B)
CSIMRI-10	9/24/2008	Ł	NT	NT	100	NT	NT	NT	4.6	42	NT	41	1		NT	NT
	12/4/2008	ΝΤ	NT	NT	100	NT	NT	NT	4.8	41	NT	43	-		NT	NT
	3/16/2009	NT	NT	NT	110	NT	NT	TN	4.5	43	NT	43			NT	NT
	6/22/2009	0.00078	0.0032	0.09	100	0.00016	0.00041	0.00002	4.5	41	40	0,0019			NT	NT
	9/25/2009	IN	IN	IN	120 (J)	IN	NT	NT	3.8	47 (J)	NT	43 (J)	-		NT	NT
	6002/01/71	LN LN	IN	Z L	130	IN	Z	NI TH	4.9	21	IN	49	+		IN	NT
	0102/11/0	2	N	IN IN	130	z	IN STORE	N Socoo	4,4	75	IZ	45	+		TN	IN
	2/2//2007	UN.	ON C	0.0/3 (B)	(5	Q	0.00013 (B)	0.000023 (B)	9.7	29	0.033	33	QN	0.013	0.00073 (B)	0.0023 (B)
	1002/02/0	ON.	UN ND	0.096 (B)	110	N	0.0012 (B)	0.0000071 (B)	5.4	44	0.0014 (B)	39	QN		0.00059 (B)	QN
CSMRI-11	1007/01/6	UN.	U.UU4 (B)	0.0/1 (B)	36	ON	0.00083 (B)	0.000016 (B)	4.5	39	0.0016 (B)	44	QN		0.00078(B)	0.0033 (B)
	11/26/2007			0.11	110	ON	Q	0.000028 (B)	4.9	44	0.0012 (B)	40	QN		0.0013 (B)	QN
	Z/15/2008	ON		0.12	110	ND			4.0	74		44	ON CIN			0,0048
	12/3/08 (DRV)	NT	TN	NT	NT	NT	TN	TN	NIT NIT	TIN TIN	TIN	11 LV	UN TH		UN L	IN
	3/16/09 (DRY)	NT	NT	NT	11	NT	NT	LN N	NT	NT	TIN	LIN	EN FIN		NIT	NT
CCMPL11B	1	0.00078	0.0032	0.22	89	0.00043	0,00041	0.000027	19	43	48	0.0019	0.00066		NT	NT L
		NT	NT	NŢ	130 (J)	NT	NT	ΤN	6.2	57 (J)	NT	49 (J)	NT	11.0	NT	NT
	12/15/2009	IN	NT	NT	NT	NT	IN	NT	NT	NT	NT	NT	NT	NT	NT	NT
at 1 and	3/8/2010	IN	IN	NT	IN,	IN	NT	TN	NT	NT	NT	NT	IN		NT	NT
LUBICOUN LIMIS	UIS	10.0	10.0	1.0	-	cnn'n	0.01	0.0002		-	0.01		0.003		0.01	0.02
			200	c	NE	0.005	14	N DAM	LIN	111	LIN		ALAN		1.1.	

not tested

(B) - Detected above Instrument Detection Level but below Reported D

ND - non detect NE - not established NT - not tested (B) - Detected above [Page G-4

Sample	Sample	Ra-226	Ra-228	Th-228	Th-230	Th-232	U-234	U-235	U-238	Total U
Station	Date	(pCi/l)	(l/brl)							
	2/25/2005	0	0.58	0.018	-0.026	-0.001	0.89	0.083	0.65	1.97
	6/14/2005	0.14	0.05	0.05	-0.025	0.016	0.246	0.021	0.251	0.75
	9/7/2005	0.18	0.42	0.041	0.25	0.102	0.35	0.031	0.35	1.04
	12/20/2005	-0.31	0.47	0.028	0.197	-0.005	0.64	0.041	0.7	2.11
	3/15/2006	-0.16	0.35	0.059	0.125	0.005	0.6	0.029	0.53	1.59
	6/14/2006	0.13	0.45	0.16	0.53	0.062	0.11	0.08	0.19	0.61
	9/13/2006	-0.03	0.25	-0.019	-0.035	0.01	0.37	-0.005	0.34	L
	3/1/2007	-0.1	0.25	-0.038	0.15	0.026	NT	NT	NT	1.7
	6/27/2007	0.13	0.77	0.006	0.016	0.014	NT	NT	NT	0.6
CINI 4	9/11/2007	0.15	0.74	0.063	0.088	0.012	NT	NT	NT	0.94
1-140	11/27/2007	0.2	0.24	0.026	0.049	0.025	NT	NT	NT	1.8
	2/27/2008	0.1	0.48	0.014	0.002	0.024	NT	NT	NT	2
	4/18/2008	0.06	-0.07	-0.023	-0.026	0.012	NT	NT	NT	1.9
	9/25/2008	0.18	-0.01	NT	NT	NT	NT	NT	NT	1.1
	12/3/2008	-0.06	0.34	NT	NT	NT	NT	NT	NT	1.6
	3/16/2009	0.14	0.73	NT	NT	NT	NT	NT	NT	1.9
	6/24/2009	0.33	1.228 J	NT	NT	NT	NT	NT	NT	0.55
	9/24/2009	-0.08	0.37	NT	NT	NT	NT	NT	NT	1.1
	12/17/2009	0.1	0.42	IN	NT	NT	NT	NT	NT	1.7
	3/9/2010	-0.04	0.2	NT	LN	NT	NT	NT	NT	2

Table G-3

Sample	Sample	Ra-226	Ra-228	Th-228	Th-230	Th-232	U-234	U-235	U-238	Total U
Station	Date	(pCi/l)	(pCi/l)	(pCi/l)	(pCi/l)	(pCi/l)	(pCi/l)	(pCi/l)	(pCi/l)	(l/brl)
	2/25/2005	0.45	0.06	0.011	-0.016	0.033	0.8	0.066	0.42	1.29
	6/14/2005	0.04	0.29	0.071	-0.028	0.007	0.259	0.032	0.23	0.69
	9/7/2005	-0.08	0.24	-0.013	0.107	0.051	0.54	0.014	0.54	1.62
	12/20/2005	0.09	0.07	-0.003	0.126	0	0.71	0.067	0.49	1.5
	3/15/2006	-0.04	-0.15	0.009	0.184	0.01	0.79	0.004	0.51	1.52
	6/14/2006	0.03	0.04	0.172	0.24	0.1	0.39	0	0.48	1.44
	9/13/2006	0.11	0.35	0.009	-0.03	0.01	0.43	-0.006	0.3	0.89
	3/8/2007	0.12	0.73	0.047	-0.055	0	NT	NT	NT	1.7
	6/28/2007	0.02	0.78	0.028	0.014	0	NT	NT	NT	0.57
0 1010	9/11/2007	0.1	0.27	0.066	0.068	0.002	NT	NT	NT	0.97
7-10	11/26/2007	0.11	0.36	0.007	0	0.012	NT	NT	NT	1.7
	2/26/2008	0.1	0	-0.01	0.113	0.011	NT	NT	NT	2
	4/18/2008	0.13	0.58	0.015	0.24	0.024	NT	NT	NT	1.8
	9/24/2008	-0.16	-0.02	NT	NT	NT	NT	NT	NT	0.99
	12/3/2008	0.1	0.46	NT	NT	NT	NT	NT	NT	1.5
	3/16/2009	0.2	0.29	NT	NT	NT	NT	NT	NT	1.9
	6/24/2009	0.03	0.47 J	NT	NT	NT	NT	NT	NT	0.059
	9/24/2009	0	0.28 (J)	IN	NT	NT	NT	NT	NT	1.1
	12/17/2009	0.03	0.44	IN	NT	NT	NT	NT	NT	1.9
	3/9/2010	-0.03	0.27	NT	NT	NT	NT	NT	NT	2
N.A.	+ 10+ 1	(}	1		- 000	1400 000 H 000 H	Lix	1	L	20

Table G-3

 MCL*
 Total Ra = 5
 NE
 Th 230 + Th 232 = 60**

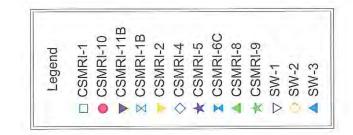
 *Maximum Contaminant Level – National Primary Drinking Water Regulations

**5 CCR 1002-31 Reg 31 - Colorado Surface Water Standards

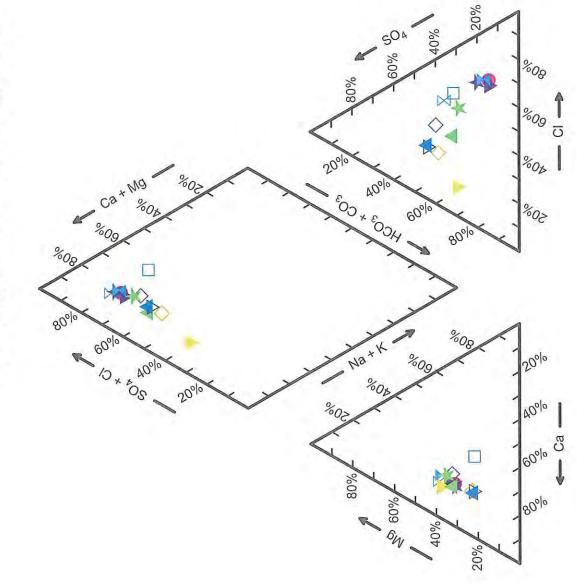
pCi/l - picoCuries per liter

μg/l - micrograms per liter

Appendix H Anion and Cation Balances and Piper Diagram







Water Type	Ca-Cl		
Dissolved Solids	360.97 mg/kg	360 mg/L	Measured
Density	0.9973 g/cm ³		Calculated
Conductivity	871 µmho/cm		Measured
Hardness (as CaCO	3)		
Total	189.63 mg/kg	189.12 mg/L	Calculated
Carbonate	136.52	136.15	
Non-Carbonate	53.112	52.968	
Primary Tests			
Anion-Cation Bala	nce		
Anions	5		
Cations	5.	71	
% Difference	6.	639	Not within $\pm 2\%$
Measured TDS = C	alculated TDS		
Measured		50.973	
Calculated	30	50.713	
Ratio		001	OK
Measured EC = Ca			
Measured	87	71.000	
Calculated		59.202	
Ratio	- L	558	Not within range 0.9 to 1.
Secondary Tests			
Measured EC and			
Anions	0.	574073	Not within preferred range
(0.9-1.1)			
Cations	0.	655723	Not within preferred range
(0.9-1.1)			
Calculated TDS to	EC ratio 0.	414	Not within preferred range
(0.55-0.7)			
Measured TDS to I	EC ratio 0.	414	Not within preferred range
(0.55-0.7)			
Organic Mass Bala			
DOC \geq Sum of Org		a.a.a.	
Dissolved Organi		000 mg/L	
Sum of Organics	0.	000 mg/L	OK

CSMRI-1B

Water Type	Ca-Cl		
Dissolved Solids	932.11 mg/kg	930 mg/L	Measured
Density	0.99773 g/cm ³		Calculated
Conductivity	1420 µmho/cm		Measured
Hardness (as CaCO	3)		
Total	776.28 mg/kg	774.52 mg/L	Calculated
Carbonate	378.14	377.28	
Non-Carbonate	398.14	397.24	
Primary Tests			
Anion-Cation Bala	nce		
Anions	15	.7	
Cations	18	.3	
% Difference	7.2	392	Not within $\pm 5\%$
Measured TDS = C	alculated TDS		
Measured	93	2.112	
Calculated	10	77.942	
Ratio		365	Not within range 1.0 to 1.2
Measured EC = Ca	red EC = Calculated EC		
Measured	14	20.000	
Calculated	15	59.433	
Ratio	0.9	011	OK
Secondary Tests			
Measured EC and I			
Anions	1.1	.08533	Not within preferred range
(0.9-1.1)			
Cations	1.2	285505	Not within preferred range
(0.9-1.1)			
Calculated TDS to	EC ratio 0.7	759	Not within preferred range
(0.55-0.7)			
Measured TDS to F		556	OK
Organic Mass Bala			
DOC ≥Sum of Org			
Dissolved Organi		000 mg/L	
Sum of Organics	0.0	000 mg/L	OK

Water Type	Ca-HCO ₃		
Dissolved Solids	511.32 mg/kg	510 mg/L	Measured
Density	0.99742 g/cm^3	STO INDER	Calculated
Conductivity	986 µmho/cm		Measured
Hardness (as CaCC			Measured
Total		400 2 //	Calculated
- A.M.	410.36 mg/kg	409.3 mg/L	Calculated
Carbonate Non-Carbonate	410.36 0.0	409.3 0.0	
ivon-Carbonate	0.0	0.0	
Primary Tests			
Anion-Cation Bala	nce		
Anions	6	.73	
Cations	9	.44	
% Difference	1	6.764	Not within $\pm 2\%$
Measured TDS = C	Calculated TDS		
Measured	5	11.321	
Calculated	6	23.511	
Ratio	C	.820	Not within range 1.0 to 1.2
Measured EC = Ca	lculated EC		the summer design of the
Measured	9	86.000	
Calculated	7	34.422	
Ratio	1	.343	Not within range 0.9 to 1.1
Secondary Tests			
Measured EC and			
Anions	0	.682736	Not within preferred range
(0.9-1.1)			
Cations	0	.957751	Within preferred range (0.9-
1.1)			
Calculated TDS to	EC ratio 0	.632	OK
Measured TDS to I	EC ratio 0	.519	Not within preferred range
(0.55-0.7)			
Organic Mass Bala			
DOC ≥Sum of Org			
Dissolved Organi		.000 mg/L	
Sum of Organics	0	.000 mg/L	OK

Water Type	Ca-SO ₄		
Dissolved Solids	821.93 mg/kg	820 mg/L	Measured
Density	0.99765 g/cm ³		Calculated
Conductivity	990 µmho/cm		Measured
Hardness (as CaCC			
Total	552.4 mg/kg	551.1 mg/L	Calculated
Carbonate	460.38	459.3	
Non-Carbonate	92.019	91.803	
Primary Tests			
Anion-Cation Bala	nce		
Anions		12	
Cations	1	[4	
% Difference		7.593	Not within $\pm 5\%$
Measured TDS = C	Calculated TDS		
Measured	8	321.931	
Calculated	9	012.142	
Ratio	(0.901	Not within range 1.0 to 1.2
Measured EC = Ca	lculated EC		
Measured	ç	990.000	
Calculated	1	215.773	
Ratio	C).814	Not within range 0.9 to 1.1
Secondary Tests			
Measured EC and			
Anions	1	.213595	Not within preferred range
(0.9-1.1)			
Cations	1	.413026	Not within preferred range
(0.9-1.1)			
Calculated TDS to	EC ratio ().921	Not within preferred range
(0.55-0.7)			
Measured TDS to I	EC ratio (0.830	Not within preferred range
(0.55-0.7)			
Organic Mass Bala			
DOC ≥Sum of Org		400	
Dissolved Organi		5.400 mg/L	OZ
Sum of Organics	C	0.000 mg/L	OK

Water Type	Ca-Cl		
Dissolved Solids	841.96 mg/kg	840 mg/L	Measured
Density	0.99767 g/cm^3	o to mg/n	Calculated
Conductivity	$1130 \mu\text{mho/cm}$		Measured
Hardness (as CaCO			Wedsured
Total	598.32 mg/kg	596.92 mg/L	Calculated
Carbonate	328.84	328.07	Calculated
Non-Carbonate	269.48	268.85	
rion carbonate	209.40	200.05	
Primary Tests			
Anion-Cation Bala			
Anions		2.6	
Cations		.4	
% Difference		600	Not within $\pm 5\%$
Measured $TDS = C$			
Measured		1.965	
Calculated		4.655	
Ratio		985	Not within range 1.0 to 1.2
Measured EC = Ca			
Measured		30.000	
Calculated		.79.460	
Ratio	0.8	883	Not within range 0.9 to 1.1
Secondary Tests			
Measured EC and			
Anions	1.	113615	Not within preferred range
(0.9-1.1)			
Cations	1.2	271006	Not within preferred range
(0.9-1.1)			
Calculated TDS to	EC ratio 0.7	756	Not within preferred range
(0.55-0.7)			
Measured TDS to E	EC ratio 0.3	745	Not within preferred range
(0.55-0.7)			
Organic Mass Bala			
DOC \geq Sum of Org			
Dissolved Organi		400 mg/L	
Sum of Organics	0.0	000 mg/L	OK

CSMRI-6C

Water Type	Ca-Cl		
Dissolved Solids	701.71 mg/kg	700 mg/L	Measured
Density	0.99756 g/cm ³		Calculated
Conductivity	990 µmho/cm		Measured
Hardness (as CaCO	(3)		
Total	506.51 mg/kg	505.28 mg/L	Calculated
Carbonate	361.76	360.88	
Non-Carbonate	144.75	144.4	
Primary Tests			
Anion-Cation Bala	nce		
Anions		10.3	
Cations		12.4	
% Difference	(9.027	Not within $\pm 5\%$
Measured TDS = C	alculated TDS		
Measured		701.712	
Calculated		759.052	
Ratio		0.924	Not within range 1.0 to 1.2
Measured EC = Ca	leulated EC		
Measured	9	990.000	
Calculated		1090.243	
Ratio	(0.908	OK
Secondary Tests			
Measured EC and			
Anions		1.041939	Within preferred range (0.9-
1.1)			
Cations)]	1.248705	Not within preferred range
(0.9-1.1)			
Calculated TDS to	EC ratio (0.767	Not within preferred range
(0.55-0.7)			
Measured TDS to E (0.55-0.7)	EC ratio ().709	Not within preferred range
Organic Mass Bala	nce		
DOC ≥Sum of Org			
Dissolved Organi		1.400 mg/L	
Sum of Organics		0.000 mg/L	OK

Water Type	Ca-HCO ₃		
Dissolved Solids	912.08 mg/kg	910 mg/L	Measured
Density	0.99772 g/cm ³		Calculated
Conductivity	1160 µmho/cm		Measured
Hardness (as CaCO) ₃)		
Total	648.08 mg/kg	646.6 mg/L	Calculated
Carbonate	575.43	574.12	
Non-Carbonate	72.645	72.479	
Primary Tests			
Anion-Cation Bala	nce		
Anions		3.1	
Cations	1	5.4	
% Difference	8	.020	Not within \pm 5%
Measured TDS = C	alculated TDS		
Measured	9	12.080	
Calculated	9	85.247	
Ratio	0	.926	Not within range 1.0 to 1.2
Measured EC = Ca	lculated EC		
Measured	1	160.000	
Calculated		293.482	
Ratio	0	.897	Not within range 0.9 to 1.1
Secondary Tests			
Measured EC and			
Anions	1	.131277	Not within preferred range
(0.9-1.1)			
Cations	1	.328556	Not within preferred range
(0.9-1.1)			
Calculated TDS to	EC ratio 0	.849	Not within preferred range
(0.55-0.7)	and the second		
Measured TDS to I (0.55-0.7)	EC ratio 0.	.786	Not within preferred range
Organic Mass Bala	nce		
DOC ≥Sum of Org			
Dissolved Organi		.700 mg/L	
Sum of Organics		.000 mg/L	OK

Water Type	Ca-Cl		
Dissolved Solids	922.1 mg/kg	920 mg/L	Measured
Density	0.99773 g/cm ³		Calculated
Conductivity	1380 µmho/cm		Measured
Hardness (as CaCC	0 ₃)		
Total	660.19 mg/kg	658.69 mg/L	Calculated
Carbonate	493.23	492.1	
Non-Carbonate	166.97	166.59	
Primary Tests			
Anion-Cation Bala	nce		
Anions	13.	2	
Cations	16.		
% Difference		640	Not within $\pm 5\%$
Measured TDS = C	alculated TDS		
Measured	922		
Calculated	996	5.165	
Ratio	0.9	26	Not within range 1.0 to 1.2
Measured EC = Ca			
Measured	138	0.000	
Calculated	135	4.380	
Ratio	1.0	19	OK
Secondary Tests			
Measured EC and			
Anions	s 0.956093		Within preferred range (0.9-
1.1)			
Cations	1.183782		Not within preferred range
(0.9-1.1)			
Calculated TDS to (0.55-0.7)	EC ratio 0.722		Not within preferred range
Measured TDS to I	C ratio 0.60	58	ОК
Organic Mass Bala			
DOC ≥Sum of Org			
Dissolved Organi		00 mg/L	
Sum of Organics		00 mg/L	OK

Water Terre	0.01		
Water Type Dissolved Solids	Ca-Cl	760	Measured
	761.82 mg/kg	760 mg/L	
Density	0.99761 g/cm ³		Calculated
Conductivity	1110 µmho/cm		Measured
Hardness (as CaCO			
Total	523.53 mg/kg	522.27 mg/L	Calculated
Carbonate	295.97	295.26	
Non-Carbonate	227.56	227.01	
Primary Tests			
Anion-Cation Bala	nce		
Anions	1	1	
Cations		2.7	
% Difference		923	Not within $\pm 5\%$
Measured $TDS = C$			
Measured		51.824	
Calculated		43.881	
Ratio		024	OK
Measured EC = Ca			
Measured		110.000	
Calculated	1	138.712	
Ratio	0.	975	OK
Secondary Tests			
Measured EC and I	fon Sums:		
Anions	0.	995104	Within preferred range (0.9-
1.1)			
Cations	1.	143142	Not within preferred range
(0.9-1.1)			
Calculated TDS to	EC ratio 0.	670	OK
Measured TDS to E	C ratio 0.	686	OK
Organic Mass Bala	nce		
DOC ≥Sum of Org			
Dissolved Organi		000 mg/L	
Sum of Organics		000 mg/L	OK

CSMRI-11B

Water Type	Ca-CI		
Dissolved Solids	721.75 mg/kg	720 mg/L	Measured
Density	0.99758 g/cm ³		Calculated
Conductivity	1000 µmho/cm		Measured
Hardness (as CaCO	(3)		
Total	552.44 mg/kg	551.1 mg/L	Calculated
Carbonate	361.75	360.88	
Non-Carbonate	190.69	190.22	
Primary Tests			
Anion-Cation Bala	nce		
Anions	11.	5	
Cations	13.		
% Difference	7.3		Not within $\pm 5\%$
Measured TDS = C			
Measured	721	.750	
Calculated	794	.927	
Ratio	0.9	08	Not within range 1.0 to 1.2
Measured EC = Ca	lculated EC		
Measured	100	0.000	
Calculated	117	3.475	
Ratio	0.8	52	Not within range 0.9 to 1.1
Secondary Tests			
Measured EC and I			
Anions	1.1.	46848	Not within preferred range
(0.9-1.1)			
Cations	1.3.	28795	Not within preferred range
(0.9-1.1)			
Calculated TDS to 1 (0.55-0.7)	EC ratio 0.79	95	Not within preferred range
Measured TDS to E (0.55-0.7)	C ratio 0.72	22	Not within preferred range
Organic Mass Bala	nce		
DOC ≥Sum of Org			
Dissolved Organi		00 mg/L	
Sum of Organics	0.00	00 mg/L	OK

SW-1

Water Type	Ca-SO ₄		
Dissolved Solids	61.179 mg/kg	61 mg/L	Measured
Density	0.99708 g/cm ³		Calculated
Conductivity	100 µmho/cm		Measured
Hardness (as CaCC) ₃)		
Total	28.633 mg/kg	28.549 mg/L	Calculated
Carbonate	24.677	24.605	
Non-Carbonate	3.9557	3.9441	
Primary Tests			
Anion-Cation Bala	nce		
Anions	57	1×10^{-3}	
Cations		4×10 ⁻³	
% Difference		.480	OK
Measured $TDS = C$	Calculated TDS		
Measured	61.	.179	
Calculated	45.	.002	
Ratio		59	Not within range 1.0 to 1.2
Measured EC = Ca	Iculated EC		designed and series of
Measured		0.000	
Calculated	72.	.736	
Ratio		75	Not within range 0.9 to 1.1
Secondary Tests			
Measured EC and			
Anions	0.5	70818	Not within preferred range
(0.9-1.1)			
Cations		04463	Not within preferred range
(0.9-1.1)	and all the second s	2.4	
Calculated TDS to EC ratio		50	Not within preferred range
(0.55-0.7)		1.2	
Measured TDS to EC ratio		12	OK
Organic Mass Bala			
DOC ≥Sum of Org		00 //	
Dissolved Organi		00 mg/L	OV
Sum of Organics	0.0	00 mg/L	OK

SW-2

Water Type	Ca-HCO ₃		
Dissolved Solids	61.179 mg/kg	61 mg/L	Measured
Density	0.99708 g/cm ³		Calculated
Conductivity	107 µmho/cm		Measured
Hardness (as CaCO	3)		
Total	27.882 mg/kg	27.8 mg/L	Calculated
Carbonate	24.677	24.605	
Non-Carbonate	3.2044	3.195	
Primary Tests			
Anion-Cation Balar	nce		
Anions	59.	4×10^{-3}	
Cations		6×10^{-3}	
% Difference	7.1	36	OK
Measured TDS = C	alculated TDS		
Measured	61.	179	
Calculated	43.	818	
Ratio		96	Not within range 1.0 to 1.2
Measured EC = Cal	lculated EC		
Measured	10	7.000	
Calculated	72.	278	
Ratio		80	Not within range 0.9 to 1.1
Secondary Tests			
Measured EC and I			
Anions		55359	Not within preferred range
(0.9-1.1)			
Cations		40715	Not within preferred range
(0.9-1.1)			
Calculated TDS to EC ratio		10	Not within preferred range
(0.55-0.7)			
Measured TDS to EC ratio		72	OK
Organic Mass Balan			
$DOC \ge Sum of Orga$		0.0	
Dissolved Organic		00 mg/L	~~~
Sum of Organics	0.0	00 mg/L	OK

	SW-3			
Water Type	Ca-SO ₄			
Dissolved Solids	61.179 mg/kg	61 mg/L	Measured	
Density	0.99708 g/cm ³		Calculated	
Conductivity	103 µmho/cm		Measured	
Hardness (as CaCO				
Total	28.883 mg/kg	28.799 mg/L	Calculated	
Carbonate	23.032	22.965		
Non-Carbonate	5.8513	5.8342		
Primary Tests				
Anion-Cation Bala	nce			
Anions	568×10-3			
Cations	706×10 ⁻³			
% Difference	10	.847	OK	
Measured TDS = C	alculated TDS			
Measured	61	.179		
Calculated	44	.129		
Ratio		86	Not within range 1.0 to 1.2	
Measured EC = Ca	lculated EC			
Measured		3.000		
Calculated		.810		
Ratio		15	Not within range 0.9 to 1,1	
Secondary Tests				
Measured EC and l				
Anions		51170	Not within preferred range	
(0.9-1.1)				
Cations		85292	Not within preferred range	
(0.9-1.1)				
Calculated TDS to EC ratio		28	Not within preferred range	
(0.55-0.7)				
Measured TDS to EC ratio		94	OK	
Organic Mass Bala				
DOC ≥Sum of Org		00 //		
Dissolved Organi		00 mg/L		
Sum of Organics	0.0	00 mg/L	OK	